

# The protective effects of Curcumin and Caffeic acid alone or in combination on Nicotine-induced lung injury in rats

Sylvia A. Boshra<sup>1\*</sup>

## \*Corresponding author:

Sylvia A. Boshra

<sup>1</sup>Biochemistry Department, Faculty of Pharmacy, October 6 University, 6<sup>th</sup> of October City, Egypt

## Abstract

The present study was performed to explore the protective effects of caffeic acid (20 mg/kg.b.w) and curcumin (50mg/k.g.b.w.) on nicotine-induced lung injury alone and in combination. Their effect was compared to N-acetylcysteine (500mg/k.g.b.w.) as known modulator of oxidative stress. Nicotine treatment (0.6mg/kg/day, i.p, for 21 consecutive days) resulted in a significant increase ( $p < 0.05$ ) in plasma alanine aminotransferase (ALT), aspartate aminotransferase (AST), lactate dehydrogenase (LDH), total cholesterol (TC), triglycerides (TG), low density lipoprotein cholesterol (LDL-C) and well as plasma and lung thiobarbituric acid reactive substances (TBARS), nitric oxide (NO) and tumor necrosis factor- (TNF-) concomitant with significant decline in non-enzymatic antioxidant like reduced glutathione (GSH) and in enzymatic antioxidants like catalase (CAT) and superoxide dismutase (SOD) as well as high density lipoprotein cholesterol (HDL-C). Furthermore, nicotine treatment caused severe injury indicated by the histopathological examination of lung tissue compared to normal control group. Oral treatment with caffeic acid alone or curcumin alone or in combination as well as N-acetylcysteine alone prevented the elevation in plasma ALT, AST, LDH, TC, TG, LDL-C, NO, TNF- and TBARS levels concomitant with an increment in the HDL-C, reduced glutathione GSH and antioxidant enzymes (CAT and SOD) and amelioration in histopathological changes and injury induced by nicotine. Lung protection was prominent in curcumin and N-acetylcysteine alone more than caffeic acid alone or caffeic acid and curcumin combination. Moreover, curcumin has the potential to be used in a combination therapy with caffeic acid, with decreasing the therapeutic dose of caffeic acid and therefore its side-effects.

**Keywords:** Nicotine, lung, curcumin, caffeic acid, N-acetylcysteine, oxidative stress biomarkers

## Introduction

Nicotine, a major toxic component of cigarette smoking, has long been recognized to result in oxidative stress by inducing the generation of reactive oxygen species [1]. Nicotine, found in tobacco (*Nicotiana glauca*), is a natural alkaloid and an agent that weakens the immune system. Nicotine and metabolites increase lipid peroxidation and also affect the activities of antioxidant enzymes, thus, causing oxidative damage [2]. Nicotine increased oxidative stress results from excess generation of reactive chemical species called free radicals from a number of sources and/or from decreased enzymic and nonenzymic antioxidant defenses [3]. Oxygen and oxygen-derived free radicals are very important mediators of cell and tissue injury [3 & 4]. Free radicals are atoms or molecules that have one or more unpaired electrons in their outer orbital, which increase reactivity of the chemical

derivatives extraordinarily [5]. As a consequence, these radicals interact with molecules such as DNA, RNA and several proteins in their surroundings [2-6]. The interaction usually causes a decrease or even loss of function of these molecules. Although many different radicals occur in our body, the most effective ones are related to oxygen [5, 6]. Natural antioxidants are best obtained from plants. These phytonutrients serve as potential therapeutic agents against a wide variety of environmental stresses and pathological conditions. Curcumin is a  $\beta$ -diketone compound which contains two ferulic acid molecules linked via a methylene bridge at the carbon atoms of the carbonyl groups [7]. Various curcumin-related phenols (*Curcuminoids*) have also been found in edible plants, especially *Zingiberaceae* plants [8-10]. Extracts of rhizomes of turmeric have been widely used in Indian medicine and they are considered to be efficacious in the treatment of liver disorders and certain pyrogenic infections [11]. Curcumin exhibits anti-inflammatory [12], hepatoprotective [13] and inhibits of tumor initiation by various



carcinogens [14,15]. Caffeic acid (3,4-dihydroxycinnamic acid) is present in many plants and occurs in diet as part of fruits, tea, coffee and wine[16]. *In vivo* and *in vitro* studies have reported that caffeic acid showed antioxidant, free radical scavenging, antitumor and anti-inflammatory[17-19]. In addition; N-acetylcysteine is also widely used as an antiangiogenic [20], antifibrotic [21], neuroprotective [22], renoprotective [23], antioxidant [24] and as a chelating agent in the treatment of heavy metal poisoning [25]. N-acetylcysteine effectiveness is primarily attributed to its ability to reduce extracellular cystine to cysteine, and as a source of sulfhydryl groups [26]. The aim of this study was to elucidate the role of oxidative stress in nicotine-induced lung injury. In addition, studying the possible protective effects of different antioxidants as curcumin and caffeic acid alone and in combination on nicotine-induced lung injury and compare their effects with N-acetylcysteine; known modulator of oxidative stress.

## Materials and Methods

### Chemicals

Nicotine, Curcumin, caffeic acid and N-acetylcysteine were purchased from Sigma, USA.

#### Rats

This experiment was conducted in accordance with guidelines established by the Animal Care and use Committee of Faculty of Pharmacy, October 6 University, Egypt. Adult rats weighing around  $180 \pm 20$ gms were purchased from National Cancer Institute, Cairo University, Egypt. They were individually housed in cages in an air-conditioned room with a temperature of  $22 \pm 2^\circ\text{C}$ , a relative humidity of 60%, and an 8:00 to 20:00 light cycle. During the acclimatization period, each animal was raised on a regular diet *ad libitum*.

### Experimental design

The animals enrolled in the present study were divided into 6 groups, each group consists of 8 animals, two controls groups and four treatment groups:

Group (1): Control negative (0.9% saline, 3ml/kg.b.w., orally).

Group (2): Positive control (nicotine 0.6 mg/kg.b.w. suspended in 1ml 0.9% saline) was given I.P daily for 21 days [27].

Group (3): Nicotine 0.6 mg/kg.b.w./day(I.P.)+ caffeic acid (20mg/kg.b.w, orally) daily for 21 days [28].

Group (4): Nicotine 0.6 mg/kg.b.w.(I.P.)+ curcumin (50mg/k.g.b.w., orally) daily for 21 days [29].

Group (5): Nicotine 0.6 mg/kg.b.w.(I.P.)+ caffeic acid (20mg/kg.b.w orally) + curcumin (50mg/k.g.b.w.) daily for 21 days.

Group (6): Nicotine 0.6 mg/kg.b.w.+N-acetylcysteine (500mg/k.g.b.w., orally) daily for 21 days [30].

At the end of the experiment, rats of each group were sacrificed by cervical decapitation.

### Blood samples

Blood samples were collected at the end of experimental period in dry, clean, and screw capped tubes. Blood was divided into two parts; first part used for GSH [31], SOD [32] and CAT [33] estimation, Also, the second part was centrifugated at 2500r.p.m for 15 minutes. Plasma was separated by automatic pipette and received in dry sterile samples tube and kept in a deep freeze at  $-20^\circ\text{C}$  until used for subsequent biochemical analysis ALT [34], AST [34], LDH [35], TC [36], TG [37], HDL-C[38], LDL-C [39], NO[40], TNF- [41] and TBARS [42].

### Tissue specimen (lung tissue)

The abdomen and chest were opened and the lung specimen was quickly removed and opened gently using a scrapper, cleaned by rinsing with ice-cold isotonic saline to remove any blood cells, clots, then blotted between 2 filter papers and quickly stored in a deep freezer at  $(-20^\circ\text{C})$  for subsequent biochemical estimation of lung GSH [31], SOD [32], CAT [33] and TBARS [42]. Lung protein and blood hemoglobin (Hb) were estimated by the method of Lowry et al[43] and Jain [44], respectively.

### Histological assessment

lungs from rats of different groups were fixed in 10% neutral formalin solution, dehydrated in graded alcohol and embedded in paraffin. Fine sections obtained were mounted on glass slides and counter-stained with Hematoxylin and Eosin (H&E) for light microscopic analyses according to the method of Bancroft and Steven [45]. The slides were coded and examined by a histopathologist who was ignorant about the treatment groups after which photographs were taken.

### Statistical analysis

The obtained data were statistically analyzed and using the statistical package for social science (SPSS, 13.0 software, 2003) [46], for obtaining mean and standard deviation and error. The data were analyzed using one-way ANOVA to determine the statistical significance of differences among groups. Duncan's test was used for making a multiple comparisons among the groups for testing the inter-grouping homogeneity.

#### Results

Table 1 showed that I.P. administration of nicotine (0.6mg/kg.b.w) resulted in a significant increase in plasma ALT, AST and LDH compared to the normal control group ( $p < 0.05$ ). Supplementation of caffeic acid (20 mg/kg.b.w.) and curcumin (50 mg/k.g.b.w.) alone and in combination and N-acetylcysteine (500mg/k.g.b.w.) resulted in a significant decrease in plasma ALT, AST and ALP compared to the group that received nicotine ( $p < 0.05$ ). Effect of curcumin and N-

acetylcysteine alone was more than caffeic acid alone or caffeic acid and curcumin in combination.

Table 2 showed that I.P. administration of nicotine (0.6 mg/kg.b.w) resulted in a significant increase in plasma total cholesterol (TC), triglycerides (TG) and LDL-C as well as a significant decrease in plasma HDL-C compared to the normal control group ( $p < 0.05$ ). Oral supplementation of caffeic acid (20 mg/kg.b.w.), curcumin (50 mg/k.g.b.w.) alone or in combination and N-acetylcysteine (500mg/k.g.b.w.) resulted in a significant decrease in plasma total cholesterol (TC), triglycerides (TG) and LDL-C as well as a significant increase in plasma HDL-C compared to the group that received nicotine ( $p < 0.05$ ).

Table 3 showed that I.P. administration of nicotine (0.6 mg/kg.b.w) resulted in a significant decrease in blood reduced glutathione (GSH) and activities of superoxide dismutase (SOD) and catalase (CAT) as well as a significant increase in plasma TBARs compared to the normal control group ( $p < 0.05$ ). Supplementation of caffeic acid (20 mg/kg.b.w.) and curcumin (50 mg/k.g.b.w.) alone and in combination and N-acetylcysteine (500mg/k.g.b.w.) resulted in a significant increase in blood GSH, SOD and CAT as well as a significant decrease in plasma TBARs compared to the group that received nicotine ( $p < 0.05$ ).

Table 4 showed that I.P. administration of nicotine (0.6 mg/kg.b.w) resulted in a significant decrease in lung reduced glutathione (GSH) and activities of superoxide dismutase (SOD) and catalase (CAT) as well as a significant increase in lung TBARs compared to the normal control group ( $p < 0.01$ ). Supplementation of caffeic acid (20 mg/kg.b.w.) and curcumin (50 mg/k.g.b.w.) alone and in combination and N-acetylcysteine (500 mg/k.g.b.w.) resulted in a significant increase in lung GSH, SOD and CAT as well as a significant decrease in lung TBARs compared to the group that received nicotine ( $p < 0.05$ ).

Table 5 showed that administration of nicotine (0.6 mg/kg.b.w., I.P.) resulted in a significant increase in plasma and lung nitrous oxide (NO) and tumor necrosis factor- (TNF-) compared to the normal control group ( $p < 0.05$ ). Administration of caffeic acid (20 mg/kg.b.w.) and curcumin (50 mg/k.g.b.w.) alone and in combination and N-acetylcysteine (500 mg/k.g.b.w.) to rats resulted in a significant decrease in plasma and lung nitrous oxide (NO) and tumor necrosis factor- (TNF-) compared to the group that received nicotine ( $p < 0.05$ ).

**Table 1:** Activity of alanine transaminase (ALT), aspartate transaminase (AST), lactate dehydrogenase (LDH) of normal and experimental groups of rats.

No.	Groups	ALT (U/L)	AST (U/L)	LDH (U/L)
(I)	Normal 0.9% saline	35.47 ± 2.36	17.60 ± 3.10	110.96 ± 6.44
(II)	Positive control: Nicotine (0.6mg/Kg.b.w.)	83.11 ± 4.50 <sup>a</sup>	52.80 ± 4.67 <sup>a</sup>	225.08 ± 11.39 <sup>a</sup>
(III)	Nicotine (0.6mg/Kg.b.w.) + Caffeic acid (20mg/Kg.b.w.)	61.48 ± 3.08 <sup>ab</sup>	44.23 ± 5.70 <sup>ab</sup>	146.00 ± 8.50 <sup>ab</sup>
(IV)	Nicotine (0.6mg/Kg.b.w.) + Curcumin (50mg/Kg.b.w.)	40.83 ± 5.24 <sup>bc</sup>	19.40 ± 4.64 <sup>bc</sup>	110.22 ± 12.80 <sup>bc</sup>
(V)	Nicotine (0.6mg/Kg.b.w.) + Caffeic acid (20mg/Kg.b.w.) + Curcumin (50mg/Kg.b.w.)	57.33 ± 4.20 <sup>bcd</sup>	26.15 ± 2.98 <sup>abcd</sup>	128.07 ± 11.40 <sup>bcd</sup>
(VI)	Nicotine (0.6mg/Kg.b.w.) + N-acetylcysteine (500mg/Kg.b.w.)	38.60 ± 2.68 <sup>bce</sup>	20.59 ± 3.26 <sup>bcde</sup>	109.25 ± 9.80 <sup>bce</sup>

Values are given as mean ± SD for groups of eight animals each. \* Significantly different at  $p < 0.05$ : a: significant from normal control; b: significant from nicotine (0.6mg/Kg.b.w.) supplement group; c: significant from caffeic acid (20mg/Kg.b.w.); d: significant from curcumin (50mg/Kg.b.w.); e: significant from caffeic acid+ curcumin group of rats.

**Table 2:** Level of plasma total cholesterol (TC), triglycerides (TG), HDL-C and LDL-C of normal and experimental groups of rats.

No.	Groups	TC (mg/dl)	TG (mg/dl)	HDL-C (mg/dl)	LDL-C (mg/dl)
(I)	Normal 0.9% saline	129.06 ± 7.87	105.38 ± 11.24	42.25 ± 4.03	65.71 ± 6.11
(II)	Positive control: Nicotine (0.6mg/Kg.b.w.)	212.38 ± 8.56 <sup>a</sup>	162.80 ± 5.73 <sup>a</sup>	25.47 ± 3.66 <sup>a</sup>	154.4 ± 9.80 <sup>a</sup>
(III)	Nicotine (0.6mg/Kg.b.w.) + Caffeic acid (20mg/Kg.b.w.)	163.46 ± 9.97 <sup>ab</sup>	129.29 ± 5.40 <sup>ab</sup>	30.29 ± 2.60 <sup>ab</sup>	107.67 ± 11.25 <sup>ab</sup>
(IV)	Nicotine (0.6mg/Kg.b.w.)+ Curcumin (50mg/Kg.b.w.)	133.60 ± 12.04 <sup>bc</sup>	106.74 ± 5.24 <sup>bc</sup>	39.06 ± 4.58 <sup>bc</sup>	73.24 ± 6.48 <sup>bc</sup>
(V)	Nicotine (0.6mg/Kg.b.w.)+ Caffeic acid (20mg/Kg.b.w.) + Curcumin (50mg/Kg.b.w.)	145.87 ± 9.98 <sup>abcd</sup>	115.59 ± 5.07 <sup>abcd</sup>	34.80 ± 6.68 <sup>abcd</sup>	87.96 ± 9.16 <sup>abcd</sup>
(VI)	Nicotine (0.6mg/Kg.b.w.) + N-acetylcysteine(500mg/Kg.b.w.)	131.25 ± 11.69 <sup>bce</sup>	102.65 ± 6.20 <sup>bce</sup>	38.17 ± 3.65 <sup>bce</sup>	72.58 ± 5.00 <sup>abc</sup>

Values are given as mean ± SD for groups of eight animals each. \* Significantly different at  $p < 0.05$ . a: significant from normal control; b: significant from nicotine (0.6mg/Kg.b.w.) supplement group; c: significant from caffeic acid (20mg/Kg.b.w.); d: significant from curcumin (50mg/Kg.b.w.); e: significant from caffeic acid+ curcumin (50mg/Kg.b.w.) group of rats. LDL-C (mg/dl) = TC-HDL-[TG / 5].

**Table 3:** Level of blood reduced glutathione (GSH) and activities of superoxide dismutase (SOD), catalase (CAT) and plasma Thiobarbituric acid reactive substances (TBARs) in normal and experimental groups of rats.

No.	Groups	GSH (mg %)	SOD (U/g Hb)	CAT (U/g Hb)	TBARs (mmol/dL)
(I)	Normal 0.9% saline	34.64 ± 4.09	14.38 ± 0.34	3.68 ± 0.55	4.23 ± 0.41
(II)	Positive control: Nicotine (0.6mg/Kg.b.w.)	12.59 ± 3.84 <sup>a</sup>	6.70 ± 0.29 <sup>a</sup>	1.65 ± 0.43 <sup>a</sup>	8.05 ± 1.29 <sup>a</sup>
(III)	Nicotine (0.6mg/Kg.b.w.) + Caffeic acid (20mg/Kg.b.w.)	19.88 ± 1.49 <sup>ab</sup>	13.15 ± 0.34 <sup>ab</sup>	2.75 ± 0.24 <sup>ab</sup>	5.80 ± 0.79 <sup>b</sup>
(IV)	Nicotine (0.6mg/Kg.b.w.)+ Curcumin (50mg/Kg.b.w.)	30.19 ± 4.23 <sup>bc</sup>	14.08 ± 0.21 <sup>b</sup>	3.88 ± 0.44 <sup>bc</sup>	4.15 ± 0.44 <sup>bc</sup>
(V)	Nicotine (0.6mg/Kg.b.w.)+ Caffeic acid (20mg/Kg.b.w.) + Curcumin (50mg/Kg.b.w.)	23.27 ± 2.23 <sup>abcd</sup>	10.30 ± 0.14 <sup>bd</sup>	3.00 ± 0.45 <sup>abc</sup>	4.62 ± 0.45 <sup>bd</sup>
(VI)	Nicotine (0.6mg/Kg.b.w.) + N-acetylcysteine(500mg/Kg.b.w.)	33.40 ± 2.74 <sup>bce</sup>	13.96 ± 0.43 <sup>bce</sup>	3.76 ± 0.25 <sup>bce</sup>	4.03 ± 0.60 <sup>bce</sup>

Values are given as mean ± SD for groups of eight animals each. \* Significantly different from normal group at  $p < 0.05$ . a: significant from normal control; b: significant from nicotine (0.6mg/Kg.b.w.) supplement group; c: significant from caffeic acid (20mg/Kg.b.w.); d: significant from curcumin (50mg/Kg.b.w.); e: significant from caffeic acid+ curcumin (50mg/Kg.b.w.) group of rats.

**Table 4:** Level of lung reduced glutathione (GSH) and activities of superoxide dismutase (SOD), catalase (CAT) and Thiobarbituric acid reactive substance (TBARs) in normal and experimental groups of rats.

No.	Groups	GSH ( $\mu\text{g}/\text{mg}$ protein)	SOD	CAT	TBARs ( $\text{mmol}/\text{g}$ tissue)
(I)	Normal 0.9% saline	113.58 $\pm 6.98$	15.69 $\pm 2.38$	48.37 $\pm 4.50$	1.17 $\pm 0.28$
(II)	Positive control: Nicotine (0.6mg/Kg.b.w.)	66.38 $\pm 3.36^{\text{a}}$	8.86 $\pm 1.20^{\text{a}}$	26.80 $\pm 3.15^{\text{a}}$	2.84 $\pm 0.35^{\text{a}}$
(III)	Nicotine (0.6mg/Kg.b.w.) + Caffeic acid (20mg/Kg.b.w.)	85.25 $\pm 8.27^{\text{ab}}$	11.45 $\pm 0.88^{\text{b}}$	31.35 $\pm 3.10^{\text{ab}}$	1.80 $\pm 0.21^{\text{ab}}$
(IV)	Nicotine (0.6mg/Kg.b.w.)+ Curcumin (50mg/Kg.b.w.)	106.63 $\pm 5.72^{\text{bc}}$	15.37 $\pm 2.05^{\text{bc}}$	42.68 $\pm 3.03^{\text{bc}}$	1.03 $\pm 0.19^{\text{bc}}$
(V)	Nicotine (0.6mg/Kg.b.w.)+ Caffeic acid (20mg/Kg.b.w.) + Curcumin (50mg/Kg.b.w.)	95.88 $\pm 8.63^{\text{bc}}$	10.52 $\pm$ 0.99 <sup>abcd</sup>	36.08 $\pm 5.95^{\text{bcd}}$	1.66 $\pm 0.13^{\text{abcd}}$
(VI)	Nicotine (0.6mg/Kg.b.w.) + N-acetylcysteine(500mg/Kg.b.w.)	109.75 $\pm 4.43^{\text{bce}}$	14.68 $\pm 2.21^{\text{bce}}$	45.26 $\pm 4.16^{\text{bce}}$	1.15 $\pm 0.49^{\text{bce}}$

Values are given as mean  $\pm$  SD for groups of eight animals each. \* Significantly different from at  $p < 0.05$ . SOD; one unit of enzyme activity was taken as the enzyme reaction, which gave 50% inhibition of pyrogallol autoxidation in one minute/mg protein; CAT;  $\mu\text{mol}$  of  $\text{H}_2\text{O}_2$  consumed/min/mg protein; a: significant from normal control; b: significant from nicotine (0.6mg/Kg.b.w.) supplement group; c: significant from caffeic acid (20mg/Kg.b.w.); d: significant from curcumin (50mg/Kg.b.w.); e: significant from caffeic acid+ curcumin (50mg/Kg.b.w.) group of rats.

**Table 5:** Levels of plasma and lung nitrous oxide (NO) and tumor necroses factor- (TNF- ) of normal and experimental groups of rats

No.	Groups	NO		TNF-	
		Plasma ( $\mu\text{mol}/\text{ml}$ )	Lung ( $\mu\text{mol}/\text{gtissue}$ )	Plasma pg/ml	Lung (U/g tissue)
(I)	Normal 0.9% saline	15.86 $\pm 2.98$	50.06 $\pm 8.47$	225.35 $\pm 11.42$	20.76 $\pm 3.96$
(II)	Positive control: Nicotine (0.6mg/Kg.b.w.)	37.05 $\pm 3.94^{\text{a}}$	92.74 $\pm 8.93^{\text{a}}$	298.82 $\pm 17.33^{\text{a}}$	46.38 $\pm 5.20^{\text{a}}$
(III)	Nicotine (0.6mg/Kg.b.w.) + Caffeic acid (20mg/Kg.b.w.)	25.49 $\pm 3.74^{\text{ab}}$	78.55 $\pm 4.98^{\text{ab}}$	252.28 $\pm 4.15^{\text{ab}}$	31.56 $\pm 3.85^{\text{ab}}$
(IV)	Nicotine (0.6mg/Kg.b.w.)+ Curcumin (50mg/Kg.b.w.)	17.27 $\pm 2.97^{\text{bc}}$	49.06 $\pm 3.41^{\text{bc}}$	216.08 $\pm 13.72^{\text{bc}}$	18.46 $\pm 3.59^{\text{bc}}$
(V)	Nicotine (0.6mg/Kg.b.w.)+ Caffeic acid (20mg/Kg.b.w.) + Curcumin (50mg/Kg.b.w.)	22.55 $\pm$ 3.43 <sup>abcd</sup>	60.93 $\pm 4.86^{\text{bcd}}$	244.63 $\pm 7.91^{\text{bcd}}$	27.90 $\pm 2.49^{\text{abcd}}$
(VI)	Nicotine (0.6mg/Kg.b.w.) + N-acetylcysteine(500mg/Kg.b.w.)	16.67 $\pm$ 2.90 <sup>bce</sup>	44.48 $\pm 5.90^{\text{bce}}$	222.39 $\pm 9.42^{\text{bce}}$	20.22 $\pm 3.83^{\text{bce}}$

Values are given as mean  $\pm$  SD for groups of eight animals each. \* Significantly different at  $p < 0.05$ . a: significant from normal control; b: significant from nicotine (0.6mg/Kg.b.w.) supplement group; c: significant from caffeic acid (20mg/Kg.b.w.); d: significant from curcumin (50mg/Kg.b.w.); e: significant from caffeic acid+ curcumin (50mg/Kg.b.w.) group of rats.

### Histopathology examination of the lung tissues

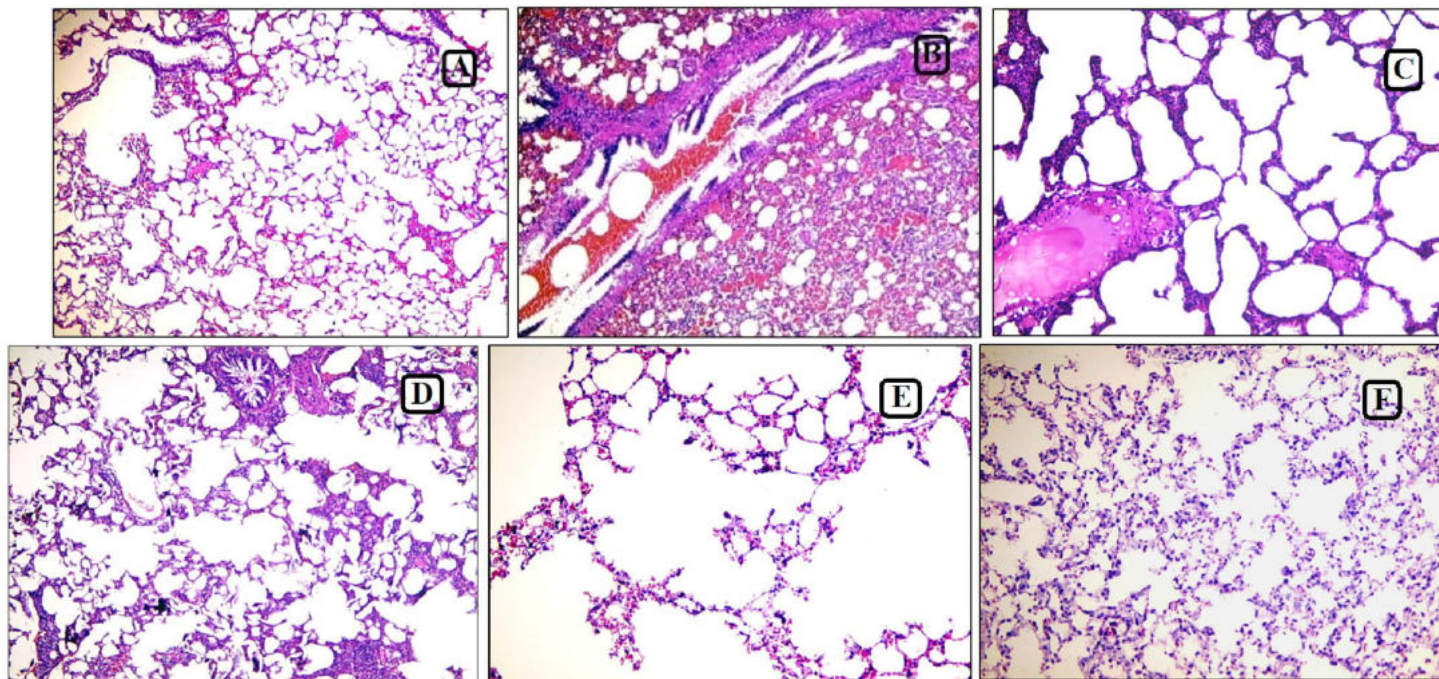
Group (1) control negative group showed micrograph of lung section is normal the bronchial veolar unit parenchyma, is normal (Figure 1A). Group 2 (control positive; rats treated with nicotine (0.6 mg/kg.b.w.) showed partially collapsed lung tissue with severe congestion and dense inflammation. (Figure 1B). Group (3); rats

treated with nicotine (0.6 mg/kg.b.w.) + caffeic acid (20mg/Kg.b.w.) showed refilling of the alveoli, marked decreased congestion and inflammation (Figure 1C). Group (4); rats treated with nicotine (0.6mg/Kg.b.w.)+ curcumin (50mg/Kg.b.w.) showed refilling of the alveoli, mild decreased congestion and inflammation. (Figure 1D). Group 5; rats treated with nicotine (0.6mg/Kg.b.w.)+ curcumin (50mg/Kg.b.w.) +caffeic acid (20mg/Kg.b.w.) showed



refilling of the alveoli with compensatory emphysema (Figure 1E). Group (6); rats treated with nicotine (0.6 mg/kg.b.w.) + N-acetylcysteine(500mg/Kg.b.w.) showed improvement with refilling

of the alveoli, marked decreased of congestion and inflammation with reparative type II pneumocytes hyperplasia.



**Figure (1A-F):** Sections in the lungs of groups at 400 magnification:

**Figure (1A):** Negative control of rat lung. **Figure (1B):** Positive control (nicotine, 0.6mg/Kg.b.w) rat lung with inflammation. **Figure (1C):** nicotine (0.6 mg/Kg.b.w.) + caffeic acid (20mg/Kg.b.w), lung tissue showed mild inflammation. **Figure (1D):** nicotine (0.6 mg/Kg.b.w) + curcumin (50 mg/Kg.b.w), the lung tissue showed refilling of the alveoli. **Figure (1E):** nicotine (0.6 mg/Kg.b.w) + curcumin (50 mg/Kg.b.w) + caffeic acid (20mg/Kg.b.w), the lung showed moderate decrease inflammation. **Figure (1F):** nicotine (50mg/Kg.b.w) + N-acetylcysteine (500mg/Kg.b.w.) showed improvement with refilling of the alveoli.

## Discussion

Nicotine, the major component of cigarette smoke, plays an important role in the development of lung complications. Early-stage disease can be treated with curative intent although the risk for relapse is notoriously high. Unfortunately, the majority of lung cancer patients present at an advanced stage. Despite an initial response to treatment, most of these late stage patients will eventually progress on standard therapy and die from their disease. Despite the complex nature of lung cancer biology, its molecular underpinnings are becoming increasingly clear [47 & 48]. Nicotine is considered a prototype polycyclic aromatic hydrocarbon (PAH), classic DNA damaging agent and carcinogen. Antioxidants are the first source of protection of the body against free radicals and other oxidants, being the compounds that attack the formation of radical species within cells. The group of antioxidants inside the organism is known as the total antioxidant state (TAS) [49]. The antioxidant

protection of human cells includes enzyme mediated and non-enzymatic defense mechanisms. Superoxide dismutase (SOD), catalase (CAT) and glutathione-peroxidase (Gpx) are the most important antioxidant enzymes. SOD catalyses the reaction of superoxide anion to hydrogen peroxide ( $H_2O_2$ ); in turn, CAT converts  $H_2O_2$  into water and oxygen. The affinity of CAT for  $H_2O_2$  is relatively low, therefore, some  $H_2O_2$  remains in the cell. GSH-px is capable of detoxifying the remaining  $H_2O_2$  [50]. The present study showed that oral administration of nicotine (0.6 mg/kg.b.w.) resulted in a significant increase in plasma ALT, AST and LDH. Some investigators claimed ALT, AST and LDH increased by cigarette smoking [51] which agreed with our results. Liver enzymes were strongly influenced by smoking, consistent with other studies [52, 53] concerning osteoporosis have documented increased serum ALT, AST and LDH levels in current smokers, as a mainly marker of the liver and bones turnover and also present in the kidneys and leukocytes count [54].

The present study showed that oral administration of curcumin (50 mg/kg.b.w.) resulted in a significant decrease in plasma ALT, AST

and LDH. Curcumin, an antioxidant and anticarcinogenic substance, was reported to have a protective effect against liver damage [55]. The protective action of curcumin against adverse effects of toxicants had also been reported [56]. Curcumin could exert antioxidative effects either directly as a chemical antioxidant due to its ability to scavenge reactive oxygen and nitrogen free radicals or by modulating cellular defenses which themselves exert antioxidant effects [57, 58].

Oral administration of caffeic acid at the present dose (20mg/kg b.w) attenuated the nicotine induced elevation of the plasma levels of these marker enzymes indicating that caffeic acid may have the capacity to provide protection to the rat hepatic tissue. This hepatoprotection might

have been exerted through its membrane stabilizing and antilipoperoxidative property. Pari and Prasath [59] have already shown the membrane stabilizing and anti-lipoperoxidative effects of caffeic acid in protecting cell membrane against peroxidative damage. Furthermore, N-acetylcysteine (500mg/Kg.b.w.) treatment ameliorated hepatic injury in terms of the AST, ALT and LDH levels against nicotine treated rats. The reason may be due to N-acetylcysteine with its antioxidant effects could prevent nicotine-induced oxidative damage. This evidence of liver damage are consistent with the previous findings by Santra et al [60] who mentioned that administration of N-acetylcysteine stimulates the synthesis of glutathione, which exhibited therapeutic effects on nicotine-induced liver fibrosis in rats.

The present study showed that administration of nicotine resulted in a significant increase in plasma total cholesterol, triglycerides and LDL-cholesterol as well as significant decrease in HDL-cholesterol compared to the normal control group which are in agreement with the earlier studies [61, 62]. The increased free fatty acids in tissues of nicotine treated rats may serve as the substrate for lipid peroxidation. In the present study the cholesterol level was elevated in the nicotine-treated animals. The prevalence of hypercholesterolemia and triglyceridemia has been reported in heavy smokers [63]. This increased level of cholesterol is attributed to the increased activity of 3-hydroxy-3-methyl-glutaryl CoA reductase (HMG-CoA reductase) and increased incorporation of labelled acetate into cholesterol [64]. In curcumin treated group the level of cholesterol is decreased probably due to the increase in the CYP7A1 gene expression which is a rate limiting enzyme in the biosynthesis of bile acid from cholesterol as suggested by Kim and Kim [65]. Curcumin showed an obvious hypocholesterolemic effect that could be due to an effect on total cholesterol absorption [66], degradation or elimination [67]. Triglyceride lowering effect is probably due to multiple inductions of fatty acid catabolism [68]. The influence of Caffeic acid at the TG, TC and LDL-cholesterol might be attributed to Caffeic acid decrease triglyceride creation by decrease the countenance of both HMG-CoA reductase and glycerol three phosphate acyltransferase in the liver by arrangement of adenosine mono phosphate activated kinase [69]. While that increase of HDL-cholesterol, in all study groups treated with caffeic acid might be indicated that doses of caffeic acid may

have beneficial effect on HDL-cholesterol, by its effect as anti-lipid oxidation and suggested that caffeic acid lead to diminish oxygen species, and thus reduces DNA from impairment, which could be important in the regulation of liver function, this result agreement with result recorded by Codrington, et al [70]. N-acetylcysteine has been shown to reduce cholesterol levels in plasma of rats [71]. Krieger et al. [72] mentioned a slight reduction in plasma lipid fractions by means of N-acetylcysteine supplementation in hypercholesterolemic LDL receptor. Putative mechanisms accounting for the lipid-lowering effects of N-acetylcysteine might be related to its antioxidant properties. N-acetylcysteine in our study led to reduced plasma LDL-C. The maintenance of the normal structure of lipoprotein receptors is indispensable for their function, improving the cellular uptake of plasma lipids from the blood. Reactive oxygen species produced during oxidative stress react with lipoproteins to produce oxidation states, diminishing the cellular uptake of lipids from the blood [73-75]. Thus the antioxidant action of N-acetylcysteine might contribute to elevated cellular lipid uptake, resulting in the decrease of serum cholesterol levels. According to Lin and Yin [76], the lipid lowering action of N-acetylcysteine in mice consuming a high fat diet is attributed partially to the suppression of mRNA expression of three lipogenic-related enzymes (malic enzyme, fatty acid synthase and 3-hydroxy-3-methylglutaryl coenzyme A reductase).

The present study showed that oral administration of nicotine (0.6 mg/kg.b.w) resulted in a significant increase in plasma and lung TBARs as well as significant decrease in blood and lung SOD, GSH and CAT compared to the normal control group. Nicotine is a highly addictive alkaloid induced oxidative stress both *in vivo* and *in vitro* [77]. It has been reported that the nicotine disrupts the mitochondrial respiratory chain leading to an increase generations of super oxide ions and hydrogen peroxide [78]. Superoxide anion and hydrogen peroxide are the main sources of the nicotine induced free radical generation and depletion of the cellular antioxidant [79]. Glutathione being an important cellular reductant involves in protection against free radicals, peroxides and toxic compounds [80]. Therefore depletion of GSH not only impairs cell defense against toxic compounds but also results in enhanced oxidative stress and tissue damage [81, 82]. Our observation shows that nicotine treatment more significantly ( $p < 0.05$ ) depletes GSH, SOD and CAT level of blood and lung of both dietary groups indicating higher level of tissue damage. Curcumin reduces the effect in nicotine treated group due to its high antioxidant capacity [83]. The supplementation of caffeic acid is found to be effective in decreasing the lipid peroxidation, lipid hydroperoxides and conjugated diene levels in the hepatic tissue. Lipid peroxidation, which is self-perpetuating, terminates when chain breaking antioxidants acts in any step of the chain. This indicates that caffeic acid has antilipoperoxidative and anti-oxidative properties. The above effect, of caffeic acid is favored by the presence of two hydroxyl groups attached to its main ring that may produce a site for chelation. In this context, caffeic acid has already been shown to chelate the free radicals [84]. N-

acetylcysteine treatment enhancement the levels of antioxidant enzymes SOD and CAT beside GSH content and ameliorated the levels of MDA in plasma and lung as compared to nicotine group. These results may be due to the antioxidant scavenging properties of N-acetylcysteine to remove the ROS like OH<sup>-</sup>, O<sup>-</sup> and H<sub>2</sub>O<sub>2</sub> radicals which librated from nicotine treatment. This result was coincide with Gurer et al [85] who reported that, N-Acetylcysteine is a thiolcontaining antioxidant that has been used to reduce various conditions of oxidative stress. Its antioxidant action is attributed to GSH synthesis; therefore maintaining intracellular GSH levels and scavenging reactive oxygen species (ROS) [86].

The present study showed that oral administration of nicotine (0.6 mg/kg.b.w) resulted in a significant increase in plasma and lung NO and TNF- compared to the normal control group.

Curcumin is a potent "scavenger" of the superoxide radical, a free radical that initiates potentially harmful oxidative processes such as lipid peroxidation [87]. Through in Curcumin also increases survival of cells exposed *in vitro* to the enzyme hypoxanthine/xanthine oxidase, which stimulates superoxide and hydrogen peroxide production. Curcumin demonstrates several other *in vitro* effects linked to free radical scavenging. Moreover, curcumin has also been shown to quench reactive oxygen species and scavenge superoxide anion radicals and hydroxyl radicals and strongly inhibits nitric oxide (NO) production by down-regulating inducible nitric oxide synthase gene expression[88, 89].Furthermore, caffeic acid has been reported to up-regulate the expression of Nrf-2 gene and downregulate the expression of NF-kB gene by which it can reduce the generation of free radicals via the augmentation of

cellularanti-oxidative machinery in cells [90]. Also, In the current study, N-acetylcysteine significantly attenuated NO and TNF- -induced reduction of the activities of SOD and GSH, two major endogenous antioxidants, and attenuated TNF- -induced increase in the lipid peroxidation product malondialdehyde. This may represent a major mechanism by which N-acetylcysteine attenuates TNF- -induced cell toxicity [91, 92].

Finally, histopathological examination of lung showed severe congestion and dense inflammation in nicotine-treated rats (Figure 1B). Comparing the beneficial effect of caffeic acid and curcumin alone and in combination as well as N-acetylcysteinewith that of nicotine-induced lung toxicity. Results indicated that curcumin, caffeic acid and N-acetylcysteine showed refilling of the alveoli, decreased congestion and inflammation(fig. 1C-F).

In conclusion, the present study showed that caffeic acid and curcumin alone and in combination as well as N-acetylcysteinehas a powerful antioxidant and lung protective activity against nicotine induced lungtoxicity. These effects could be due to membrane protective action of tested compounds by scavenging the free radicals and its antioxidant action.

## Acknowledgement

Thanks to Ass. Prof. Dr Amal Haridy, National Cancer Institute, Cairo University, Egypt, for her involvement in histopathological examination.

## References

- [1]. Sener G, Toklu HZ, Cetinel S.  $\beta$ -Glucan protects against chronic nicotine induced oxidative damage in rat kidney and bladder. *Environmental Toxicology Pharmacology*, 2007; 23: 25–32.
- [2]. Siktar E, Ekinci D, Siktar E. Protective role of L-carnitine supplementation against exhaustive exercise induced oxidative stress in rats. *European Journal of Pharmacology*, 2011; 668(3): 407–413.
- [3]. Nallella A, Allamaneni SS, Said TM. Role of antioxidants in treatment of male infertility Agarwal: An overview of the literature. *Reprod Biomed Online*, 2004; 8:616–27.
- [4]. Gitto E, Reiter RJ, Karbownik M. Causes of oxidative stress in the pre- and perinatal period. *Biology of the Neonate*, 2002; 81: 146–157
- [5]. Osuna C, Reiter R, Garcia JJ. Inhibitory effect of melatonin on homocysteine-induced lipid peroxidation in rat brain homogenate. *Pharmacology and Toxicology*, 2002; 90: 32–37.
- [6]. Halliwell B, Gutteridge JMC. Oxygen toxicity, oxygen radicals, transition metals and disease. *Biochemical Journal*, 1984; 219: 1–14.
- [7]. Sharma OP. Antioxidant activity of curcumin and related compounds. *Biochem. Pharmacol.*, 1979; 25, 2811–1812.
- [8]. Toda S, Miyase T, Arichi H, Tanizawa H, Takino Y. Natural antioxidants III, antioxidant compounds isolated from rhizomes of curcuma Longa. *Chem. Pharm. Bull.*, 1985; 33, 1725-1728.
- [9]. Masuda T, Isobe J, Jitoe A, Nakatani N. Antioxidative curcuminoids from rhizomes of curcuma xanthorrhiza. *Phytochemistry*, 1992; 31, 3645-3647.
- [10]. Masuda T, Hidaha K, Shinohara A, Mackawa T, Takeda Y, Yamaguchi H. Chemical studies on antioxidant mechanism of curcuminoid: analysis of radical reaction products from curcumin. *J. Agric. Food. Chem.*, 1999; 47, 71-77.
- [11]. Nadakarni AK (1954). *Indian Materia Medica*, Popular Book Depot, Bombay, 1<sup>st</sup> ed., pp.414-42.
- [12]. Joe B, Vijaykumar M, Lokesh B. Biological properties of curcumin-cellular and molecular mechanisms of



- action. *Crit. Rev. Food Sci. Natur.*, 2004; 44, 97-111.
- [13]. Sahu SC, Washington MC. Effect of ascorbic acid and curcumin on quercetin-induced nuclear DNA damage, lipid peroxidation and protein degradation. *Cancer Lett.*, 1992; 63, 237-241.
- [14]. Lim G, Chu T, Young F. The curry spice curcumin reduces oxidative damage and amyloid pathology in an Alzheimer transgenic mouse. *J. Neurosci.*, 2001; 21, 8370-8377.
- [15]. Kawamori T, Lubet R, Steele V. Chemopreventive effect of curcumin, a naturally occurring anti-inflammatory agent, during the promotion/progression stages of colon cancer. *Cancer Res.*, 1999; 59, 597-601.
- [16]. Challis L, Beutler E. Active transport of glutathionedisulfide from erythrocytes. In: Larson, A., Orrenius, S., Holmgren, A., Mannerwik, B., editors. *Functions of glutathione, biochemical, physiological, toxicological and clinical aspects*, New York, USA, Raven Press, 1987. 65.
- [17]. Chan JH, Ho CT. Antioxidant activities of caffeic acid and its related hydroxy cinnamic acid compounds. *J. Agric. Food Chem.* 1997; 45, 2374-2378.
- [18]. Chen YJ, Shiao MS, Wang SY. The antioxidant caffeic acid phenethyl ester induces apoptosis associated with selective scavenging of hydrogen peroxide in human leukemic HL-60 cells. *Anticancer Drugs* 2001; 12, 143-149.
- [19]. Lee KJ, Choi JH, Khanal T, Hwang YP, Chung YC, Jeong HG. Protective effect of caffeic acid phenethyl ester against carbon tetrachloride induced hepatotoxicity in mice. *Toxicology*. 2008; 248, 18-24.
- [20]. Albin A, Morini M, Agostini FD, Ferrari N, Campelli F, Arena G, Noonan DM, Pesce C, Flora SD. Inhibition of angiogenesis driven Kaposi's, sarcoma tumor growth in nude mice by oral N-acetylcysteine. *Cancer Res*, 2001; 61: 8171-8178.
- [21]. Filho GP, Ferriera C, Schwengber A, Marroni C, Zettler C, Marroni N. Role of N-acetylcysteine on fibrosis and oxidative stress in cirrhotic rats. *Arq. Gastroenterol.*, 2008; 45: 156-162.
- [22]. Eskiocak S, Altaner S, Bayir S, Cakir E. The effect of N-acetylcysteine on brain tissue of rats fed with high cholesterol diet. *Turk. J. Biochem*, 2008; 33:58-63.
- [23]. Heyman SN, Goldfarb M, Shina A, Karmeli F, Rosen S. N-acetylcysteine ameliorates renal microcirculation: studies in rats. *Kidney Int*, 2003; 63: 634-641.
- [24]. Modi M, Kaul RK, Kannan GM, Flora SJS. Co-administration of zinc and N-acetylcysteine prevents arsenic induced tissue oxidative stress in male rats. *J. Trace Elem. Med. Biol*, 2006; 20: 197-204.
- [25]. De Vries N, De Flora S. N-acetylcysteine. *J. cell. Biochem. (suppl.)*, 1993; 17F: 270 - 277.
- [26]. Varadharajan V, Ganesan J. Restoration of Antioxidant Activity by N-acetylcysteine and Gallic Acid on Kidney Tissue of Mercuric Chloride Intoxicated Wistar Rats. *International Journal of Biological & Pharmaceutical Research*, 2013; 4(4): 302-307.
- [27]. Liu, RH. Potential synergy of phytochemicals in cancer prevention: mechanism of action. *The Journal of Nutrition*, 2004; 134: 3479S-3485S.
- [28]. Miltonprabu S, Shagirtha. K. Caffeic acid potentially attenuates cadmium induced oxidative stress mediated hepatotoxicity in rats. *The Journal of Free Radicals and Antioxidants*. 2013; 139: 141 -152.
- [29]. Choudhary D, Chandra D, Kale R. Modulation of radioresponse of glyoxalase system by curcumin. *Ethnopharmacol.*, 1999; 64, 1-7.
- [30]. Kucukardali Y, Cinan U, Acar HV, Ozkan S, Top C, Nalbant S, Cermik H, Cankir Z, Danaci M. Comparison of the C.L. Sprague, A.A. Therapeutic efficacy of 4-methylpyrazole and N-acetylcysteine on acetaminophen (paracetamol) hepatotoxicity in rats. *Curr. Med. Res. Opin*, 2005; 18: 78-81.
- [31]. Chanarin I. *Text book of Laboratory Haematology: An Account of Laboratory techniques*, Churchill Livingstone, New York 1989. 107.
- [32]. Marklund S, Marklund D. Involvement of the superoxide anion radical in the autoxidation of pyrogallol and a convenient assay for superoxide dismutase. *Eur. J. Biochem.*, 1974; 47:469.
- [33]. Sinha AK. Colorimetric assay of catalase. *J. Anal Biochem.* 1972; 47 (2): 389-94.
- [34]. Reitman S, Frankel SA. Colorimetric method for the determination of serum oxaloacetic acid and glutamic pyruvic transaminases. *Am. j. Clin. Pathol.* 1957; 28: 56 - 63.
- [35]. Buhl SN, Jackson K. Optimal conditions and comparison of lactate dehydrogenase catalysis of the lactate to pyruvate to lactate reactions in human serum at 25, 30 and 37 °C. *Clin. Chem.* 1978; 24:15: 828-833.
- [36]. Fossati P, Prencipe L. Serum triacylglycerols determined calorimetrically with an enzyme that produces hydrogen peroxide. *Clin Chem.* 1982; 1: 2077-2080.
- [37]. Allain CC, Poon LS, Chan CS. Enzymatic determination of total serum cholesterol. *Clin Chem.* 1974; 4: 470-475.
- [38]. Burnstein M, Selvenick HR, Morfin R. Rapid method for isolation of lipoprotein from human serum with polyanions. *J Lipid Res.*, 1970; 11: 583-395.
- [39]. Friedewald WT. Estimation of concentration of low-density lipoprotein cholesterol in plasma without use of the preparative ultracentrifuge. *Clin Chem.* 1972; 18:499-502.
- [40]. Miranda KM, Espey MG, Wink DA. A rapid, simple spectrophotometric method for simultaneous detection of nitrate and nitrite. *Nitric Oxide*. 2001; 5: 62-71.
- [41]. Beyaert R, Fiers W (1998): Tumor Necrosis Factor and Lymphotoxin. In Cytokines, A.R.M.-S. a. R. Thorpe, eds. Academic Press, San Diego, 1998; 335-360.
- [42]. Nichans WH, Samulelson B. Formation of malondialdehyde from phospholipid

- arachidonate during microsomal lipid peroxidation. *Eur J Biochem*, 1968;6: 126-30.
- [43]. Lowry OH, Rosebrough NJ, Farr AL, Randall R: Protein measurement with Folin's phenol reagent. *J Biol Chem* 1952;193: 265-275.
- [44]. Jain WC: Schalm's Veterinary Hematology, ed 4, Lea and Febiger, Philadelphia. 1986, pp. 69 - 71.
- [45]. Bancroft GD, Steven A. In. Theory and practice of histological technique 4<sup>th</sup> Ed. London; Churchill Livingstone. 1983; Pp. 99 – 112.
- [46]. Abo-Allam RM, 2003. Data Statistical Analysis using SPSS Program. 1 Ed., Publication for 1<sup>st</sup> Universities, Cairo.
- [47]. Salgia R, Hensing T, Campbell N. Personalized treatment of lung cancer. *Semin Oncol*. 2011; 38: 274–283.
- [48]. Teixeira V, Valente H, Casal S. Blood antioxidant and oxidative stress biomarkers acute responses to a 1000-m kayak sprint in elite male kayakers. *J Sports Med physical fitness* 2013; 53(1):71–79.
- [49]. Boisseau p, Loubaton B. Nanomedicine, nanotechnology in medicine. *C. R. Physique J*. 2011; 620-630
- [50]. Arrigoni O, De Tullio MC. Ascorbic acid: much more than just an antioxidant. *Biochim Biophys Acta*, 2002; 1569: 1-9.
- [51]. Chan-Yeung M, Ferreira P, Frohlich J, Schulzer M, Tan F. The effects of age, smoking, and alcohol on routine laboratory tests. *Am. J. Clin. Pathol*. 1981; 75(3): 320-326.
- [52]. Cheung BM, Ong K L, Wong L Y. Elevated serum alkaline phosphatase and peripheral arterial disease in the United States National Health and Nutrition Examination Survey 1999–2004. *Int. J. Cardiol*. 2009; 135(2): 156-161.
- [53]. Wannamethee S G, Lowe G D, Shaper A G, Rumley A, Lennon L, Whincup P. Associations between cigarette smoking, pipe/cigar smoking, and smoking cessation, and haemostatic and inflammatory markers for cardiovascular disease. *Eur. Heart J*. 2005; 26(17):1765-1773.
- [54]. Friedman L S, Martin P, Munoz S J. Liver function tests and the objective evaluation of the patient with liver disease, In: Zakin D, boyer TD, eds. *Hepatology: A text book of liver disease* (Vol. 41). Philadelphia: PA: WB saunders(1996).
- [55]. Iqbal M, Okazaki Y, and Okada S. "In vitro curcumin modulates ferric nitrilotriacetate (Fe-NTA) and hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>)-induced peroxidation of microsomal membrane lipids and DNA damage," *Teratogenesis Carcinogenesis and Mutagenesis*, 2003; 23, 151–160.
- [56]. Antunes L M G, Araújo M C P, Darin J D C, and Bianchi M D LP. "Effects of the antioxidants curcumin and vitamin C on cisplatin-induced clastogenesis in Wistar rat bone marrow cells," *Mutation Research*, 2000; 465: 131–137.
- [57]. Priyadarsini K I, Maity D K, Naik G H. et al., "Role of phenolic O-H and methylene hydrogen on the free radical reactions and antioxidant activity of curcumin," *Free Radical Biology and Medicine*, 2003; 35: 475–484.
- [58]. Awasthi S, Pandya U, Singhal S S. et al., "Curcumin-glutathione interactions and the role of human glutathione S-transferase P1-1," *Chemico-Biological Interactions*, 2000; 128: 19–38.
- [59]. Pari L, Prasath A. Efficacy of caffeic acid in preventing nickel induced oxidative damage in liver of rats. *Chem. Biol. Interact*. 2008; 173, 77-83.
- [60]. Santra A, Chowdhury A, Ghatak S, Biswas A, Dhali GK. Arsenic induces apoptosis in mouse liver is mitochondria dependent and is abrogated by N-acetylcysteine. *Toxicol. Appl. Pharmacol*, 2007; 220: 146–155.
- [61]. Chattopadhyay K, Mandal S, Chattopadhyay BD, Ghosh S. Ameliorative effect of sesame lignans on nicotine toxicity in rats. *Food Chem Toxicol* 2010; 48: 3215–3220.
- [62]. Sinha S, Maiti M, Chattopadhyay K, Chattopadhyay BD. Potential amelioration of curcumin against nicotine-induced toxicity of protein malnourished female rats. *J Pharmacol Toxicol*. 2012; 4: 166-180.
- [63]. Masora E J. Lipids and lipid metabolism, *Annu Rev Physiol*, 1977; 39: 301-308.
- [64]. Brunzell JD, Miller N E, Alaupovic P, St. Hilaire R J, Wang C S. Familial chylomicronemia due to a circulation inhibitor of lipoprotein lipase activity, *J Lipid Res*, 1983; 24: 12-17.
- [65]. Kim M, Kim Y. Hypocholesterolemic effects of curcumin via up-regulation of cholesterol 7 $\alpha$ -hydroxylase in rats fed a high fat diet. *Nutr. Res. Pract.*, 2010; 4: 191-195.
- [66]. Hasimun P, Sukandar EY, Adnyana IK, Tjahjono, DH. Synergistic effect of curcuminoid and s-methyl cysteine in regulation of cholesterol homeostasis. *Int. J. Pharmacol.*, 2011; 7: 268-272.
- [67]. Arafa HM. Curcumin attenuates diet induced hypercholesterolemia in rats. *Med. Sci. Monit.*, 2005; 11: 228-234.
- [68]. Asai A, Miyazawa T. Dietary curcuminoids prevent high-fat diet-induced lipid accumulation in rat liver and epididymal adipose tissue. *J. Nutr.*, 2001; 131: 2932-2935.
- [69]. Chung C L, Ting T O, Hui P H, Chau J W. The inhibition of oleic acid induced hepatic lipogenesis and the promotion of lipolysis by caffeic acid via up-regulation of AMP-activated kinase. *2013;55:122-199.*
- [70]. Codrington A M, Hales BF, Robaire B. anti lipid oxidation of caffeic acid. 2007; 10-1095.
- [71]. Lin CC, Yin MC, Hsu CC, Lin MP. Effect of five cysteine-containing compounds on three lipogenic enzymes in Balb/cA mice consuming a high saturated fat diet. *Lipids*. 2004;39:843–848.
- [72]. Krieger MH, Santos KF, Shishido SM, Wanschel AC, Estrela HF, Santos L, De Oliveira MG, Franchini KG, Spadari-Bratfisch RC, Laurindo FR. Antiatherogenic effects of S-nitroso-N-acetylcysteine in hypercholesterolemic LDL receptor knockout mice. *Nitric Oxide*. 2006;14:12–20.
- [73]. Diniz YS, Rocha KK, Souza GA, Galhardi CM, Ebaid GM, Rodrigues HG, Novelli Filho JL, Cicogna AC, Novelli EL. Effects of N-acetylcysteine on sucrose-rich diet-induced

- hyperglycaemia, dyslipidemia and oxidative stress in rats. *Eur J Pharmacol.* 2006;543:151–157.
- [74]. Schaffer JE. Lipotoxicity: when tissues overeat. *Curr Opin Lipidol.* 2003;14:281–287.
- [75]. Brizzi P, Tonolo G, Carusillo F, Malaguarnera M, Maioli M, Musumeci S. Plasma lipid composition and LDL oxidation. *Clin Chem Lab Med.* 2003;41:56–60
- [76]. Lin CC, Yin MC. Effects of cysteine-containing compounds on biosynthesis of triacylglycerol and cholesterol and anti-oxidative protection in liver from mice consuming a high-fat diet. *Br J Nutr.* 2008;99:37–43.
- [77]. Mahapatra, S.K., Das, S, Bhattacharjee, S, Gautam, N, Majumdar S, Roy, S. 2009. *In vitro* nicotine-induced oxidative stress in mice peritoneal macrophages: A dose-dependant approach. *Toxicol. Mech. Methods*, 2009' 19: 100-108.
- [78]. Yildiz D, Ercal N, Armstrong, DW. Nicotine enantiomers and oxidative stress. *Toxicology*, 1998; 130: 155-165.
- [79]. Chattopadhyay K, Chattopadhyay, B. Effect of nicotine on lipid profile, peroxidation and antioxidant enzymes in female rats with restricted dietary protein. *Indian J. Med. Res.*, 2008; 127: 571-576.
- [80]. Grester H.  $\beta$ -Carotene, vitamin E and vitamin C in different stages of carcinogenesis. *Eur. J. Clin. Nutr.*, 1995; 149: 155-168.
- [81]. Sener G, Toklu HZ, Cetinel S.  $\beta$ -Glucan protects against chronic nicotine-induced oxidative damage in rat kidney and bladder. *Environ. Toxicol. Pharmacol.*, 2007; 23: 25-32.
- [82]. Sreekala S, Indira M. Effects of exogenous selenium on nicotine-induced oxidative stress in rats. *Biol. Trace Elem. Res.*, 2009; 130: 62-71.
- [83]. Zava, D.T., Dollbaum CM, Blen M. Estrogen and progestin bioactivity of foods, herbs and spices. *Proc. Soc. Exp. Biol. Med.*, 1998; 217: 369-378.
- [84]. Chimi H, Cillard J, Cillard P, Rahmani M. Peroxyl and hydroxyl radical scavenging activity of some natural phenolic antioxidants. *J. Am. Oil.Chem. Soc.* 1991; 68, 307-312.
- [85]. Gurer H, Ozunes H, Neal R, Spitz DR, Ercal N. Antioxidant effects of N-acetylcysteine and succimer in red blood cells from lead exposed rats. *Toxicol.* 1998;120:181-9.
- [86]. Hemalatha P, Reddy AG, Reddy YR, Shivakumar P. Evaluation of protective effect of N-acetyl cysteine on arsenic-induced hepatotoxicity. *J Nat Sc Biol Med*, 2013; 4:393-5.
- [87]. Sreejayan N, Rao MN. Free radical scavenging activity of curcuminoids. *Arzneimittelforschung.* 1996; 46, 169–171.
- [88]. Ghoneim AI. Effects of curcumin on ethanol-induced hepatocyte necrosis and apoptosis: implication of lipid peroxidation and cytochrome c. *Naunyn Schmiedebergs Arch Pharmacol.* 2009; 379:47–60.
- [89]. Wang WZ, Cheng J, Luo J., Zhuang, S.M. Abrogation of G2/M arrest sensitizes curcumin-resistant hepatoma cells to apoptosis. *FEBS Lett.* 2008; 582:2689–95.
- [90]. Lee Y, Shin D, Kim KH, Hong S, Choi D, Kim YJ, Kwak MK, Jung Y. Caffeic acidphenethyl ester-mediated Nrf2 activation and I $\kappa$ Bkinase inhibition are involved in NF $\kappa$ B inhibitory effect: Structural analysis for NF $\kappa$ B inhibition. *European Journal of Pharmacology*, 2010; 643,21 -28.
- [91]. Jain SK, Kannan K, Lim G, McVie R, Bocchini Jr, JA. Hyperketonemia increases tumor necrosis factor-alpha secretion in cultured U937 monocytes and Type 1 diabetic patients and is apparently mediated by oxidative stress and cAMP deficiency. *Diabetes* 2002; 51, 2287–2293.
- [92]. Ohta MY, Nagai Y, Takamura T, Nohara E, Kobayashi K. Inhibitory effect of troglitazone on TNF-alpha-induced expression of monocyte chemoattractant protein-1 (MCP-1) in human endothelial cells. *Diabetes Res. Clin. Pract.* 2000; 48, 171–176.