

International Journal of Phytomedicine 3 (2011) 09-26 http://www.arjournals.org/index.php/ijpm/index

Review



Natural sources as potential anti-cancer agents: A review Abhishek Bhanot, Rohini Sharma, Malleshappa N. Noolvi*

*Corresponding author:

Malleshappa N. Noolvi

Department of Natural Chemistry, ASBASJSM College of Pharmacy, Bela, Ropar, Punjab (India)-140111 E-mail: <u>mnoolvi@yahoo.co.uk</u> Bhanot <u>abhi82bhanot@gmail.com</u> Mo. No: +91 9417563874 +91 9878371275 Fax: +91 1881 263655.

Abstract

Natural products remain an important source of new drugs, new drug leads and new chemical entities. The plant based drug discovery resulted mainly in the development of anticancer agents including plants (vincristine, vinblastine, etoposide, paclitaxel, camptothecin, topotecan and irinotecan), marine organisms (citarabine, aplidine and dolastatin 10) and micro-organisms (dactinomycin, bleomycin and doxorubicin). Beside this there is numerous agents identified from fruits and vegetables can used in anticancer therapy. The agents include curcumin (turmeric), resveratrol (red grapes, peanuts and berries), genistein (soybean), diallyl sulfide (allium), S-allyl cysteine (allium), allicin (garlic), lycopene (tomato), capsaicin (red chilli), diosgenin (fenugreek), 6-gingerol (ginger), ellagic acid (pomegranate), ursolic acid (apple, pears, prunes), silymarin (milk thistle), anethol (anise, camphor, and fennel), catechins (green tea), eugenol (cloves), indole-3-carbinol (cruciferous vegetables), limonene (citrus fruits), beta carotene (carrots), and dietary fiber. In this review active principle derived from natural products are offering a great opportunity to evaluate not only totally new chemical classes of anticancer agents, but also novel lead compound and potentially relevant mechanisms of action.

Keywords: Cancer, vincristin, vinblastin, fruit, vegetables.

Introduction

Cancer continues to be one of the major causes of death worldwide and only modest progress has been made in reducing the morbidity and mortality of this disease [1]. Cancers may be caused in one of three ways, namely incorrect diet, genetic predisposition, and via the environment. As many as 95% of all cancers are caused by life style and may take as long as 20– 30 years to develop. Current estimates from the American Cancer Society and from the International Union Against Cancer indicate that 12 million cases of cancer were diagnosed last year, with 7 million deaths worldwide; these numbers are expected to double by 2030 (27 million cases with 17 million deaths) [2].

According to a report of World Health Organization, more than 80% of world's populations depend on traditional medicine for their primary health care needs [3,4]. Plants have a long history of use in the treatment of cancer and it is significant that over 60% of currently used anti-cancer agents are come from natural sources [5]. Naturally occurring drugs that are part of the war against cancer include (vincristine, vinca alkaloids vinblastine, vindesine, vinorelbine), taxanes (paclitaxel, docetaxel), podophyllotoxin and its derivative (etoposide, teniposide), camptothecin and its derivatives (topothecan, irinothecan), anthracyclines (doxorubicin, daunorubicin. epirubicin, idarubicin) and others. In fact, half

This work is licensed under a Creative Commons Attribution 3.0 License.

of all anti-cancer drugs approved internationally were either natural products or their derivatives and were developed on the basis of knowledge gained from small molecules or macromolecules that exist in nature [6,7]. In between 2001 and 2005, 23 new drugs derived from natural products were introduced for the treatment of disorders such as bacterial and fungal infections, cancer, diabetes, dyslipidemia, atopic dermatitis, Alzheimer's disease and genetic diseases such as tyrosinaemia and Gaucher disease out of these 4 drugs have been approved as anti cancer agents. The approved anti cancer agents in 2002 doxorubicin, in 2002 estradiol, in 2004 cholorophyll and 1- aspartic acid and taxol nanoparticles in 2005 [8]. Three new drugs also introduced in 2007 originate from microbial sources for the treatment of cancer is marine alkaloid trabectedin, epothilone derivative ixabepilone and temsirolimus [9].

Nature is an attractive source of new therapeutic candidate compounds as a tremendous chemical diversity is found in millions of species of plants. animals. marine organisms and microorganisms as potential anti-cancer agent [10,11]. In this present study the potential anticancer agent from plants. marines. microorganisms and dietary (fruits, vegetables, spices) sources with some recent and advancement in the field of cancer research were discussed.

Plants as source of anti-cancer agents:

The history of plant as source of anti-cancer agents started in earnest in the 1950s with the discovery and development of the vinca alkaloids (vinblastine and vincristine) and the isolation of the cytotoxic podophyllotoxins. Vinca alkaloid was responsible for an increase in the cure rates for Hodgkin's disease and some forms of leukemia [12]. Vincristine inhibits microtubule assembly, inducing tubulin selfassociation into coiled spiral aggregates [13]. Etoposide is a epipodophyllotoxin, derived from the mandrake plant Podophyllum peltatum and

the wild chervil Podophyllum emodi [14]. It has also significant activity against small-cell lung carcinoma [15]. Etoposide is a topoisomerase II inhibitor, stabilizing enzyme-DNA cleavable complexes leading to DNA breaks [16]. The taxanes paclitaxel and docetaxel has been show antitumor activity against breast, ovarian and other tumor types in the clinic trial. Paclitaxel stabilizes microtubules and leading to mitotic arrest [17]. In addition, the camptothecin derivatives irinotecan and topotecan, have shown significant antitumor activity against colorectal and ovarian cancer respectively [18,19]. These compounds were initially obtained from the bark and wood of Nyssacea Camptotheca accuminata and act by inhibiting topoisomerase I [20]. The taxanes and the camptothecins are presently approved for human use in various countries (Table 1).

Table 1: Plant based anticancer agents in clinicalpractice.

| S.No. | Compound | Uses | Status |
|-------|-------------|--|-----------|
| 1. | Vincristine | Leukemia, lymphoma, | Phase |
| | | breast, lung, pediatric solid cancers and others | III/IV |
| 2. | Vinblastine | Breast, lymphoma, | Phase |
| | | germ-cell and renal | III/IV |
| | | cancer | |
| 3. | Paclitaxel | Ovary, breast, lung, | Phase |
| | | bladder and head and neck cancer | III/IV |
| 4. | Docetaxel | Breast and lung cancer | Phase III |
| 5. | Topotecan | Ovarian, lung and | Phase |
| | - | pediatric cancer | II/III |
| 6. | Irinotecan | Colorectal and lung | Phase |
| | | cancer | II/III |

Rohitukine the plant alkaloid, isolated from the leaves and stems of Dysoxylum binectariferum (Maliaceae) [21,22]. Synthetic flavone derived from rohitukine, Flavopiridol representing the first cyclin-dependent kinase inhibitor to enter the clinical trial [23]. The mechanism of action involves interfering with the phosphorylation of cyclin-dependent kinases and arrest cell-cycle progression at growth phase G1 or G2 [24,25]. Homoharringtonine an alkaloid isolated from the Chinese tree Cephalotaxus harringtonia (Cephalotaxacea) [26]. The mechanism of action is the inhibition of protein synthesis and blocking cell-cycle progression [27]. It has shown efficacy against various leukemias [28]. A lung-cancer-specific antineoplastic agent 4-Ipomeanol is isolated from the sweet potato Ipomoea batata (Convolvulaceae) [29]. The mechanism of action is converted into DNAbinding metabolites upon metabolic activation by cytochrome P450 enzymes that are present in cells of the lung [30]. DNA topoisomerase I inhibitor β -lapachone, that induces cell-cycle delay at G1 or S (synthesis) phase before inducing either apoptotic or necrotic cell death in a variety of human carcinoma cells, including ovary, colon, lung, prostate and breast [31].

Beside this there are so many plants which are used in cancer; following enlist the plant which prevent and target for future studies as potential anticancer agent (Table 2):

| | Diant Spacing | Family | Dlant Dant | Deference |
|-----|---|-----------------|--------------------------|------------|
| - | Plant Species | Family | Plant Part | References |
| 1. | Salvia officinalis | Labiatae | Leaves | [32] |
| 2. | Viscum album | Loranthaceae | Leaves | [33] |
| 3. | Combretum caffrum | Combretaceae | Bark | [34] |
| 4. | Melaleuca alternifolia | Myrtaceae | Leaves | [35] |
| 5. | Lavandula angustifolia | Labiatae | Leaves | [35] |
| 6. | Aglaia foveolata | Meliaceae | Fruit | [36] |
| 7. | Maytenus serrata | Celastraceae | Seed | [37] |
| 8. | Tabebuia impetiginosa | Bignoniaceae | Stem bark and trunk wood | [38,39] |
| 9. | Tabebuia rosea | Bignoniaceae | Stem bark and trunk wood | [38,39] |
| 10. | Tabebuia serratifolia | Bignoniaceae | Stem bark and trunk wood | [38,39] |
| 11. | Dipteryx odorata | Fabaceae | Seed | [40] |
| 12. | Thapsia garganica | Apiaceae | Fruit | [41] |
| 13. | Indigofera tinctoria | Leguminosae | Aerial part | [42] |
| 14. | Matricaria chamomilla | Asteraceae | Flower | [43] |
| 15. | Erythroxylum pervillei | Erythroxylaceae | Root | [44] |
| 16. | Broussonetia papyrifera | Urticaceae | Entire | [45] |
| 17. | Cyclopia intermedia | Fabaceae | Leaves | [46] |
| 18. | Scutellariae radix, Scutellariae indica | Labiatae | Root | [47] |
| 19. | Physalis philadelphica | Solanaceae | Seed | [48] |
| 20. | Dysoxylum binectariferum | Meliaceae | Stem bark | [49] |
| 21. | Aristotelia chilensis | Elaeocarpaceae | Leaf and Stem | [50] |
| 22. | Cyathostemma argentium | Annonaceae | Root | [51] |
| 23. | Epimedium hunanense | Berberidaceae | Aerial parts | [52] |
| 24. | Croton urucurama | Euphorbiacaeae | Bark | [53] |
| 25. | Epilobium hirsutum | Onagraceae | Entire | [54] |
| 26. | Pleione bulbocodioides | Orchidaceae | Tuber | [55] |
| 27. | Cassia quinquangulata | Caesalpiniaceae | Root | [56] |
| 28. | Begonia glabra | Begoniaceae | Entire | [57] |
| 29. | Celastrus orbiculatus | Celastraceae | Entire | [57] |
| 30. | Croton draco | Euphorbiacaeae | Aerial parts | [57] |
| 31. | Smilax sieboldii | Liliaceae | Entire | [58] |
| 32. | Ximenia Americana | Olacaceae | Root | [58] |

| Table 2: | Plants | used as | s anti-cancer. |
|----------|--------|---------|----------------|
|----------|--------|---------|----------------|

| 33. | Maytenus emarginata | Celastraceae | Entire | [59] |
|-----------------|--|-------------------------------|-----------------------|--------------|
| 34. | Sarcandra glabra | Choranthaceae | Entire | [60] |
| 35. | Salvia plebeian | Labiatae | Aerial | [61] |
| 36. | Scutellaria barbata | Labiatae | Entire | [62] |
| 37. | Ocotea caparrapi | Lauraceae | Essential oil | [63] |
| 38. | Caragana cuneata | Leguminosae | Leaf | [64] |
| 39. | Croton flavens | Euphorbiacaeae | Leaf | [65] |
| 40. | Euphorbia heterophylla | Euphorbiacaeae | Stem | [65] |
| 41. | Echites vucatanensis | Apocynaceae | Latex | [65] |
| 42. | Thevetia ahouia | Apocynaceae | Leaf and Stem | [65] |
| 43 | Thevetia gaumeri | Apocynaceae | Leaf and Stem | [65] |
| 44. | Thevetia peruciana | Apocynaceae | Leaf and Stem | [65] |
| 45. | Euphorbia ebracteolata | Euphorbiacaeae | Aerial parts | [66] |
| 46. | Dioscorea collettii | Dioscoreaceae | Rhizome | [67] |
| 47. | Juglans mandshurica | Juglandaceae | Root | [68] |
| 48. | Maackia tenuifolia | Leguminosae | Root | [69] |
| 49 | Juncus acutus | Juncaceae | Leaf | [09] [70] |
| 5 0. | Hedyotis chrysotricha | Rubiaceae | Entire | [70] |
| 50. 51. | Arisaema erubescens | Araceae | Root | [71] |
| 52. | Leptadenia hastate | Asclepiadaceae | Bark | [72] |
| 52. 53. | Viscum calcaratum | Loranthaceae | Entire | [73] |
| 55. 54 | Aphanamixis polystachya | Meliaceae | Stembark | [74] |
| 55. | Pratia nummularia | | Entire | |
| 55. 56. | Aeonium arboretum | Campanulaceae Crassulaceae | Leaf | [76] |
| 50. 57. | Ocotea foetens | Lauraceae | Branchlets | [77] |
| 57. 58. | | Celastraceae | | [77] |
| 58. 59. | Maytenus canariensis Sedum alboroseum | Crassulaceae | Fruit juice Entire | [78] |
| 59. 60. | | | Entire | [79] |
| 60. 61. | Euphorbia micractina | Euphorbiacaeae | | [80] |
| 61. 62. | Euphorbia prolifera | Euphorbiacaeae | Latex | [81] |
| 63. | Scirpus holoschoenus | Cyperaceae Dilleniaceae | Inflorescence | [82] |
| 63. 64 | Dillenia suffruticosa | | Fruit | [83] |
| | Hypoxis rooperii | Hypoxiaceae | Tuber | [84] |
| 65. | Inula linariaefolia | Compositae | Flowers | [85] |
| 66. 67 | Ziziphus mauritiana | Rhamnaceae | Stem bark and Fruit | [86] |
| 67. | Adiantum macrophyllum | Pteridaceae | Entire | [87] |
| 68. | Thalictrum fabri | Ranunculaceae | Root | [88] |
| 69. 70 | Scutellaria indica | Labiatae | Root | [89] |
| 70. | Hypericum japonicum | Guttiferae | Entire | [90] |
| 71. | Cyathea fauriei | Cyatheaceae | Shoot | [91] |
| 72. | Fissistigma oldhamii | Annonaceae | Stem | [92] |
| 73. | Monnina obtusifolia | Polygalaceae | Aerial parts | [93] |
| 74. | Coriolus versicolor | Polyporaceae | Fruitbody | [94] |
| 75. 76 | Melastoma malabathricum | Melatomataceae | Flower | [95] |
| 76. | Carapa guianensis | Meliaceae | Seed oil | [96] |
| 77. 79 | Swietenia humilis | Meliaceae | Seed | [97] |
| 78. | Ficus pretoiae | Moraceae | Sap | [98] |

| 79. | Croton lechleri | Euphorbiacaeae | Latex | [99] |
|------|----------------------------|-----------------|-----------|-------|
| 80. | Aster amellus | Compositae | Entire | [100] |
| 81. | Crassocephalum bojeri | Compositae | Entire | [101] |
| 82. | Echinops grijisii | Compositae | Root | [101] |
| 83. | Adenium obesum | Apocynaceae | Leaf | [102] |
| 84. | Ipomea batata | Convolvulaceae | Rhizome | [103] |
| 85. | Uncaria tomentosa | Rubiaceae | Bark | [104] |
| 86. | Plantago asiatica | Plantaginaceae | Leaf | [105] |
| 87. | Phymatosorus diversifolium | Polydiaceae | Root | [105] |
| 88. | Rabdosia rubescens | Labiatae | Leaf | [106] |
| 89. | Salvia chinensis | Labiatae | Entire | [107] |
| 90. | Ganoderma lucidum | Ganodermataceae | Fruitbody | [108] |
| 91. | Euphorbia kansui | Euphorbiacaeae | Root | [109] |
| 92. | Echinops latifolius | Compositae | Root | [110] |
| 93. | Euphorbia marginata | Euphorbiacaeae | Entire | [111] |
| 94. | Ligustrum lucidum | Oleaceae | Seed | [112] |
| 95. | Phytolacca esculenta | Phytolaccaceae | Root | [113] |
| 96. | Pinus parviflora | Pinaceae | Strobilus | [114] |
| 97. | Dysosma pleiantha | Berberidaceae | Root | [115] |
| 98. | Alnus japonica | Betulaceae | Wood | [116] |
| 99. | Ruellia tuberose | Acanthaceae | Bark | [117] |
| 100. | Acacia xanthophloea | Leguminosae | Fruit | [118] |
| 101. | Lannea stuhlmannii | Anacardiaceae | Root | [118] |
| 102. | Maytenus obscura | Celastraceae | Leaf | [118] |
| 103. | Plicosepalus sagittifolius | Loranthaceae | Branches | [118] |
| 104. | Piper latifolium | Piperaceae | Leaf | [119] |
| 105. | Morinda citrifolia | Rubiaceae | Root | [119] |
| 106. | Knema tenuinervia | Myristicaceae | Stembark | [120] |
| 107. | Deeringia amaranthoides | Amaranthaceae | Fruit | [121] |
| 108. | Cynanchum hancoekianum | Asclepiadaceae | Entire | [122] |
| 109. | Azadirachta indica | Meliaceae | Leaf | [123] |
| 110. | Virola bicuhyba | Myristicaceae | Seed | [124] |
| 111. | Sempervivum armenum | Crassulaceae | Leaf | [125] |
| 112. | Sempervivum arvense | Crassulaceae | Leaf | [125] |
| 113. | Hippophae salicifolia | Elaeagnaceae | Fruit | [126] |
| 114. | Hypoxis nyasica | Hypoxiaceae | Rhizome | [127] |
| 115. | Astragalus membranaceus | Leguminosae | Root | [128] |
| 116. | Maytenus macrocarpa | Celastraceae | Stembark | [129] |
| 117. | Cephalotaxus Harrington | Cephlotaxaceae | Entire | [130] |

Dietary source of anti cancer agents:

Natural dietary agents including fruits, vegetables, and spices have drawn a great deal of attention from both the scientific community and the general public owing to their demonstrated ability to suppress cancers. Recent

studies suggest that the consumption of food rich in fruits, vegetables and spices have a lower incidence of cancers (stomach, esophagus, lung, oral cavity and pharynx, endometrium, pancreas and colon) [131-133]. Dietary agents consist of a wide variety of biologically active components that are responsible for the anti-cancer effects like curcumin, genistein, resveratrol, diallyl sulfide, S-allyl cysteine, allicin, lycopene, capsaicin, diosgenin, gingerol, ellagic acid, ursolic acid, silymarin, anethol, catechins, eugenol, isothiocyanates, isoeugenol, dithiolthiones, indole-3-carbinol, isoflavones, saponins, phytosterols, inositol hexaphosphate, Vitamin C, D-limonene, lutein, folic acid, beta carotene,

selenium, Vitamin E and flavonoids (Table 3). Many of which have been used in traditional medicines for thousands of years. These dietary agents are believed to suppress the inflammatory processes that lead to transformation, hyperproliferation, and initiation of carcinogenesis. Their inhibitory influences may ultimately suppress the final steps of carcinogenesis i.e angiogenesis and metastasis [134].

| S. No. | Botanical Name | Source | Compound | Reference |
|--------|---|--------------|-------------------|-----------|
| 1 | Carica papaya, Family- Caricaceae | Berries | β-Cryptoxanthin | [135] |
| 2 | Glycyrrhiza glabra; Glycyrrhiza radix; Licorice roo | | Glycyrrhizin | [136] |
| | Glycyrrhiza uralensis, Family- Leguminosae | | | |
| 3 | Cannabis sativa, Family- Cannabiaceae | Hemp | Cannabinol | [137] |
| 4 | Rosmarinus officinalis, Family- Lamiaceae | Rosemary | Carnosol | [138] |
| 5 | Pueraria lobata radix, Family- Fabaceae | | Genistein | [139] |
| 6 | Glycine max, Family- Fabaceae | Soybeans | Genistein | [139] |
| 7 | Prunus armeniaca, Family- Rosaceae | Apricots | Carotenoids | [140] |
| 8 | Zingiber officinale, Family- Zingiberaceae | Tuber | Gingerol | [141] |
| 9 | Lycopersicon esculentum, Family- Solanaceae | Tomato | Lycopene, Lutein, | [141] |
| | | | Kaempferol | |
| 10 | Piper nigrum; Piper longum, Family- Piperaceae | Black pepper | Purpurogallin; | [142] |
| | | | Piperine | |
| 11 | Ocimum sanctum, Family-Lamiaceae | Basil | Ursolic acid | [143] |
| 12 | Betula alba, Family- Betulaceae | Birch tree | Betulinic acid | [144] |
| 13 | Crocus sativus, Family- Iridaceae | Saffron | Carotenoids | [146] |
| 14 | Silymarin marianum, Family-Asteraceae | Milk thistle | Silymarin | [147] |
| 15 | Capsaicum annum; Capsaicum frutens, Family- | Red chilli | Capsaicinoids, | [148] |
| | Solanaceae | | Capsaicin | |
| 16 | Camellia sinensis, Family- Theaceae | Green and | Catechin and | [149] |
| | | black teas | theaflavins | |
| 17 | Vitis vinifera, Family- Vitaceae | Grapes | Resveratrol | [150] |
| 18 | Daucus carota sativus, Family- | Carrot | β-Carotene | [151] |
| | Apiaceae/umbelliferae | | | |
| 19 | Tabebuia avellanedae, Family- Bignoniaceae | Lapacha tree | Lapachone | [31] |
| 20 | Citrus aurantium, Family- Rutaceae | Orange | Hesperidin | [152] |
| 21 | Prunus dulcis, Family- Rosaceae | Almond | Morin | [153,154] |
| 22 | Aloe arborescens, Family- Asphodelaceae | Aloe vera | Emodin | [155] |
| 23 | Opium poppy, Family- Paparveraceae | Рорру | Morphine and its | [157] |
| | | | analogues | |
| 24 | Curcurbita moschata, Family-Cucurbitaceae | Pumpkin | β-Carotene | [158] |
| 25 | Azadirachata indica, Family- Meliaceae | Neem | Polyphenolics | [159] |

Table 3: Dietary sources as anticancer agent.

Marines as source of anti-cancer agents:

Marine organisms are a rich source for natural products [160]. In recent time, advancement in deep-sea collection and aqua culture technology gives significant number of compounds derived from marine organisms entering preclinical and early clinical evaluation as potential anticancer agent [161,162]. Overall, more than 3000 new substances have been identified from marine organisms that demonstrate the great potential as a source of novel chemical classes [163]. Marine belongs to very diverse structural classes including polyketides, terpenes, steroids and peptides. The organisms yielding these bioactive marine compounds include invertebrate animals, algae, fungi and bacteria [164].

The first anticancer product didemnin B, a cyclic depsipeptide isolated from the tunicate Trididemnum solidum from marine source enter in clinical trials. Preliminary results showed a against non-Hodgkin's partial activity lymphoma [165]. It can inhibit protein synthesis and arrest G1 phase of cell-cycle. Another depsipeptide Aplidine appear to be more active as comparison with didemninB in preclinical trial and does not produce life-threatening neuromuscular toxicity. Preclinical data indicate that aplidine is active against several tumors through blockade of cell-cycle progression at phase [166]. There are number of G1 ecteinascidins have been isolated from the marine source tunicate Ecteinascidia turbinata. One of these ecteinascidins (ET-743) was selected for clinical trials and antitumor effects have been observed in phase I studies [167]. ET-743 is a tetrahydroisoquinilone alkaloid and they acts by selective alkylation of guanine residues in the DNA minor groove [168] and also interacts with nuclear proteins [169]. In Europe and the United States ET-743 is currently in phase II clinical trials [167]. The dolastatins are a class of peptides obtained from the Indian Ocean, Dolabella auricularia. These peptides have cytotoxic activity and now a day.

dolastatin10 and dolastatin15 of this class have received the greatest clinical interest Dolastatin10 has entered in Phase I and Phase II clinical trials. after showing significant antitumor activity in preclinical models [170]. Its mechanism of action involves inhibition of microtubule assembly ultimately result in cellcycle arrest in metaphase [171,172]. The bryostatins, 20 macrocyclic lactones isolated from Bugula neritina and other marine bryozoa. These macrocyclic compounds have shown activity lymphocytic significant against leukemia cell line [173]. Bryostatin1 has recently entered phase II clinical trials for the of melanoma, non-Hodgkin's treatment lymphoma, renal cancer and colorectal cancer [174-176] and continues to be evaluated in phase I clinical trials. Bryostatin1 has been found to promote the normal growth of bone marrow progenitor cells, to provide in vivo protection against normally lethal doses of ionizing radiation and to serve as an immune stimulant, enhancing the normal production of interleukin2 and interferons [177].

Beside this there are the number of compounds isolated from marine as potential anti-cancer agents included in Table 4 [178,179].

Microorganisms as source of anticancer agents:

Antitumor antibiotics are among the most important cancer chemotherapeutic agents, and members of the anthracycline, include actinomycin. mitomycin bleomycin, and aureolic acid families [6]. Clinically useful agents from these above families are the daunomycin and related agents like doxorubicin, idarubicin and epirubicin; the peptolides (exemplified by dactinomycin), the mitosanes (such as mitomycin C) and the glycosylated anthracenone mithramycin. The anthracyclines are among the most used antitumor antibiotics in the clinic and exert antitumor activity mainly by inhibiting topoisomerase II [180,181].

Table 4: Marine derived potential anticancer agent.

| S.No. | Compound | Organism | Chemistry | Mechanism of action |
|-------|-------------------|--------------|-----------------|--|
| 1. | Aaptamine | Sponge | Alkaloid | Induction of p21 and G2/M cell cycle |
| | | | | arrest |
| 2. | Cortistatin A | Sponge | Alkaloid | Selective inhibiton of angiogensis |
| 3. | Aplidine | Ascidian | Depsipeptide | Oxidation and inactivation of low |
| | | | | molecular weight-protein tyrosine |
| | | | | phosphatase activity |
| 4. | Bastadine 6 | Sponge | Alkaloid | Inhibition of angiogenesis in vitro and |
| | | | | in vivo involves apoptosis |
| 5. | Fucoxanthinol | Ascidian | Carotenoid | Induction of apoptosis |
| 6. | Lamellarin D | Mollusk | Alkaloid | ErbB3 protein and PI3K- Akt pathway |
| | | | | involved in necrosis induction |
| 7. | Clavulone II | Soft coral | Prostanoid | G1 cell cycle arrest and apoptosis |
| 8. | Geodiamolides | Sponge | Peptide | Disorganization of actin filaments |
| 9. | Ircinin-1 | Sponge | Sesterterpene | G1 phase inhibition and apoptosis |
| | | | | induction |
| 10. | Laxaphycins A and | Bacterium | Cyclic peptides | Increased polyploidy by putative |
| | В | | | topoisomerase II alterations |
| 11. | Leptosins C and F | Fungus | Alkaloid | DNA topoisomerase I and II inhibition |
| | | | | and apoptosis induction |
| 12. | Onnamide A | Sponge | Polyketide | Protein synthesis inhibition |
| 13. | Philinopside A | Sea cucumber | Saponin | Inhibition of angiogenesis and |
| | | | | receptor tyrosine kinases |
| 14. | Variolin B | Sponge | Alkaloid | Inhibition of cyclin-dependent kinases |
| | | | | and apoptosis induction |
| 15. | Aplidine | Ascidian | Depsipeptide | Induction of apoptosis with |
| | | | | concomitant G1 arrest and G2 |
| | | | | blockage |
| 16. | Ascididemin | Ascidian | Alkaloid | Direct iminoquinone reduction and |
| | | | | reactive oxygen species generation |
| 17. | Cammbrescidin | Sponge | Alkaloid | Induction of eythroid differentiation |
| | 800 | | | and cell cycle arrest |
| 18. | Dideoxypetrosynol | Sponge | Fatty acid | Induction of apoptosis via |
| | А | | | mitochondrial signaling pathway |
| 19. | Dolastatin 10 | Mollusc | Peptide | Binds to amino-terminal peptide of β - |
| | | | | tubulin containing cysteine |
| 20. | Girolline | Sponge | Alkaloid | Induction of G2/M cell cycle arrest |
| | | | | and p53 proteasome recruitment |
| 21. | Halichondrin B | Sponge | Macrolide | Induction of mitotic blockage and |
| | analogues | | derivative | apoptosis |
| 22. | Lissoclinolide | Ascidian | Fatty acid | G2/M cell cycle arrest |
| 23. | Neoamphimedine | Sponge | Alkaloid | Induction of topoisomerase II α - |
| 49. | | sponge | AINAIUIU | |
| | | | | mediated catenation of DNA |

| 24. | Psammaplin A | Sponge | Alkaloid | Inhibition of aminopeptidase N and suppression of angiogenesis in vitro |
|-----|--------------------------|----------|--------------------------|---|
| 25. | Alkylpyridinium | Sponge | Alkaloid | Induction of apoptosis and reduced cell adhesion |
| 26. | Aeroplysinin | Sponge | Alkaloid | Induction of apoptosis on proliferating endothelial cells |
| 27. | Bryostatin-1 | Bryozoan | Macrolide | Potentiation of ara-C induced apoptosis by PKC-dependent release of TNF-α |
| 28. | Cephaiostatin | Worm | Steroid | Apoptosis and increased mitochondrial matrix density |
| 29. | Chondropsin A | Sponge | Macrolide | In Vitro inhibition of V-ATPase enzyme |
| 30. | Dehydrothrysiferol | Alga | Triterpene | Enhanced apoptosis induction in estrogen receptor negative breast cancer cells |
| 31. | Diazonamide-A | Ascidian | Peptide | Disruption of mitosis and cellular microtubules with inhibition of GTP hydrolysis |
| 32. | Dictyostatin | Sponge | Polyketide | Induction of tubulin polymerization |
| 33. | Dolastatin 11 | Mollusc | Peptide | F-actin stabilization by connection between two long-pitch strands |
| 34. | Ecteinascidin- 743 | Ascidian | Isoquinoline alkaloid | Telomere dysfunction increases susceptibility to ET-743 |
| 35. | GA3 polysaccharide | Alga | Polysaccharide | Inhibition of topoisomerase I and II |
| 36. | Hemiasterlin analogue | Sponge | Tripeptide | Induction of microtubule depolymerisation |
| 37. | Kahalalide F | Mollusc | Depsipeptide | Potent cytotoxicity and induction of necrosis |
| 38. | Lamellarin D | Mollusc | Alkaloid | Potent inhibition of topoisomerase I |
| 39. | omega-3 fatty acids | Fish | Fatty acid | |

Many pharmaceutical agents have been discovered by screening natural products from a wide range of microorganisms. Rapamycin and its analogs are products of Streptomyces hygroscopicus have potent immunosuppressive activity. They inhibit signaling pathways required for T-cell activation and proliferation. Rapamycin blocks progression of the cell cycle at middle-to-late G1 phase in T cells and B cells, and osteosarcoma and rhabdomyosarcoma cell lines, among others [182]. Geldanamycin is a benzoquinone ansamycin natural fermentation product and inhibits heat-shock protein HSP 90 [183].

| S.No. | Compound | Microorganism | Used in Cancer |
|-------|--------------|------------------------------|-------------------------------------|
| 1. | Actinomycin | Streptomyces spp. | Sarcoma and germ-cell tumors |
| 2. | Bleomycin | Streptomyces verticillus | Germ-cell, cervix and head and neck |
| | | | cancer |
| 3. | Daunomycin | Streptomyces coeruleorubidus | Leukemia |
| 4. | Doxorubicin | Streptomyces Pneuceticus | Lymphoma, breast, ovary, lung and |
| | | | sarcomas |
| 5. | Epirubicin | Streptomyces pneuceticus | Breast cancer |
| 6. | Idarubicin | Streptomyces Pneuceticus | Breast cancer and leukemia |
| 7. | Mitomycin C | Streptomyces caespitosus | Gastric , colorectal, anal and lung |
| | | | cancer |
| 8. | Geldanamycin | Streptomyces Hygroscopicus | Experimental |
| 9. | Rapamicin | Streptomyces hygroscopicus | Experimental |
| 10. | Wortamannin | Talaromyces wortmanni | Experimental |

Table 5: Microorganism derived anti-cancer agents.

Wortmannin is a product of the fungus Talaromyces wortmanni and inhibits signal transduction pathways by forming a covalent complex with an active-site residue of phosphoinositide 3 kinase (PI3K), inhibiting PI3K activity [184] (Table 5). Thus, toxins that evolved originally to kill competing micoorganisms can have а variety of physiological effects in animals. In many cases, the targets of these compounds are components of signal transduction cascades that are conserved in many species, and that have been considered novel targets for anticancer drug discovery [185].

Conclusion:

Natural products have been a prime source for the treatment of many forms of cancer, many of which are consumed daily with the diet. They provide significant protection against various cancers and many other diseases. The antioxidant medicinal plants and their products prevent from the cancer and other diseases by protecting cells from damage. Thus, consuming a diet rich in antioxidant fruits, vegetables, herbs etc. will provide health-protective effects. Microbes and marine organisms also have been offering the great role in the prevention and treatment of cancer. All the natural products discussed in this review exhibit anticancer activities. Natural products offer a great opportunity to evaluate not only totally new chemical classes of anticancer agents, but also novel and potentially relevant mechanisms of action.

References

- Hail Jr N. Mitochondria: a novel target for the chemoprevention of cancer. Apoptosis. 2005;10:687–705.
- 2. Aggarwal BB, Danda D, Gupta S, Gehlot P. Models for prevention and treatment of cancer: Problems vs promises. Biochem Pharmacol. 2009;78:1083–1094.
- 3. Farnsworth NR. Ethnobotany and the Search for New Drugs, Wiley Willett WC. Diet and health: what should we eat. Science. 1994;264(5158):532–7.
- 4. Duraipandiyan V, Ayyanar M, Ignacimuthu S. Antimicrobial activity of some ethnomedicinal plants used by Paliyar tribe from Tamil Nadu, India. BMC Comp Alter Med. 2006;6:35-41.
- Cragg GM, Kingston DGI, Newman DJ. Anticancer Agents from Natural Products. Taylor & Francis Group, Boca Raton, FL. Brunner-Routledge Psychology Press; 2005.
- 6. Cragg GM, Newman D J, Weiss RB. Coral reefs, forests, and thermal vents: the worldwide exploration of nature for novel antitumor agents. Semin Oncol. 1997;24:156-163.

- 7. Newman DJ, Cragg GM. Natural products as sources of new drugs over the last 25 years. J Nat Prod. 2007;70:461–477.
- 8. Butler MS. Natural products to drugs: natural product derived compounds in clinical trials. Nat Prod Rep. 2005;22:162–195.
- 9. Bailly C. Ready for a comeback of natural products in oncology. Biochem Pharmacol. 2009;77:1447–1457.
- Newman DJ, Cragg GM, Snader KM. Natural products as sources of new drugs over the period 1981–2002. J Nat Prod. 2003;66: 1022–1037.
- 11. Butler MS. The role of natural product chemistry in drug discovery. J Nat Prod. 2004;67:2141–2153.
- 12. DeVita VT Jr, Serpick AA, Carbone PO. Combination chemotherapy in the treatment of advanced Hodgkin's disease. Ann Intern Med. 1970;73:881-895.
- 13. Noble RL. The discovery of the vinca alkaloids chemotherapeutic agents against cancer. Biochem Cell Biol. 1990;68:1344-1351.
- 14. Stähelin H. Actvity of a new glycosidic lignan derivative (VP-16-213) related to podophyllotoxin in experimental tumors. Eur J Cancer. 1973;9:215-221.
- 15. Harvey AL. Medicines from nature: are natural products still relevant to drug discovery. Trends Pharmacol Sci. 1999;20:196-198.
- Liu LF. DNA topoisomerase poisons as antitumor drugs. Annu Rev Biochem. 1989;58:351-375.
- 17. Wani MC, Taylor HL, Wall ME, et al. Plant antitumor agents. VI. The isolation and structure of taxol, a novel antileukemic and antitumor agent from Taxus brevifolia. J Am Chem Soc. 1971;93:2325-2327.
- 18. Creemers GJ, Bolis G, Gore M, et al. Topotecan, an active drug in the second-line treatment of epithelial ovarian cancer. J Clin Oncol. 1996;14:3056-3061.
- 19. Bertino JR. Irinotecan for colorectal cancer. Semin Oncol. 1997;24:S18-S23.
- 20. Liu LF, Desai SD, Li TK, et al. Mechanism of action of camptothecin. Ann New York Acad Sci. 2000;922:1-10.
- 21. Harmon AD, Weiss U, Silverton JV. The structure of rohutukine, the main alkaloid of

Amoora rohituka (syn. Aphanamixis polystachya) (Maliaceae). Tetrahydron. 1979;20:721-724.

- 22. Cragg G, Suffness M. Metabolism of plantderived anticancer agents. Pharmacol Ther. 1988;37:425-432.
- 23. Losiewicz MD, Carlson BA, Kaur G, et al. Potent inhibition of cdc2 kinase activity by the flavonoid L86-8275. Biochem Biophys Res Commun. 1994;201:589-595.
- 24. Worland PJ, Kaur G, Stetler-Stevenson M, et al. Alteration of the phosphorylation state of p32cdc2 kinase by the flavone L86-8275 in breast carcinoma cells. Biochem Pharmacol. 1993;46:1831-1836.
- 25. Kelland LR. Flavopiridol, the first cyclindependent kinase inhibitor to enter the clinic: current status. Ex Opin Inv Drugs. 2000;9:2903-2911.
- 26. Powell RG, Weisleder D, Smith CR Jr, et al. Structures of harringtonine, isoharringtonine, and homoharringtonine. Tetrahydron Lett. 1970;11:815-818.
- 27. Zhou DC, Zittoun R, Marie JP. Homoharringtonine: an effective new natural product in cancer chemotherapy. Bull Cancer. 1995;82:987-995.
- 28. Kantarjian HM, O'Brien S, Anderlini P, et al. Treatment of myelogenous leukemia: current status and investigational options. Blood. 1996;87:3069-3081.
- 29. Rowinsky EK, Noe DA, Ettinger DS, et al. PhaseI and pharmacological study of the pulmonary cytotoxin 4-ipomeanol on a single dose schedule in lung cancer patients: hepatotoxicity is dose limiting in humans. Cancer Res. 1993;53:1794-1801.
- 30. Rehm S, Devor DE. Acute effects of 4ipomeanol on experimental lung tumors with bronchiolar or alveolar cell features in Syrian hamsters or C3H/HeNCr mice. J Cancer Res Clin Oncol. 1993;120:41-50.
- 31. Li YZ, Li CJ, Pinto AV, et al. Release of mitochondrial cytochrome c in both apoptosis and necrosis induced by β -lapachone in human carcinoma cells. Mol Med. 1999;4:232-239.
- 32. Vukovic-Gacic B, Nikcevic S, Beric-Bjedov T, et al. Antimutagenic effect of essential oil of sage (Salvia officinalis L.) and its monoterpenes against UV-induced mutations

in Escherichia coli and Saccharomyces cerevisiae. Food Chem Toxicol. 2006;44:1730–1738.

- ÖnayUçar E, Karagöz A, Arda N. Antioxidant activity of Viscum album sp. Fitoterapia. 2006;77: 556–560.
- 34. Pinney KG, Jelinek C, Edvardsen K, et al. The discovery and development of the combretastatins. In: Cragg GM, Kingston DGI, Newman DJ. Anticancer Agents from Natural Products. Taylor & Francis Group, Boca Raton, FL Brunner-Routledge Psychology Press; 2005:23–46 (Chapter 3).
- 35. Evandri MG, Battinelli L, Daniele C, et al. The antimutagenic activity of Lavandula angustifolia (lavender) essential oil in the bacterial reverse mutation assay. Food Chem Toxicol. 2005;43:1381–1387.
- 36. Hwang BY, Su BN, Chai H, et al. Silvestrol and episilvestrol, potential anticancer rocaglate derivatives from Aglaia silvestris. J Org Chem. 2004;69(10):3350-3358.
- 37. Cassady JM, Chan KK, Floss HG, et al. Recent developments in the Maytansanoid antitumor agents. Chem Pharma Bulletin. 2004;52:1–26.
- Suffness M, Douros J. Miscellaneous natural products with antitumor activity. In: Cassady JM, Douros JD. Anticancer Agents Based on Natural Product Models. New York, Academic Press; 1980:474 (Chapter 14).
- 39. Ravelo AG, Estevez-Braun A, Chavez-Orellana H, et al. Recent studies on natural products as anticancer agents. Curr Topics Med Chem. 2004;4:241–265.
- 40. Jang DS, Park EJ, Hawthorne ME, et al. Potential cancer chemopreventive constituents of the seeds of Dipteryx odorata (tonka bean). J Nat Prod. 2003;66(5):583– 587.
- 41. Denmeade SR, Jakobsen CM, Janssen S, et al. Prostate-specific antigen-activated thapsigargin prodrug as targeted therapy for prostate cancer. J Natl Cancer Inst. 2003;95:990–1000.
- 42. Newman DJ, Cragg GM, Holbeck S, et al. Natural products as leads to cell cycle pathway targets in cancer chemotherapy. Curr Cancer Drug Targets. 2002;2:279–308.
- 43. Hernandez-Ceruelos A, Madrigal-Bujaidar E, de la Cruz C. Inhibitory effect of chamomile

essential oil on the sister chromatid exchanges induced by daunorubicin and methyl methanesulfonate in mouse bone marrow. Toxicol Lett. 2002;135:103–110.

- 44. Silva GL, Cui B, Chavez D, et al. Modulation of the multidrug-resistance phenotype by new tropane alkaloid aromatic esters from Erythroxylum pervillei. J Nat Prod. 2001;64(12):1514–1520.
- 45. Lee D, Bhat KPL, Fong HHS, et al. Aromatase inhibitors from Broussonetia papyrifera. J Nat Prod. 2001;64(10):1286– 1293.
- 46. Marnewick JL, Gelderblom WCA, Joubert E. An investigation on the antimutagenic properties of South African herbal teas. Mutat Res. 2000;471:157–166.
- 47. Abdulla M, Gruber P. Role of diet modification in cancer prevention. Biofactors. 2000;12:45–51.
- 48. Dinkova-Kostova AT, Talalay P. Persuasive evidence that quinine reductase type 1 (DT diaphorase) protects cells against the toxicity of electrophiles and reactive forms of oxygen. Free Radic Biol Med. 2000;29(3–4): 231–240.
- 49. Sausville EA, Zaharevitz D, Gussio Z, et al. Cyclin-dependent kinases: initial approaches to exploit a novel therapeutic target. Pharmacol Thera. 1999;82:285.
- 50. He K, Valcic S, Timmerman BN, et al. Indole alkaloids from Aristotelia chilensis (Mol.) Stuntz. Int J Pharmacog. 1997;35:215–217.
- 51. Khamis SB, Brown JE, Bibby MC, et al. Evaluation of two plants species used in Malay traditional medicine for the treatment of breast cancer. J Pharma Pharmacol. 1997;49:113.
- 52. Liang HR, Vuorela P, Vuorela H, et al. Isolation and immunomodulatory effect of flavonol glycosides from Epimedium hunanense. Planta Medica. 1997;63:316 – 319.
- Peres MTLP, Monache FD, Cruz AB, et al. Chemical composition and antimicrobial activity of Croton urucurana Baillon (Euphorbiaceae). J Ethnopharmacol. 1997;56:223 – 226.

- 54. Barakat HH, Hussein SAM, Marzour MS, et al. Polyphenolic metabolites of Epilobium hersutum. Phytochemistry. 1997;46:935–941.
- 55. Bai L, Yamaki M, Takagi S. Lignans and a bichroman from Pleione bulbocodioides. Phytochemistry. 1997;44:341–343.
- 56. Jang M, Cai L, Udeani GO, et al. Cancer chemopreventive activity of resveratrol, a natural product derived from grapes. Science. 1997;275(5297):218–220.
- 57. Gupta MP, Monge A, Karikas GA, et al. Screening of Panamanian medicinal plants for brine shrimp toxicity, crown gall tumor inhibition, cytotoxicity and DNA intercalation. Int J Pharmacog. 1996;34:19 – 27.
- Suh HW, Song DK, Son KH, et al. Antinociceptive effect of Smilaxin B administered intracerebroventricularly in the mouse. Planta Med. 1996;62:141 – 145.
- 59. Shirota O, Tamemura T, Morita H, et al. Triterpenes from Brazilian medicinal plant "Chuchuhuasi" (Maytenus krukobii). J Nat Prod. 1996;59:1072–1075.
- 60. Tsui WY, Brown GD. Cycloeudesmanolides from Sarcandra glabra. Phytochemistry. 1996;43:819–821.
- Um SH, Lee KR, Zee OP, Pyo S. The effect of Salvia plebeia on murine macrophagemediated cytotoxicity. Nat Prod Sci. 1996;2: 43 – 47.
- 62. Ducki S, Hadfield JA, Lawrence CY, et al. Isolation of E-1-(4'-hydroxyphenyl)-But-1en-3-one from Scutellaria barbata. Planta Med. 1996;62:185–186.
- Palomino E, Maldonado C, Kempff MB, et al. Caparratriene, an active sesquiterpene hydrocarbon from Ocotea caparrapi. J Nat Prod. 1996;59:77 79.
- 64. Singh VK, Ali ZA, Zaidi STH, et al. Ethnomedicinal uses of plants of Gonda district forests of Uttar Pradesh, India. Fitoterapia. 1996;67:129 – 139.
- 65. Flores JS, Ricalde RV. The secretions and exudates of plants used in mayan traditional medicine. J Herbs Spices Med Plants. 1996;4:53–59.
- 66. Ahn BT, Oh KJ, Ro JS, Lee KS. A new flavonoid from Euphorbia ebracteolata. Planta Med. 1996;62:383–384.

- 67. Hu K, Dong A, Yao XS, et al. Antineoplastic agents I. Three spirostanol glycosides from rhizomes of Dioscorea collettii var. hypoglauca. Planta Med. 1996;62:573 575.
- Joe YK., Son JK, Park SH, et al. New naphthalenyl glucosides from the roots of Juglans mandshurica. J Nat Prod. 1996;59:159 – 160.
- Zeng, JF, Li GL, Zhu DY. Two isoprenoidsubsititued isoflavans from roots of Maackia tenuifolia. Phytochemistry. 1996;43:893 – 896.
- Martinez-Lirola MJ, Gonzalez-Tejero MR, Molero-Mesa J. Ethnobotanical resources in the province of Almeria, Spain, Campos De Nijar. Econ Bot. 1996;50:40 – 56.
- 71. Peng JN, Feng XZ, Liang XT. A new Iridoid from Hedyotis crysotrichae. Chin Chem Lett. 1995;6:965–966.
- 72. Ducki S, Hadfield JA, Lawrence NJ, et al. Isolation of paeonol from Arisaema eubescens. Planta Med. 1995;61:586–587.
- 73. Aquino R, Pizza C, De Tommasi N, et al. New polyoxypregnane ester derivatives from Leptadenia hastata. J Nat Prod. 1995;58:67.
- 74. Tiwari VJ. Ethnobotanical survey of Halbi tribe of Chandrapur and Gadchiroli districts of Maharashtra State, India. Fitoterapia. 1995;66:346 – 350.
- 75. Rabi T, Gupta RC. Antitumor and cytotoxic investigation of Amoora rohituka. Int J Pharmacog. 1995;33:359–361.
- Ho LK, Ou JC, Sun ML, Sun CM. Two rare alkaloids from Pratia nummularia. Planta Med. 1995;61:567–568.
- 77. Rivera D, Obon C. The Ethnopharmacology of Madeira and Porto Santo islands, a review. J Ethnopharmacol. 1995;46:73–93.
- 78. Shirota O, Morita H, Takeya K, Itokawa H. Structures of xuxuarines, stereoisomeric triterpene dimmers from Maytenus chuchuhuasca. Tetrahedron. 1995;51:1107– 1120.
- 79. Fujita T, Sezik E, Tabata M, et al. Traditional medicine in Turkey VII. Folk medicine in middle and west Black sea regions. Econ Bot. 1995;49:406–422.
- Shi JG, Jia ZJ, Cui YX. Novel Tricyclic Diterpenoids from Euphorbia micractina. J Nat Prod. 1995;58:51–56.

- Wu DG, Sorg B, Hecker E. New myrsinolrelated polyfunctional pentacyclic diterpene esters from roots of Euphorbia prolifera. J Nat Prod. 1995;58:408–413.
- Gonzalez-Tejero MR, Molero-Mesa J, Casares-Porcel M, et al. New Contributions to the ethnopharmacology of Spain. J Ethnopharmacol. 1995;45:157–163.
- Ahmad FB, Holdsworth DK. Traditional medicinal plants of Sabah, Malaysia part III. The Rungus people of Kudat. Int J Pharmacog. 1995;33:262–264.
- 84. Amusan OOG, Msonthi JD, Makhubu LP. Molluscicidal activity of Spathodea campanulata, Andrachneovalis, Phytolacca dodecandra and Hypoxis rooperi. Fitoterapia. 1995;66:113–116.
- 85. Zani CL, Chaves PPG, Queiroz R, et al. Brine shrimp lethality assay as a prescreening system for anti-trypanosoma cruzi activity. Phytomedicine. 1995;2:47–50.
- Pisha E, Chai H, Lee IS, et al. Discovery of betulinic acid as a selective inhibitor of human melanoma that functions by induction of apoptosis. Nat Med. 1995;1(10):1046– 1051.
- Laferriere JE. Medicinal plants of the lowland Inga people of Colombia. Int J Pharmacog. 1994;32:90–4.
- Lin LZ, Hu SF, Zaw K, et al. Thalifaberidine, a cytotoxic aporphine-benzylisoquinoline alkaloid from Thalictrum faberi. J Nat Prod. 1994;57:1430–1436.
- 89. Bae K H, Min BS, Park KI, Ahn BZ. Cytotoxic flavonoids from Scutellaria indica. Planta Med. 1994;60:280–281.
- 90. Ishiguro K, Nagata S, Fukumoto H, et al. A 2-pyrone derivative from Hypericum japonicum. Phytochemistry. 1994;37:283– 284.
- 91. Chiang HC, Juilo Y, Lu FJ. Xanthine oxidase inhibitors from the leaves of Alsophila spinulosa (Hook) Tryon. J Enzyme Inhib. 1994;8:61–71.
- 92. Wu JB, Cheng YD, Kuo SC, et al. Fissoldhimine, a novel skeleton alkaloid from Fissistigma oldhamii. Chem Pharm Bulletin. 1994;42:2202–2204.
- 93. Pinto DCG, Fuzzati N, Pazmino XC, Hostettmann K. Xanthone and antifungal

constituents from Monnina obtusifolia. Phytochemistry. 1994;37:875–878.

- 94. Liu WK, Ng TG, Sze SF, Tsui KW. Activation of peritoneal macrophages by polysaccharopeptide from the mushroom, Coriolus 6ersicolor. Immunopharmacology. 1993;26:139–146.
- 95. Mohandoss S, Ravindran P. Flavonoids from Melastoma melabathricum. Fitoterapia. 1993;64:277–278.
- 96. Hammer M L A, Johns EA. Tapping an Amazonian plethora, four medicinal plants of Marajo Island, Para (Brazil). J Ethnopharmacol. 1993;40:53-75.
- 97. Segura-Correa R, Mata R, Anaya AL, et al. New tetranortriterpenoids from Swietenia humilis. J Nat Prod. 1993;56:1567–1574.
- 98. Ghazanfar SA, Al-Sabahi MA. Medicinal plants of Northern and Central Oman (Arabia). Econ Bot. 1993;47:89–98.
- 99. Cai Y, Chen JP, Phllipson J. Diterpenes from Croton lechleri. Phytochemistry. 1993;32:755–760.
- 100. Hussain SJ, Alvi AB, Jahan M. A study on Unani medicinal plants, Asteratiqus. J Res Educ Indian Med. 1993;2:35–39.
- 101. Lin CC, Lin CH. Pharmacological and pathological studies on Taiwan folk medicine. IX. The hepatoprotective effect of the methanolic extract from Echinops grijisii. Am J Chin Med. 1993;21:33–44.
- 102. Omino EA, Kokwaro JO. Ethnobotany of Apocynaceae species in Kenya. J Ethnopharmacol. 1993;40:167–180.
- 103. Rehm S, Devor DE. Acute effects of 4ipomeanol on experimental lung tumors with bronchiolar or alveolar cell features in Syrian hamsters or C3H/HeNCr mice. J. Cancer Res Clin Oncol. 1993;120:41-50.
- 104. Stuppner H, Sturm S, Geisen G, et al. A differential sensitivity of oxindole alkaloids to normal and leukemic cell lines. Planta Med. 1993;supplement A:583.
- 105. Holdsworth DK. A preliminary study of medicinal plants of Easter Island, South Pacific. Int J Pharmacog. 1992;30:27–32.
- 106. Sun HD, Zhou QZ, Fujita T, et al. Rubescensin D, a diterpenoid from Rabdosia rubescens. Phytochemistry. 1992;31:1418– 1419.

- 107. Qian TX, Li LN. Isosalvianolic acid C, a depside possessing a dibenzooxepin skeleton. Phytochemistry. 1992;31:1068–1070.
- 108. Furusawa E, Chou SC, Furusawa S, et al. Antitumour activity of Ganoderma lucidum, an edible mushroom, on intraperitoneally implanted lewis lung carcinoma in synergenic mice. Phytother Res. 1992;6:300 – 304.
- Ding YL, Jia ZA. Two phenolic derivatives from Euphorbia kansui. Phytochemistry. 1992;31:1435 – 1436.
- 110. Lin CC, Lin CH, Chiu HF, et al. The Pharmacological and pathological studies on Taiwan folk medicine (VII). The antiinflammatory effect of Echinops grjiisii. Am J Chin Med. 1992;22:127 – 134.
- 111. De Feo V, Aquino R, Menghini A, et al. Traditional phytotherapy in the peninsula Sorrentina, Campania, Southern Italy. J Ethnopharmacol. 1992;36:113 – 125.
- 112. Wong BYY, Lau BHS, Teel RW. Chinese medicinal herbs modulated mutagenesis, DNA binding and metabolism of Benzo[A]Pyrene. Phytother Res. 1992;6:10 14.
- 113. Hua YY. Two new saponins from the roots of Phytolacca esculenta. Planta Med. 1992;58:99 101.
- 114. Sakagami H, Konno K, Kawazoe Y, et al. Multiple immunological functions of extract from the cone of Japanese white pine, Pinus par6iflora Sieb. et Zucc. Adv Exp Med Biol. 1992;319:331 – 335.
- 115. Kao WJ, Hung DZ, Tsai WJ, et al. Podophyllotoxin intoxication. Toxic effect of Bajiaolian in herbal therapeutics. Hum Exp Toxicol. 1992;11:480 – 487.
- Lee IR, Song JY, Lee YS. Cytotoxicity of folkloric medicines in murine and human cancer cells. Kor J Pharmacog. 1992;23:132 - 136.
- 117. Reddy MB, Reddy KR, Reddy MN. Ethnobotany of Cuddapah district, Andhra Pradesh, India. Int J Pharmacog. 1991;29:273 – 280.
- 118. Chhabra SC, Mahunnah RLA, Mshiu MN. Plants used in traditional medicine in Eastern Tanzania. V. Angiosperms (Passifloraceae to Sapindaceae). J Ethnopharmacol. 1991;33:143 – 157.

- 119. Holdsworth DK. Traditional medicinal plants of Rarotonga, Cook Islands. Part II. Int J Pharmacog. 1991;29:71–79.
- 120. Kijjoa A, Gonzalez MJTG, Pinto MMM, et al. Constituents of Knema laurina and Knema tenuiner6ia subsp. setosa. Planta Med. 1991;57:575–577.
- 121. Sati OP, Bahuguna S, Uniyal S, et al. A new saponin from Deeringia amaranthoides. J Nat Prod. 1990;53:466–469.
- 122. Konda Y, Iguchi M, Harigaya Y, et al. Hancokinol, a novel triterpene, from Cynanchum hancokianum. Tetrahedron Lett. 1990;31:5315 – 5318.
- 123. Bhat RB, Eterjere EO, Oladipo VT. Ethnobotanical studies from Central Nigeria. Econ Bot. 1990;44:382 – 390.
- 124. Plotkin MJ, Schultes RE. Virola, a promising genus for ethnopharmacological investigation. J Psychoactive Drugs. 1990;22:357–361.
- 125. Novaretti R, Lemordant D. Plants in the traditional medicine of the Ubaye valley. J Ethnopharmacol. 1990;30:1–34.
- 126. Uniyal MR. Utility of hitherto unknown medicinal plants traditionally used in Ladakh. J Res Educ Indian Med. 1990;9:89 – 95.
- 127. Vinesi P, Serafini M, Nicoletti M, et al. Plant regeneration and hypoxoside content in Hypoxis obtusa. J Nat Prod. 1990;53:196 – 199.
- 128. Chu DT, Wong WL, Mavlight GM. Immunotherapy with Chinese medicinal herbs: immune restoration of local xenogenic graft-versus-host reaction in cancer patients by fractionated Astragalus membranacueus in vitro. J Clin Lab Immunol. 1988;25(3):19-23.
- 129. Kokwaro JO. Medicinal plants of east Africa. East African Literature Bureau, Nairobi;1976.
- 130. Powell RG, Weisleder D, Smith CRJr, et al. Structures of harringtonine, isoharringtonine, and homoharringtonine. Tetrahydron Lett. 1970;11: 815-818.
- 131. Block G, Patterson B, Subar A. Fruit, vegetables, and cancer prevention: a review of the epidemiological evidence. Nutr Cancer. 1992;18(1):1–29.

- 132. Steinmetz KA, Potter JD. Vegetables, fruit, and cancer prevention: a review. J Am Diet Assoc. 1996;96(10):1027–39.
- 133. Reddy L, Odhav B, Bhoola KD. Natural products for cancer prevention: a global perspective. Pharmacol Ther. 2003;99(1):1–13.
- 134. Aggarwal BB, Shishodia S. Molecular targets of dietary agents for prevention and therapy of cancer. Biochem Pharmacol. 2006;71:1397–1421.
- 135. Hoyoku N, Michiaki M, Harukuni T, et al. Cancer prevention by carotenoids. Arch Biochem Biophys. 2009;483:(2):165-168.
- 136. Wan X, Luo M, Li X, et al. Hepatoprotective and anti-hepatocarcinogenic effects of glycyrrhizin and matrine. Chem Biol Interact. 2009;181:(1)15-19.
- 137. Caihua L, Michael DMc, Carmen M, et al. A Population-Based Case-Control Study of Marijuana Use and Head and Neck Squamous Cell Carcinoma. Cancer Prev Res. 2009;2(8):759-768.
- 138. Shiu-Chen H, Chi-Tang H, Shoei-Yn L, et al. Carnosol inhibits the invasion of B16/F10 mouse melanoma cells by suppressing metalloproteinase-9 through down-regulating nuclear factor-kappa B and c-Jun. Biochem Pharmacol. 2005;69(2):221-232.
- 139. Li M, Zhang Z, Hill DL, et al. Genistein, a dietary isoflavone, down-regulates the MDM2 oncogene at both transcriptional and posttranslational levels. Cancer Res. 2005;65(18):8200–8.
- 140. Ruiz D, Egea J, Tomas-Barberan FA, et al. Carotenoids from new apricot (Prunus armeniaca L.) varieties and their relationship with flesh and skin color. J Agric Food Chem. 2005;53:6368-74.
- 141. Kim GY, Kim JH, Ahn SC, et al. Lycopene suppresses the lipopolysaccharide-induced phenotypic and functional maturation of murine dendritic cells through inhibition of mitogen-activated protein kinases and nuclear factor-kappaB. Immunology. 2004;113(2):203–11.
- 142. Sunila ES, Kuttan G. Immunomodulatory and antitumor activity of Piper longum Linn. and piperine. J Ethnopharmacol. 2004;90:(2-3):339-346.

- 143. Shishodia S, Majumdar S, Banerjee S, et al. Ursolic acid inhibits nuclear factor-kappaB activation induced by carcinogenic agents through suppression of IkappaBalpha kinase and p65 phosphorylation: correlation with downregulation of cyclooxygenase 2, matrix metalloproteinase 9, and cyclin D1. Cancer Res. 2003;63(15):4375–83.
- 144. Takada Y, Aggarwal BB. Betulinic acid suppresses carcinogen-induced NF-kappa B activation through inhibition of I kappa B alpha kinase and p65 phosphorylation: abrogation of cyclooxygenase-2 and matrix metalloprotease-9. J Immunol. 2003;171(6):3278–86.
- 145. Cheng YH., Shen TF, Pang VF, et al. Effects of aflatoxin and carotenoids on growth performance and immune response in mule ducklings. Comp Biochem Physiol Toxicol Pharmacol. 1999;128:19–26.
- 146. Abdullaev FI. Cancer chemopreventive and tumoricidal properties of saffron (Crocus sativus L.). Exp Biol Med. 2002;227:20-5.
- 147. Kohno H, Marda M, Honjo S. Prevention of colonic preneoplastic lesions by the betacryptoxanthine and hesperidine rich powder prepared from Citrus Unshiu Marc. Juice in male F344 rats. J Toxicol Pathol. 1999;12:209-215.
- 148. Han SS, Keum YS, Seo HJ, et al. Capsaicin suppresses phorbol ester-induced activation of NF-kappaB/Rel and AP-1 transcription factors in mouse epidermis. Cancer Lett. 2001;164(2):119–26.
- 149. Yang F, Oz HS, Barve S, et al. The green tea polyphenol (-)-epigallocatechin-3-gallate blocks nuclear factor-kappa B activation by inhibiting I kappa B kinase activity in the intestinal epithelial cell line IEC-6. Mol Pharmacol. 2001;60(3):528–33.
- 150. Manna SK, Mukhopadhyay A, Aggarwal BB. Resveratrol suppresses TNF-induced activation of nuclear transcription factors NF-kappa B, activator protein-1, and apoptosis: potential role of reactive oxygen intermediates and lipid peroxidation. J Immunol. 2000;164(12):6509–19.
- 151. Cheng YH, Shen TF, Pang VF, et al. Effects of aflatoxin and carotenoids on growth performance and immune response in mule

ducklings. Comp Biochem Physiol Toxicol Pharmacol. 1999;128:19–26.

- 152. Berkarda B, Koyuncu H, Soybir GT, et al. Inhibitory effect of hesperidin on tumor initiation and promotion in mouse skin. Res Exp Med. 1998;198:93-99.
- 153. Laughton MJ, Evans PJ, Moroney MA, et al. Inhibition of mammalian 5-lipoxygenase and cyclooxygenase by flavonoids and phenolic dietary additives. Relationship to antioxidant activity and to iron ionreducing ability. Biochem Pharmacol. 1991;42(9):1673–81.
- 154. Soliman KF, Mazzio EA. In vitro attenuation of nitric oxide production in C6 astrocyte cell culture by various dietary compounds. Proc Soc Exp Biol Med. 1998;218(4):390–7.
- 155. Kumar A, Dhawan S, Aggarwal BB. Emodin (3-methyl-1,6,8-trihydroxyanthraquinone) inhibits TNF-induced NFkappaB activation, IkappaB degradation, and expression of cell surface adhesion proteins in human vascular endothelial cells. Oncogene. 1998;17(7):913– 8.
- 156. Geng Z, Rong Y, Lau BH. S-Allyl cysteine inhibits activation of nuclear factor kappa B in human T cells. Free Radic Biol Med. 1997;23(2):345–50.
- 157. Sueoka N, Sueoka E, Okabe S, et al. Anticancer effects of morphine through inhibition of tumour necrosis factor-a release and mRNA expression. Carcinogenesis. 1996;17(11):2337-2341.
- 158. Kune GA, Bannerman S, Field B, et al. Diet, smoking, serum beta-carotene and vitamin A in male nonmelanocytic skin cancer patients and controls. Nutr Cancer. 1992;18(3):237.
- 159. Gogate SS. Cytotoxicity of neem leaf extract: an antitumor. Natl Med J India. 1991;9:297.
- 160. Pomponi AS. The bioprocess-technological potential of the sea. J Biotechnol. 1999;70:5-13.
- 161. Schwartsmann G. Marine organisms and other novel natural sources of new anticancer drugs. Ann Oncol. 2000;11:235-243.
- 162. Schwartsmann G, Rocha AB, Berlinck R, et al. Marine organisms as a source of new anticancer agents. Lancet Oncol. 2001;2:221-225.
- 163. Schweitzer J, Handley FG, Edwards J, et al. Summary of the workshop on drug

development, biological diversity, and economic growth. J Natl Cancer Inst. 1991;83:1294-1298.

- 164. Rinehart KL. Antitumor compounds from tunicates. Med Res Rev. 2000;20:1-27.
- 165. Chun HG, Davies B, Hoth D, et al.Suffness. Didemnin B The first marine compound entering clinical trials as an antineoplastic agent. Invest New Drugs. 1986;4:279-284.
- 166. Geldof AA, Mastbergen SC, Henrar REC, et al. Cytotoxicity and neurocytoxicity of new marine anticancer agents evaluated using invitro assays. Cancer Chemother Pharmacol. 1999;44:312-318.
- 167. Demetri G, Garcia-Carbonero R, Harmon D, et al. Ecteinascidin-743 (ET-743) induces objective responses and disease control in patients with advanced non-osseous sarcomas: results from phaseII trials. Ann Oncol. 2000;11(Suppl 4):126.
- 168. Erba E, Bergamaschi D, Bassano L, et al. Ecteinascidin-743 (ET-743), a natural marine compound, with a unique mechanism of action. Eur J Cancer. 2001;37:97-105.
- 169. Damia G, Silvestri S, Carrassa L, et al. Unique pattern of ET-743 activity in different cellular systems with defined deficiencies in DNA-repair pathways. Int J Cancer. 2001;92:583-588.
- 170. Poncet J. The dolastatins, a family of promising antineoplastic agents. Curr Pharm Des. 1999;5:139-162.
- 171. Bai R, Pettit GR, Hamel E. Dolastatin10, a powerful cytostatic peptide derived from a marine animal. Inhibition of tubulin polymerization mediated through the vinca alkaloid binding domain. Biochem Pharmacol. 1990;39:1941-1949.
- 172. Pathak S, Multani AS, Ozen M, et al. Dolastatin10 induces polyploidy, telomeric associations and apoptosis in a murine melanoma cell line. Oncol Res. 1998;5:373-376.
- 173. Pettit GR. The bryostatins. Fortschr Chem Org Naturst. 1991;57:153-195.
- 174. Pagliaro L, Daliani D, Amato R, et al. Phase II trial of bryostatin-1 for patients with metastatic renal cell carcinoma. Cancer. 2000;89:615-618.
- 175. Varterasian ML, Mohammad RM, Shurafa MS, et al. PhaseII trial of bryostatin1 in

patients with relapsed low-grade non-Hodgkin's lymphoma and chronic lymphocytic leukemia. Clin Cancer Res. 2000;6:825-828.

- 176. Zonder JA, Shields AF, Zalupski M, et al. A phase II trial of bryostatin 1 in the treatment of metastatic colorectal cancer. Clin Cancer Res. 2001;7:38-42.
- 177. Ahmad I, Al-Katib AM, Beck FW, et al. Sequential treatment of a resistant chronic lymphocytic leukemia patient with bryostatin1 followed by 2chlorodeoxyadenoside: case report. Clin Cancer Res. 2000;6:1328-1332.
- 178. Mayer A and Gustafson KR. Marine pharmacology in 2002–2004: Anti-tumour and cytotoxic compounds. Eur J Cancer. 2006;42:2241–2270.
- 179. Mayer A and Gustafson K R. Marine pharmacology in 2005–2006: Anti-tumour and cytotoxic compounds. Eur J Cancer. 2008;44: 2257–2287.
- 180. Binaschi M, Farinosi R, Borgnetto ME, et al. In vivo site specificity and human isoenzyme selectivity of two topoisomerase II poisoning anthracyclines. Cancer Res. 2000;60:3770-3776.

- 181. Patrick Y. Major microbial diversity initiative recommended. Am Soc Microbiol News. 1997;63:417-421.
- 182. Alberts MW, Williams RT, Brown EJ, et al. KBP-Rapamycin inhibits a cyclin-dependent kinase activity and a cyclin D1-Cdk association in early Gl of an osteosarcoma cell line. J Biol Chem. 1993;268:22825-22829.
- 183. Schulte TW, Neckers LM. The benzoquinone ansamycin 17-allylamino-17 demethoxygeldanamycin binds to HSP90 and shares important biologic activities with geldanamycin. Cancer Chemother Harmacol. 1998;42:273-279.
- 184. Cadenas ME, Sandfrison A, Cutler NS, et al. Signal transduction cascades as targets for therapeutic intervention by natural products. Trends Biotechnol. 1998;16:427-433.
- 185. Adjei AA. Signal transduction pathway targets for anticancer drug discovery. Curr Pharmaceut Design. 2000;6:361-378.