

Comparative phytochemical and antioxidant profile of crude seed powder, aqueous and methanolic seed extracts of *Buchholzia coriacea*

Godwin C Ugwu^{*1}, Joseph Effiong Eyo¹, Jude Ifeanyi Egbuji¹, Chiemekam Ezechukwu¹, Keziah Nwamaka Ibemenuga²

*Corresponding author:

Godwin C Ugwu

¹University of Nigeria, Nsukka, NIGERIA

²Anambra State University, Department of Biological Sciences, Uli, Anambra State, NIGERIA.

Received: 18 May 2017

Accepted: 24 Jul 2017

Published: 28 Dec 2017

Abstract

The increasing discovery of more medicinal plants have triggered increased scientific screening of their bioactivity in order to provide data that will help physicians and patients make wise decision before using them. This study was designed to elucidate comparative phytochemical and antioxidant properties of crude seed powder, aqueous and methanolic seed extracts of *Buchholzia coriacea*. The results showed that crude seed powder had the highest alkaloid and phenol content of $3.98 \pm 0.00\%$ and $0.92 \pm 0.00\%$, while aqueous extract had the least alkaloid and phenol content of $1.00 \pm 0.00\%$ and $0.12 \pm 0.00\%$ respectively. Methanolic extracts had the highest phytochemical components among the three extracts with flavonoids (12.03 ± 0.0), saponins (1.99 ± 0.01), terpenoids (2.00 ± 0.00), tannin (0.10 ± 0.00) and phytate (2.02 ± 0.01) compositions, while aqueous extracts had the highest hydrogen cyanide (0.30 ± 0.00) and glycoside (0.35 ± 0.00). Antioxidant (DPPH) activities of *B. coriacea* showed that aqueous extract and crude seed powder had inhibition concentration (IC_{50}) of 4.65 mg/ml while methanolic extract had IC_{50} of 5.85 mg/ml. The result of the LD_{50} of the extracts showed the each extracts was well tolerated at a dose of 5000 mg/kg, an indication of high safety profile. The study therefore clearly demonstrated that methanolic extracts of *B. coriacea* have antioxidant, antihypertensive, hypocholesterolic and anticarcinogenic properties owing to the presence of high levels of phytochemical components than the aqueous and crude seed powder.

Keywords: Phytochemical, Crude seed powder, Aqueous and methanolic extracts, *Buchholzia coriacea*

Introduction

The increasing discovery of more medicinal plants have triggered increased scientific screening of their bioactivity in order to provide data that will help physicians and patients make wise decision before using them [1]. Approximately 119 pure chemical substances extracted from higher plants are used in medicine throughout the world [2]. It has been reported that more than 80 % of the population in developing countries depends on plants for their medical needs [3] and about 2/3 of all the medicinal plant species are found in the tropics [4].

However, *Buchholzia coriacea* popularly known as wonderful kola is one of the medicinal plants that have been used in different ways as an alternative medication to promote people health in Nigeria, Africa and other parts of the world. It is a forest tree with large, glossy, leathery and conspicuous leaves at the end of the branches [5]. The common name of the plant is "magic kola" while the

English name is "musk tree" [6]. In Nigeria, *B. coriacea* is known as "Uworo", "Owi" and "Uke" among Yoruba, Edo and Igbo tribes respectively [5]. Other common names include "Ovu" in Benin, and "Aponmu" in Akure [5]. Among central Africans, the fruit is known as "esson boss" [7]. The plant is widely distributed in most African countries, and its parts have been used to treat a variety of sickness in most rural communities in Nigeria. Some of the diseases locally treated with wonderful kola include fever, gonorrhoea and gastrointestinal infections [5], [8], [9]. Topical application of wonderful kola leaves and fruits on the body are used to relieve fever in Sierra Leone while the seeds are applied on the stomach to manage difficult child-birth in Abidjan [10]. The use of wonderful kola plant parts in some traditional medicine practices have been credited to some bioactive substances which they contain in rich amounts [5][8]. It is therefore the aim of this study to elucidate comparative phytochemical and antioxidant screening of

DOI:10.5138/09750185.2117



This article is distributed under the terms of the [Creative Commons Attribution License](https://creativecommons.org/licenses/by/4.0/), which permits unrestricted use and redistribution provided that the original author and source are credited.

crude seed powder, aqueous and methanolic seed extracts of *B.coriacea*.

Materials and methods

Procurement of plant material

The seeds of *B.coriacea* were purchased from local markets around Nsukka metropolis, Nsukka Local Government Area of Enugu State, Nigeria. The seeds were identified [11] and authenticated by Mr. A.O. Ozioko of Bioresources Development and Conservation Programme (BDCP), Nsukka, Enugu State, Nigeria. Voucher specimen number (INTERCEDD/ 709) was deposited in the BDCP herbarium. The seed was deshelled, cleaned and cut into small sizes, air dried at room temperature until a constant weight was obtained (7 days) and pulverized into fine powder using a grinding machine (Honda: model 622, China).

Preparation of extracts

The method of extraction followed that of Okere *et al.* [12]. Two hundred gram (200 g) and 800 g of the powdered seeds were soaked in 500 ml of water and 700 ml of methanol respectively for 48 hours with intermittent shaking. The soaked substance were filtered using Whatman No. 1 filter paper and concentrated using rotary evaporator. The concentrated sample were further subjected to dryness using hot air oven set at 60 °C and then stored in an airtight bottle in a refrigerator for subsequent use. The crude powder was gotten by sieving the ground powder samples through muslin cloth (0.7 mm mesh size). The aqueous and methanolic extracts gave 19.18 and 5.17 % yield respectively.

Phytochemical analysis

Phytochemical analysis for the qualitative detection of biologically active compounds such as saponins, alkaloids, tannins, flavonoids, steroids, terpenoids, glycosides, cyanide and phytate on the crude powder, aqueous and methanolic seed extracts of *B. coriacea* were determined using the methods of Sofowara [13], Trease and Evans [14] and Harborne [15], while quantitative was determined using the methods of Pearson [16], Obadoni and Ochuko [17], Onwuka [18], Subhadhirasakul and Pechpongs [19], El-Olemy *et al.* [20] and Oberleas [21].

Determination antioxidant potential of crude powder, aqueous and methanolic seed extracts of *B. coriacea*

In order to evaluate the antioxidant potential through free radical scavenging by the test samples, the change in optical density of 2, 2-diphenyl-1-picrylhydrazyl (DPPH) radicals was monitored. This was determined using the method of Manzocco *et al.* [22]. The

sample of each extract (0.2 ml) is diluted with methanol and 2 ml of DPPH solution (0.5 mM) is added. After 30 minutes, the absorbance is measured at 517 nm. The percentage of the DPPH radical scavenging is calculated using the equation:

$$\% \text{ inhibition of DPPH radical} = [(A_{br} - A_{ar}) / A_{br}] \times 100,$$

where: A_{br} is the absorbance before reaction and A_{ar} is the absorbance after reaction has taken place.

Acute toxicity study

The lethal dose (LD₅₀) of the crude powder, aqueous and methanolic extracts was determined according to the method of Lorke [23]. A total of sixty three (63) mice were used for each for the study in two phases. For the first phase, twelve (12) mice were divided into 4 groups (A, B, C and D) of 3 mice per group, and treated with each extracts at the doses of 100, 200, 400 and 800 mg/kg respectively. The animals were observed for 2 hours, intermittently for 6 hours and over a period of 24 hours for signs of toxicity. For the second phase, nine (9) mice were divided into 3 groups (A, B and C) of 3 mice per group and treated with each extracts at the doses of 1000, 3000 and 5000 mg/kg respectively. They were observed for signs of toxicity for 24 hours. The numbers of animals dead within 24 hours after the administration were recorded for each group. The lethal dose was calculated as the arithmetic mean of the dose that killed the least number of animals and the one next to lower dose that did not kill any animal.

Statistical Analysis

Results obtained were presented as mean \pm S.E.M and statistical evaluation of the quantitative phytochemicals of different extracts used was done using one-way analysis of variance (ANOVA) while Duncan's Multiple Range Test (DMRT) was used to test the significant differences among the extracts and level of significance was set at $p < 0.05$ [24].

Results

Qualitative and quantitative phytochemical composition of the crude seed powder, aqueous and methanolic seed extract of *B. coriacea*

Table 1 and 2 showed qualitative and quantitative phytochemical composition of aqueous, methanolic and crude seed powder of *Buchholzia coriacea*. Qualitatively, it was observed that crude seed powder had the highest alkaloid content followed by similar alkaloid contents in methanolic aqueous extracts. Flavonoids were high and similar in all extracts. Furthermore, saponins, terpenoids, tannins, phenols and phytate were higher in methanolic than in crude powder and aqueous extracts. Steroids were absent in all the extracts. Hydrogen cyanide was present only in the aqueous



extract that also had higher level of cardiac glycoside than methanolic extract and crude powder of *B. coriacea* (Table 1).

Quantitatively, it was observed that crude seed powder had the highest alkaloid content followed by methanolic and aqueous extracts. The methanolic extract had significantly the highest flavonoid content, while the crude seed powder had the least flavonoid content, although, all the extracts had high concentrations of flavonoid (Table 2). For saponin, all the extracts were significantly different from each other, with aqueous extract having the least value, while the methanolic extract had the highest value (Table 2). The terpenoid content of methanolic extract was significantly higher ($P < 0.05$) than the aqueous and crude powder extracts which had the similar terpenoid content. There was no significant difference ($P > 0.05$) between the tannin content of aqueous and methanolic extracts of *B. coriacea*, but were

significantly higher ($P < 0.05$) than that of crude powder (Table 2). The phenolic content of crude seed powder of *B. coriacea* was highest while the aqueous extract was lower and the phenolic content of the three extract are significantly difference ($P > 0.05$) from each other (Table 2). Methanolic extract has the highest phytate content while the aqueous extract had the lowest and they were significantly different ($P < 0.05$) from each other. There were no significant difference ($P > 0.05$) among steroid content of the three extract, although the levels were very low. The aqueous extracts had the highest cyanide content, while the methanolic extract had the least cyanide content, and were significantly different from one another. Similarly, the glycoside content of aqueous extracts was the highest while that of crude seed powder was the lowest and they were significantly different from one another (Table 2).

Table-1: Qualitative phytochemical screening of the crude seed powder, aqueous and methanolic seed extract of *B. coriacea*

S.No	Phytochemicals	Aqueous	Methanol	Crude powder
1	Alkaloids	++	++	+++
2	Flavonoids	+++	+++	+++
3	Saponins	+	+++	++
4	Terpenoids	+	+++	+
5	Tannin	+	++	+
6	Phenol	+	++	++
7	Phytate	+	+++	++
8	Steroids			
9	Hydrogen cyanide	+	-	-
10	Cardiac glycoside	++	+	+

+++ = High concentration, ++ = Moderate concentration, + = Trace concentration, -= Absent

Table-2: Quantitative phytochemical screening of the crude seed powder, aqueous and methanolic seed extract of *B. coriacea*

S. No	Phytochemicals	Composition (%)		
		Aqueous Extracts	Methanolic Extracts	Crude powder
1	Alkaloids	1.00±0.00 ^a	1.49±0.01 ^b	3.98±0.01 ^c
2	Flavonoids	8.03±0.01 ^c	12.03±0.01 ^b	6.10±0.06 ^a
3	Saponins	0.50±0.00 ^a	1.99±0.01 ^c	0.99±0.01 ^b
4	Terpenoids	0.50±0.00 ^a	2.00±0.00 ^b	0.50±0.00 ^a
5	Tannin	0.09±0.00 ^a	0.10±0.00 ^a	0.02±0.00 ^b
6	Phenol	0.12±0.00 ^a	0.68±0.00 ^b	0.92±0.00 ^c
7	Phytate	0.26±0.00 ^a	2.02±0.01 ^c	0.75±0.00 ^b
8	Steroids	0.02±0.00 ^a	0.01±0.00 ^a	0.02±0.00 ^a
9	Hydrogen cyanide	0.30±0.00 ^c	0.06±0.00 ^a	0.08±0.00 ^b
10	Glycoside	0.35±0.00 ^c	0.11±0.01 ^a	0.14±0.00 ^b

Values represent mean±SEM of 3 observations. Values with the same letter superscript on the same row are not significantly different ($P > 0.05$), Values with different letter superscript on the same row are significantly different ($P < 0.05$).

Antioxidant activity of crude seed powder, aqueous and methanolic seed extract of *B. coriacea*

The result of the antioxidant activities of crude seed powder, aqueous and methanolic seed extracts of *B. coriacea* indicated that the aqueous extracts and crude seed extracts of *B. coriacea* had similar inhibition concentration (IC_{50}) of 4.65 mg/ml that were lower than that of methanolic extracts (5.85 mg/ml).

Table-3: Antioxidant activity of crude seed powder, aqueous and methanolic seed extract of *B. coriacea*

S. No	Conc. (mg/ml)	DPPH radical scavenging activities (%)		
		BCAE	BCME	BCCP
1	1.00	14.38	10.59	23.14
2	2.00	26.61	13.11	29.15
3	3.00	33.92	24.21	39.63
4	4.00	37.71	24.97	45.20
5	5.00	42.37	36.74	47.38
IC ₅₀ (mg/ml)		4.65	5.85	4.65

BCAE=*Buchholzia coriacea* aqueous extract, BCME=*Buchholzia coriacea* methanolic extract, BCCP=*Buchholzia coriacea* crude powder

Acute toxicity studies of crude seed powder, aqueous and methanolic seed extract of *B. coriacea*

There was no mortality at different doses (100, 200, 400 and 800 mg/kg) of aqueous extract, methanol extract and crude seed

powder of *B. coriacea* after 24 hours in phase one (Table 4). Also, at dose levels of 1,000 mg/kg, 3,000 mg/kg and 5,000 mg/kg, there was no death recorded, although, cage side behaviours such as shivering, bulging of eyes, sedation, depression and dullness were observed in the animals.

Table-4: Acute toxicity of crude seed powder, aqueous and methanolic seed extract of *B. coriacea*

Doses (mg/kg)	Mortality recorded		
	Aqueous extract	Methanolic extract	Crude seed powder
Phase 1			
100	0/3	0/3	0/3
200	0/3	0/3	0/3
400	0/3	0/3	0/3
800	0/3	0/3	0/3
Phase 2			
1000	0/3	0/3	0/3
3000	0/3	0/3	0/3
5000	0/3	0/3	0/3

Discussion

Phytochemicals

The curative properties of plant derived medicine are due to the presence of bioactive components of varied composition in one or more parts of the plants. The phytochemical screening of aqueous extract, methanol extract and crude seed powder of *B. coriacea* indicated the presence of some phyto-compounds such as alkaloids, tannins, flavonoids, steroids, saponins, glycosides, phytate, cyanide and terpenes. These are the most important bioactive constituents of plant [25]. Alkaloids, as a component, have therapeutic activity as it is a primary natural phyto-component of plants. The *B. coriacea* extracts had moderate concentration of alkaloids indicated that it may be used as sedative, pain-relieving drug, anaesthetic, analgesic etc. [26][27]. Alkaloids also have medical, pharmacological and veterinary importance. Alkaloids belonging to beta-carboline group have been reported to possess antimicrobial, anti-HIV and antiparasitic activities [28]. Alkaloids have a wide range of pharmacological activities including antimalarial (e.g. quinine), anticancer (e.g. homoharringtonine), antiasthma (e.g. ephedrine)[29], cholinomimetic e.g. galantamine

[30], antiarrhythmic (e.g. quinidine), vasodilatory (e.g. vincamine), analgesic (e.g. morphine) [31], antibacterial (e.g. chelerythrine) [32] and antihyperglycemic (e.g. piperine) [33]. Many have found use in traditional or modern medicine, or as starting points for drug discovery. Other alkaloids possess psychotropic (e.g. psilocin) and stimulant activities (e.g. theobromine, cocaine, nicotine, caffeine) and have been used as recreational drugs [34].

Alkaloids can also be toxic (e.g. tubocurarine, atropine) [35]. Although alkaloids act on a diversity of metabolic systems in humans and other animals, they almost uniformly evoke a bitter taste [36] and in some cases may cause serious illness, injury or even death. The manner of poisoning with plants alkaloids can be divided into unintentional ingestion of plant material, intentional ingestion of plant material, and ingestion of abused plant material [37].

The flavonoid contents were in abundance in all the extracts. The presence of flavonoids in the extract showed that the plant have antioxidant properties; this could enhance the body's defense systems against pathologically induced free-radical generation as well as modify the body's reactions to allergens and viruses [38]. Flavonoids have been demonstrated to have a wide range of biological and pharmacological functions in *in-vitro*



studies. These include anti-inflammatory [39][40], anti-allergic [39], antioxidant [38][40], anti-microbial [41][42][43][44], anti-cancer [40][45] and anti-diarrheal activities [46]. Flavonoids have also been shown to inhibit topoisomerase enzymes [47][48] and to induce DNA mutations in the mixed-lineage leukaemia gene in *in-vitro* studies [49]. However, for most of the cases cited above, no follow up *in-vivo* or clinical research has been carried out thus leaving it impossible to affirm if these activities have any detrimental or beneficial effect on human health.

Saponins and phenols in food medicine and mastic ants contribute to the low rate of atherosclerosis and coronary heart disease [50]. Similarly, saponins from soybean have been shown to have hypocholesterolemic as well as anticarcinogenic effects [51]. Saponins consumption often times causes deleterious effects such as haemolysis and permeability of the intestines [52]. Thus, the methanolic extract and crude seed powder of *B. coriacea* that had abundant and moderate amount of saponins indicated the possibility of using these extracts in the management of cardiovascular diseases [50].

Tannins have biological properties that may favour the prevention and management of many ailments [53]. However, tannins may decrease protein quality by reducing palatability and digestibility; they have good antimicrobial and anti-inflammatory activities and thus are used as alternative to antimicrobial growth promoter factors in poultry [54]. Excess tannins may be toxic because tannins as metal ion chelators can decrease the bioavailability of iron which often leads to anaemia [55]. Therefore, the presence of tannin in the plant extract showed that the extract especially the methanolic extracts that had moderate amount may play a very important therapeutic role in the field of medicine. Thus, *B. coriacea* could be used in the management of viral, bacterial and fungal diseases.

Phytic acid has been reported to lower the nutritional value due to its limiting effects on the bioavailability of dietary minerals and essential trace elements (e.g. iron, zinc, calcium) in human intestine [56]. They equally possess antioxidant, anticarcinogenic and hypoglycaemic properties [57]. The high and moderate levels of phytate in the methanolic extracts and crude seed powder of *B. coriacea* showed that it could be used in the treatment of cancer and diabetes mellitus.

Steroids to some extent maintain hormonal balance by serving as a precursor or potent material in the synthesis of sex hormone [58]. Thus, the aqueous, methanolic extracts and crude seed powder of *B. coriacea* having trace amount of steroids is an indication that the plant could not be used in management of hormonal imbalance. More so, the aqueous seed extract *B. coriacea* contains very low cyanide content while cyanide was absent in methanolic extract and crude seed powder, which means that *B. coriacea* seed is not poisonous, and could serve as food or used as a herbal medicine.

Terpenoids and phenols have antioxidant properties and protect cells from oxidative damage [59]. Therefore, the presence of both terpenoids and phenols in all the extracts especially the methanolic

extracts that have reasonable quantity of terpenoids, *B. coriacea* seed extracts could be used as an antioxidant in the management of oxidative damage.

Cardiac glycosides are found as secondary metabolites in several plants and are used in the treatment of congestive heart failure and cardiac arrhythmia. Furthermore, several cardiac glycosides such as peruvoside have been used in cancer control, especially ovary cancer and leukaemia [60]. Aqueous extract of *B. coriacea* has moderate amount of cardiac glycoside while methanolic and crude seed powder have trace quantity. Thus, the plant could be used in the management of heart failure, cancer and leukaemia.

Antioxidant activities

Natural antioxidants that are present in plants are responsible for inhibiting or preventing the deleterious consequences of oxidative stress. In the present study, we have evaluated the free radical scavenging activities of crude seed powder, aqueous and methanolic extracts of *B. coriacea* seed. However, among the three extracts tested for *in-vitro* antioxidant activity using the DPPH method, it was observed that aqueous extracts and crude seed powder of *B. coriacea* had equal and greater inhibition concentration (IC_{50}) of 4.65 mg/ml than that of methanolic extract which has 5.85 mg/ml. It is obvious that the components like tannins, saponins and other phyto-constituents of the extracts may be responsible for such activity. Several of such compounds are known to possess potent antioxidant activity [61]. Antioxidant properties of medicinal plants have been documented by several researchers [22], [38], [59], [61]. The present research however, corroborates that of Nwaehujor *et al.* [62] on the antioxidant properties of *B. coriacea* fruits. The study has therefore clearly demonstrated that methanolic extracts could have antioxidant, antihypertensive, hypocholesterolemic and anticarcinogenic effects thus; high medicinal properties owing to the presence of higher phytochemical components and antioxidant activities than the aqueous and crude seed powder.

Acute toxicity

The results of the toxicity study showed that all the *B. coriacea* extract used for the research had no lethal effect on the tested experimental animals, since no death was recorded at the dose of 5000 mg/kg, thus the plant has a wide safety margin in mice and may be safe for human consumption. The finding of this study is similar to earlier reports that no death was recorded in the acute toxicity study carried out with *B. coriacea* seed extract [62][63][64]. Also, this work also agrees with Adisa *et al.* [6] who observed that no death was recorded after the toxicity study and therefore concluded that LD_{50} of *Buchholzia coriacea* is greater than 30,000 mg/kg body weight.

Conclusion



Buchholzia coriacea extracts are rich sources of phytochemicals such as alkaloids, flavonoids, saponins, terpenoids, tannin, phenol, phytate, steroids, hydrogen cyanide and glycoside. The crude seed powder had the highest alkaloid content followed by methanolic and aqueous extracts. The methanolic extract had the highest flavonoid content, while the crude seed powder had the least flavonoid content, although, all the extracts had high concentrations of flavonoid. The extracts have antioxidant properties and as such can be used in combating or preventing the deleterious consequences of oxidative stress in animals exposed to it. The extracts were non-toxic as such the seed can be used as food and as medicine. *B. coriacea* extracts have been demonstrated to have a wide range of biological and pharmacological activities thus: anti-

inflammatory, anti-allergic, antioxidant, anti-microbial, anti-cancer, anti-diarrheal, anti-leukaemia, anti-topoisomerase, induction of DNA mutations, contribute to the low rate of atherosclerosis and coronary heart disease, are hypocholesterolemic as well as anticarcinogenic effects, used in the management of cardiovascular diseases among many others. These are the reasons why it is nicknamed "wonderful kola".

Conflicts of Interests

Non to declare

References

- [1]. Oyewole OI, Akigbala PF. Phytochemical analysis and hypolipidemic properties of *Jatropha tanjorensis* leaf extract. *European Journal of Medicinal Plants*. 2011; 1(4): 180-85.
- [2]. Hoareau L, DaSilva EJ. Medicinal plants: a re-emerging health aid. *Electronic Journal of Biotechnology*. 1999; 2(2):3-4.
- [3]. Focho DA, Ndam WT, Fonge BA. Medicinal plants of Aguambu-Bamumbu in the Lebialem highlands, southwest province of Cameroon. *African Journal of Pharmacy and Pharmacology*. 2009; 3(1):001-13.
- [4]. FAO. Some medicinal forest plants of Africa and Latin America. FAO Forestry Paper. 1986; 67:1-274.
- [5]. Mbata TI, Duru CM, Onwumelu HA. Antibacterial activity of crude seed extracts of *Buchholzia coriacea* E. on some pathogenic bacteria. *Journal of Developmental Biology and Tissue Engineering*. 2009; 1(1):001-5.
- [6]. Adisa RA, Choudhary MI, Olorunsogo OO. Hypoglycemic activity of *Buchholziacoriacea* (Capparaceae) seeds in streptozotocin-induced diabetic rats and mice. *Experimental and Toxicologic Pathology*. 2011; 63(7):619-25.
- [7]. Ibrahim TA, Fagbohun ED. Phytochemical and nutritive quality of dried seeds of *Buchholzia Coriacea*. *Greener Journal of Physical Sciences*. 2012; 2(5):185-91.
- [8]. Oluseyi EO, Onyeoziri NF. Preliminary studies on the antimicrobial properties of *Buchholzia coriacea* (wonderful kola). *African Journal of Biotechnology*. 2009; 8(3):472-4.
- [9]. Ezeja MI, Ezeigbo II, Madubuike KG. Analgesic activity of the methanolic seed extract of *Buchholzia coriacea*. *Research Journal of Pharmaceutical, Biological and Chemical Sciences*. 2011; 2(1):187-93.
- [10]. Adjounahum E, Ake-Assi J. Plantes pharmaceutiques de Côté d'Ivoire. Abidjan. 1972:117-118.
- [11]. Keay RW, Onochie CF, Stanfield DP. Nigerian trees. Federal Department of Forest Research. Ibadan; Nigeria. Printed at the Nigerian National Press Limited. 1964.
- [12]. Okere OS, Iliemene UD, Tese T, Mubarak L, Olowoniyi OD. Proximate analysis, phytochemical screening and antitrypanocidal potentials of *Bucholzia coriacea* in *Trypanosoma brucei brucei* infected mice. *IOSR Journal of Pharmacy and Biological Sciences*. 2014; 9(4):69-77.
- [13]. Sofowara A. Medicinal plants and traditional medicine in Africa. Ibadan, Nigeria: Spectrum Books Limited. 1993.
- [14]. Trease GE, Evans WC. Pharmacognosy. 11th Edition. London: Bailliere Tindall. 1989.
- [15]. Harborne AJ. Phytochemical methods a guide to modern techniques of plant analysis. Netherlands: Springer Science and Business Media. 1998.
- [16]. Pearson DA. The chemical analysis of foods. 7th Edition. Edinburgh, London: Churchill and Livingstone. 1976.
- [17]. Obadoni BO, Ochuko PO. Phytochemical studies and comparative efficacy of the crude extracts of some haemostatic plants in Edo and Delta States of Nigeria. *Global Journal of Pure and Applied Sciences*. 2002; 8(2):203-8.
- [18]. Onwuka GI. Food analysis and instrumentation theory and practice. Lagos, Nigeria: Naphtali Prints; 2005.
- [19]. Subhadhirasakul S, Pechpongs P. A terpenoid and two steroids from the flowers of *Mammea siamensis*. *Songklanakarin Journal of Science and Technology*. 2005; 27(2):555-61.
- [20]. El-Olemy MM, Al-Muhtadi FJ, Afifi AF. Experimental phytochemistry: A



- laboratory manual. Saudi Arabia: King Saud University Press. 1994.
- [21]. Oberleas D. Phytates in toxicants occurring naturally in foods. Washington DC, USA: National Academy of Sciences; 1973.
- [22]. Manzocco L, Anese M, Nicoli MC. Antioxidant properties of tea extracts as affected by processing. *LWT-Food Science and Technology*. 1998; 31(7):694-8.
- [23]. Lorke D. A new approach to practical acute toxicity testing. *Archives of Toxicology*. 1983; 54(4):275-87.
- [24]. Duncan DB. Multiple range and multiple F tests. *Biometrics*. 1955; 11(1):1-42.
- [25]. Sasidharan S, Chen Y, Saravanan D, Sundram KM, Yoga Latha L. Extraction, isolation and characterization of bioactive compounds from plants' extracts. *African Journal of Traditional, Complementary, and Alternative Medicines*. 2011; 8(1):1-10.
- [26]. Osadebe PO, Uzochukwu IC. Chromogenic and antimotility studies on extract of *Loranthus microanthus* Linn. *Journal of Pharmacy and Allied Sciences*. 2006; 3(1):263-8.
- [27]. Malu SP, Obochi GO, Edem CA, Nyong BE. Effect of methods of extraction on phytochemical constituents and antibacterial properties of *Tetracarpidium conophorum* seeds. *Global Journal of Pure and Applied Sciences*. 2009; 16(3-4):373-6.
- [28]. Bouayad N, Rharrabe K, Lamhamdi M, Nourouti NG, Sayah F. Dietary effects of harmine, a β -carboline alkaloid, on development, energy reserves and a amylase activity of *Plodia interpunctella* Hübner [Lepidoptera: Pyralidae]. *Saudi Journal of Biological Sciences*. 2011; 19(1):73-80.
- [29]. Kittakoop P, Mahidol C, Ruchirawat S. Alkaloids as important scaffolds in therapeutic drugs for the treatments of cancer, tuberculosis, and smoking cessation. *Current Topics in Medicinal Chemistry*. 2014; 14(2):239-52.
- [30]. Russo P, Frustaci A, Del Bufalo A, Fini M, Cesario A. Multitarget drugs of plants origin acting on Alzheimer's disease. *Current Medicinal Chemistry*. 2013; 20(13):1686-93.
- [31]. Sinatra RS, Jahr JS, Watkins-Pitchford JM. The essence of analgesia and analgesics. Cambridge University Press; 2010.
- [32]. Cushnie TT, Cushnie B, Lamb AJ. Alkaloids: an overview of their antibacterial, antibiotic-enhancing and antivirulence activities. *International Journal of Antimicrobial Agents*. 2014; 44(5):377-86.
- [33]. Shi QI, Hui SU, ZHANG AH, Hong-Ying XU, Guang-Li YA, Ying HA, Xi-Jun WA. Natural alkaloids: basic aspects, biological roles, and future perspectives. *Chinese Journal of Natural Medicines*. 2014; 12(6):401-6.
- [34]. Wink M. Modes of action of herbal medicines and plant secondary metabolites. *Medicines*. 2015; 2(3):251-86.
- [35]. Robbers JE, Speedie MK, Tyler VE. Pharmacognosy and pharmacobiotechnology. Philadelphia: Lippincott, Williams and Wilkins; 1996.
- [36]. Rhoades DF. Evolution of plant chemical defense against herbivores. *Herbivores: their interaction with secondary plant metabolites*. New York: Academic Press; 1979:3-54.
- [37]. Beyer J, Drummer OH, Maurer HH. Analysis of toxic alkaloids in body samples. *Forensic Science International*. 2009; 185(1-3):1-9.
- [38]. Al-Humaid AI, Mousa HM, El-Mergawi RA, Abdel-Salam AM. Chemical composition and antioxidant activity of dates and dates-camel-milk mixtures as a protective meal against lipid peroxidation in rats. *American Journal of Food Technology*. 2010; 5(1):22-30.
- [39]. Yamamoto Y, Gaynor RB. Therapeutic potential of inhibition of the NF- κ B pathway in the treatment of inflammation and cancer. *The Journal of Clinical Investigation*. 2001; 107(2):135-42.
- [40]. Cazarolli LH, Zanatta L, Alberton EH, Figueiredo B, Reis MS, Follador P, Damazio RG, Pizzolatti MG, Silva B, Mena FR. Flavonoids: prospective drug candidates. *Mini Reviews in Medicinal Chemistry*. 2008; 8(13):1429-40.
- [41]. Cushnie TT, Lamb AJ. Recent advances in understanding the antibacterial properties of flavonoids. *International Journal of Antimicrobial Agents*. 2011; 38(2):99-107.
- [42]. Manner S, Skogman M, Goeres D, Vuorela P, Fallarero A. Systematic exploration of natural and synthetic flavonoids for the inhibition of *Staphylococcus aureus* biofilms. *International Journal of Molecular Sciences*. 2013; 14(10):19434-51.
- [43]. Mandalari G, Bennett RN, Bisignano G, Trombetta D, Saija A, Faulds CB, Gasson MJ, Narbad A. Antimicrobial activity of flavonoids extracted from bergamot (*Citrus bergamia* Risso) peel, a by-product of the essential oil industry. *Journal of Applied Microbiology*. 2007; 103(6):2056-64.
- [44]. Friedman M. Overview of antibacterial, antitoxin, antiviral, and antifungal activities of tea flavonoids and teas. *Molecular Nutrition and Food Research*. 2007; 51(1):116-34.
- [45]. Ruela de Sousa RR, Queiroz KC, Souza AC, Gurgueira SA, Augusto AC, Miranda MA, Peppelenbosch MP, Ferreira CV, Aoyama H. Phosphoprotein levels, MAPK activities and NF κ B expression are affected by fisetin. *Journal of Medicinal Chemistry*. 2007; 22(4):439-44.
- [46]. Schuier M, Sies H, Illek B, Fischer H. Cocoa-related flavonoids inhibit CFTR-mediated chloride transport across T84 human colon epithelia. *The Journal of Nutrition*. 2005; 135(10):2320-5.
- [47]. Esselen M, Fritz J, Hutter M, Marko D. Delphinidin modulates the DNA-damaging properties of

- topoisomerase II poisons. *Chemical Research in Toxicology*. 2009; 22(3):554-64.
- [48]. Bandle OJ, Clawson SJ, Osheroff N. Dietary polyphenols as topoisomerase II poisons: B ring and C ring substituents determine the mechanism of enzyme-mediated DNA cleavage enhancement. *Chemical Research in Toxicology*. 2008; 21(6):1253-60.
- [49]. Van Doorn SB, Janssen J, Maas LM, Godschalk RW, Nijhuis JG, van Schooten FJ. Dietary flavonoids induce MLL translocations in primary human CD34+ cells. *Carcinogenesis*. 2007; 28(8):1703-9.
- [50]. Johns T. Phytochemicals as evolutionary mediators of human nutritional physiology. *International Journal of Pharmacognosy*. 1996; 34(5):327-34.
- [51]. Oh YJ, Sung MK. Soybean saponins inhibit cell proliferation by suppressing PKC activation and induce differentiation of HT-29 human colon adenocarcinoma cells. *Nutrition and Cancer*. 2001; 39(1):132-8.
- [52]. Francis G, Kerem Z, Makkar HP, Becker K. The biological action of saponins in animal systems: a review. *British Journal of Nutrition*. 2002; 88(06):587-605.
- [53]. James DB, Abu EA, Wurochekke AU, Orji GN. Phytochemical and antimicrobial investigation of the aqueous and methanolic extracts of *Ximenia americana*. *Journal of Medical Sciences, Peshawar*. 2007; 7(2):284-8.
- [54]. Redondo LM, Chacana PA, Dominguez JE, Fernandez Miyakawa ME. Perspectives in the use of tannins as alternative to antimicrobial growth promoter factors in poultry. *Frontiers in Microbiology*. 2014;5:118. doi:10.3389/fmicb.2014.00118.
- [55]. Ukoha PO, Cemaluk EA, Nnamdi OL, Madus EP. Tannins and other phytochemical of the *Samanaea saman* pods and their antimicrobial activities. *African Journal of Pure and Applied Chemistry*. 2011; 5(8):237-44.
- [56]. Abbaspour N, Hurrell R, Kelishadi R. Review on iron and its importance for human health. *Journal of Research in Medical Sciences*. 2014; 19(2):164-74.
- [57]. Kumar V, Sinha AK, Makkar HP, Becker K. Dietary roles of phytate and phytase in human nutrition: A review. *Food Chemistry*. 2010; 120(4):945-59.
- [58]. Hu J, Zhang Z, Shen W-J, Azhar S. Cellular cholesterol delivery, intracellular processing and utilization for biosynthesis of steroid hormones. *Nutrition and Metabolism*. 2010; 7:47. doi: 10.1186/1743-7075-7-47.
- [59]. Kasote DM, Katyare SS, Hegde MV, Bae H. Significance of antioxidant potential of plants and its relevance to therapeutic applications. *International Journal of Biological Sciences*. 2015; 11(8):982-91.
- [60]. Patel S. Plant-derived cardiac glycosides: role in heart ailments and cancer management. *Biomedicine and Pharmacotherapy*. 2016; 84:1036-41.
- [61]. Cai Y, Luo Q, Sun M, Corke H. Antioxidant activity and phenolic compounds of 112 traditional Chinese medicinal plants associated with anticancer. *Life Sciences*. 2004; 74(17):2157-84.
- [62]. Nwaehujor CO, Ode OJ, Nwinyi FC, Udeh NE. Effects of methanol extract of *Buchholzia coriacea* fruit in streptozotocin-induced diabetic rats. *Journal of Pharmacology and Toxicology*. 2012; 7(4):181-91.
- [63]. Nweze NE, Fakae NE, Asuzu IU. Trypanocidal activity of the ethanolic extract of *Buchholzia coriacea* seed. *Nigerian Veterinary Journal*. 2008; 29(4):1-6.
- [64]. Okoye TC, Akah PA, Ilogu CL, Ezike AC, Onyeto CA. Anti-diabetic Effects of Methanol Extract of the Seeds of *Buchholzia coriacea* and its Synergistic Effects with Metformin. *Asian Journal of Biomedical and Pharmaceutical Sciences*. 2012; 2(12):32-6.

