

International Journal of Phytomedicine 2 (2010) 354-362

http://www.arjournals.org/ijop.html

Research article



Immunomodulatory effects of herbal plants plus melatonin on human blood phagocytes

Eduardo Luzia França¹, Joanna Conde Maynié², Valéria Conde Correa², Uender Correia Rodrigues Pereira¹, Claudemir Batalini³, Carlos Kusano Bucalen Ferrari¹, Adenilda Cristina Honorio-França¹*

*Corresponding author:

Adenilda Cristina Honorio França

 Institute of Biological and Health Science - Federal University of Mato Grosso – Pontal do Araguaia – MT, Brazil. - Rodovia MT 100, Km 3,5 s/n°, Pontal do Araguaia - MT. CEP:78698-000 Email:- <u>denifran(at)terra.com.br</u>
Environmental Biodiversity Center EcoCerrado Reserve, Araxá, MG-Brazil.
Federal University of Mato Grosso

3. Federal University of Mato Grosse Pontal do Araguaia – MT, Brazil

Abstract

It has been shown a mixture of seven herbal plants was able to trigger cell oxidative mechanism and subsequently inducing cellular activation. Moreover, melatonin hormone has also been shown to perform different actions on the cellular oxidative metabolism. It is possible that the herbal mixture associated with melatonin can activate phagocytes, improving microbicidal activity and further ameliorating resistence to the infections. The aim of this work was to verify in vitro immunomodulatory effects of melatonin and a medicinal plants mixture on blood mononuclear phagocytes (MN). We collected 40 blood samples from normal individuals to obtain the phagocytes. The MN phagocytes were separated by Ficoll-Paque gradient. Preparation of plant extract to obtain the herbal mixture was carried through the process of maceration followed by distillation. Phagocytosis and microbicidal activity of blood phagocytes, treated or not with exogenous superoxide dismutase (SOD), against enterophathogenic Escherichia coli (EPEC) were evaluated by acridine orange method. The herbal mixture and/or melatonin were added to the cell suspensions as immunomodulators. We observed increased phagocytosis and microbicidal activities by blood MN phagocytes in the presence of melatonin or the herbal mixture. The association of both potentiated the functional activity of blood MN phagocytes. Phagocytes previously treated by exogenous SOD had decreased microbicidal activity independently of immunomodulators. These data suggest that the herbal mixture is a potent immunostimulatory agent, and that the interaction between plant and hormones may represent an alternative mechanism of defense against infection, especially in immunosuppressed patients.

Keywords: herbal mixture, melatonin, blood phagocytes, phagocytosis, superoxide

Introduction

Several studies attempt to elucidate the antimicrobial action of phagocytes. On this respect, it was assigned an important role for oxygen-derived radicals and their possible modulators [1-4]. The multiple cascades displayed by free radicals may be associated on phagocytosis and microbicidal activity designed to eliminate potentially pathogenic agents [2].

Phagocytosis represents an important defense mechanism especially against bacterial infections. In phagocytosis occurs massive activation of cellular oxidative metabolism with generation of potent active oxygen metabolites. Oxygenderived radicals are involved in many important processes such as immune reactions, longevity, and peroxidation of cell lipids, proteins, carbohydrates, and DNA [1,5].

On the order hand, the literature has been demonstrated that many plants can stimulate immune cells constituting a real promise for treatment of infections [6]. The use of medicinal plants has been increased and there is a mojor scientific breakthrough on issues related to both chemical and pharmacological studies and the search for a new therapeutic compounds [7,8] especially of alternative drugs in disease treatment, including infections diseases.

In Brazilian ethnopharmacology a popular mixture of seven plants has been shown to be useful for immune system improvement as well as increased resistance to tumor cells. This herbal mixture includes the extract preparation of the following plants: Orbignya Marti, Tabebuia avellanedae, Arctium lappa, Rosa centifolia, Maytenus ilicifolia, Vernonia condensata and Thujae occidentalis. It has been demonstrated that this herbal mixture has ability to massively activate cellular oxidative mechanisms [9], but its implications for phagocytosis and microbicidal activity are still partially understood.

Literature also reports the importance of hormones and neuropeptides as potent immunomodulators, among them the melatonin hormone [10], involved in many different aspects of the regulation of functional activity on the immune system [11].

Melatonin functions are still only partially understood [12]. Research has been pointed out that melatonin, synthesized by pineal gland, is able to modulate the immune response [10,12] in dose-dependent manner [13,14]. The a involvement of melatonin on phagocytes function is controversial. Some authors postulate that melatonin activates immune system [10,15], considered it as a whereas others had immunosuppressant factor [16], while others report that this hormone is a also produced by the activated phagocytes [17].

Previous report in vitro has been pointed out that this herbal mixture and melatonin play an important role in modulating cell activation once it triggers activation of blood phagocytes by increasing the superoxide anion production [10,11]. It is possible that this mixture associated to melatonin can activate the phagocytes and increase the microbicidal activity of these cells which can further improves resistance against infections.

The aim of the present study was to verify in vitro immunomodulatory effects of melatonin and a medicinal plants mixture on blood mononuclear phagocytes.

Materials and Methods

Subjects: After the informed consent, a sample of 15 mL of blood was collected from 40 clinically healthy men ranging from 18 to 35 years of age. All procedures were submitted to ethical evaluation and obtained institutional approve.

Separation of blood cells: Blood samples were collected into heparinized (25U/ml) tubes. The samples were separated by a Ficoll-Paque gradient (Pharmacia, Upsala, Sweden), density gradient (density 1.077 g/l), producing preparations with 98% of pure mononuclear (MN) phagocytes, analyzed by light microscopy. Purified MN phagocytes were resuspended independently in serum-free medium 199 at a final concentration of $2x10^6$ cells/mL. *E. coli strain:* The Enterophatogenic *Escherichia coli* used was the EPEC O111 – H⁻ serotype. The stock culture was cultivated in Trypic Soy Broth (TSB, Difco) for 18 hours at 37°C. Bacteria were washed twice in phosphate buffered saline (PBS) and adjusted to an concentration of 1 x 10⁷ bacteria/mL as measured by turbidimetry at 540 nm, using a spectrophotometer (Femto). This bacterial concentration was previously determined by colony unit counting on Trypic Soy Agar (TSA, Difco, Detroit) [2].

Preparation of Herbal Mixture: The herbal mixture was composed of 75% leaf dry of Orbignya martiana, 2% of the bark of Tabebuia avellanedae, 4% of leaf of Arctium lappa, 5% of petals of Rosa centifolia, 5% of leaf of Maytenus ilicifolia, 5% of the leaf of Vernonia condensata and 4% of the leaf of Thujae occidentalis. All these plants were collected and deposited in the herbarium at the Environmental Biodiversity Center - "EcoCerrado" Reserve, Araxá - MG, Brazil, localized at Lat. 19 ° 36'47, 1" Long. 47 ° 08'20, 9", with a 939m altitude.

The preparation involved the mixing process followed by maceration and distillation according to the Brazilian pharmaceutical code [9]. For processing of macerates, the plant parts were macerated by placing 200 g of the plant for one liter of alcohol 70%. The plant was left soaking for thirty days at room temperature. During the first ten days the preparations were shaken once a day. After this period the preparation was filtered. For the distillation process the samples were passed on and still were concentrated until syrupy consistency in temperature up to 60oC and in the mixture was added preservative NIPAGIN® M.

Phytochemical screening

Phytochemical screening for identification and indication for their main chemical constituents of aqueous extract of herbal mixture was done [18]. Following reagents and chemicals were used alkaloids with dragendroff's reagents, flavonoids with metallic magnesium plus HCl, saponins with the ability to produce foam, reducing sugars with Fehling's reagent, glycosides with Liberman's test, tannins with ferric chloride and polysaccharides with iodine solution.

Treatment of blood phagocytes with melatonin and with herbal mixture

To verify the activity of melatonin and the herbal mixture phagocytosis, mononuclear on phagocytes $(2x10^{6})$ were treated with 30 min before phagocytosis. To check the phagocytosis and microbicidal activity by mononuclear phagocytes (2×10^6) , the cells were treated with hormone and/or with the herbal mixture immediately after phagocytosis tests. For each test performed, MN phagocytes (2×10^6) were also incubated in culture medium 199 in the absence of the melatonin and the herbal mixture. Concentrations of the hormone melatonin were 10^{-7} molar [19] and the herbal mixture 1mg/ml [9].

Bactericidal assay

Viability test, phagocytosis and microbicidal activity were evaluated using the acridine orange method described by Bellinati-Pires et al. [20]. Equal volumes of bacteria and cell suspensions were mixed and incubated at 37°C for 30 min under continuous shaking. Phagocytosis was stopped by incubation in ice. To eliminate extracellular bacteria the suspensions were centrifuged twice (160 g, 10 min, 4° C), and the cells were resuspended in serum-free medium 199 and centrifuged. The supernatant was discarded and the sediment dyed with 200µL of acridine orange (14.4g/L) for 1 minute. The sediment was resuspended in cold culture 199, washed twice and observed under immunofluorescence microscope at 400x and 1000x magnification. The viability index was calculated by counting 100 cells. For viability, the viable cells were shown to be green and red The phagocytosis non-viable. index was calculated by counting the number of cells ingesting at least three bacteria in a pool of 100 cells. To determine the bactericidal index, cells were stained with acridine orange and the killing of the bacteria were verify in 100 cells counted per slide. The dead EPEC, viewed with orange

stain [20]. The assays of phagocytosis and microbicidal activity were made in the presence or absence of superoxide dismutase (SOD; 140 units) [3,21]. All the experiments were performed in duplicate or triplicate.

Results

Phytochemical screening

Phytochemical screening showed the presence of tannins, especially condensed type or catechist. The mixture also presented in its chemical composition leucoanthocyanidins, flavanones and catechins. To lower degree there was presence of anthocyanins, proanthocyanidins, phenols, flavonoids, flavanones, xanthones, triterpenoids and saponins. The herbal mixture was negative steroids. resins. alkaloids, quaternary for compounds, quinones, flavonoids aglycones and steroid aglycone triterpenoids (Table 1).

Table 1 – Phytochemical screening for identificationand indication of main chemical constituents of aqueousextract of herbal mixture (HM).

| Analysis | Aqueous Extract (HM) |
|---|----------------------|
| Tannins | ++ |
| Leucoanthocyanidins, catechins and flavanones | ++ |
| Phenols | + |
| Anthocyanins, anthocyanidins and flavonoids | + |
| Flavanones and xanthones | + |
| Triterpenoids | + |
| Saponins | + |
| Resins | - |
| Alkaloids | - |
| Quaternary compounds | - |
| Quinones | - |
| flavonoids aglycones | - |
| Steroid aglycone triterpenoids | - |

Blood MN phagocytes viability in the presence of the herbal mixture

The results of blood MN phagocytes viability in the presence of the herbal mixture are showed in the Table 2. It was observed untreated cells had 99% viability. Similar rates were also observed when these cells were incubated by the herbal mixture and melatonin, demonstrating the lack of toxicity of immunomodulatory agents on used dosage.

| | and index of viability of Blood phagocytes in the presence of |
|-------------------------|--|
| melatonin hormone (HM). | (MLT) or/and herbal mixture |

| | Experimental Group | | |
|--|--------------------|-------------|---------------|
| Parameter | 199 Mediu m | MLT | HM |
| Phagocyte count $(x10^{6} \text{ cells/ml})$ | 3.9 ± 0.2 | 3.8 ± 0.6 | 4.0 ± 0.3 |
| Viability of MN phagocytes (%) | 99 ± 1.2 | 97 ± 1.1 | 99 ± 1.3 |

Results represent the mean (\pm SD) of ten experiments with cells from different individuals. (ANOVA p>0.05)

The phagocytic activity of blood MN cells for EPEC in the presence of the herbal mixture and melatonin hormone was significantly increased when the cells were stimulated by herbal mixture and melatonin (Figure 1A).

The herbal mixture plus melatonin increased the bactericidal index when compared to the results presented by cells incubated only with the bacteria or melatonin (Figure 1B).

Treatment of blood phagocytes by exogenous SOD did not interfered on phagocytosis (Figure 2A). In respect of the microbicidal activity of blood MN treated by exogenous SOD, independent of the effect of immunomodulatory agents, it was observed lower bactericidal index (Figure 2B).

⁽⁺⁺⁾ strong positive reaction (+) positive reaction

^(-) negative reaction

Discussion

In this study we evaluated the in vitro immunomodulatory effect of melatonin associated with a mixture of seven herbal plants on phagocytosis and microbicidal activity of MN human blood phagocytes against enterophatogenic Escherichia coli, and their interactions with cellular oxidative mechanisms. Plants are considered an unlimited source of molecules for new treatment of diseases. There is growing interest in the investigation of different plant species to identify their potential for therapeutic application because of the large historical legacy of medicinal plants [22] with fewer side effects, lower cost and toxicity. There is general consensus that the adverse effects of herbal medicines are less frequent when compared to synthetic drugs [23].

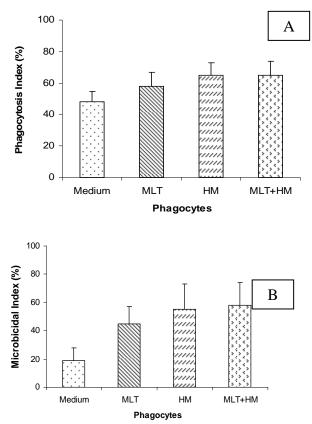


Fig 1 Phagocytosis and microbicidal index of blood MN cells for EPEC in the presence of the herbal mixture and the hormone melatonin.

Literature reports the effectiveness of medicinal plants which have been opened prospect for obtaining new drugs [24] and studies relate that a large number of plants used in traditional healing are employed in often sophisticated mixtures, rather than as individual plants [25]. Several medicinal plants have shown ability to induce immune system activation displaying beneficial effects against disease [25,26].

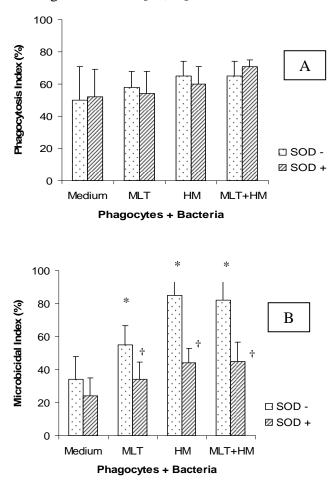


Fig 2 : Phagocytosis index and microbicidal activity of blood MN phagocytes treated with exogenous SOD in the presence of the herbal mixture and the hormone melatonin.

In this study it was found that the herbal mixture presented a potent immunstimulatory effect on the functional activity of blood phagocytes. Association of melatonin with the herbal mixture

potentiated both the phagocytic and microbicidal activities. One of the herbs used in this mixture is Thujae occidentalis. One interesting clinical controlled trial reported that an extract containing Thujae occidentalis and other two herbs improved common cold symptoms in human patients [27]. Microbicidal activity is an mechanism elimination important for of infection, particularly those caused by bacteria [2,3]. Neurohormonal control is very important to immunobiological modulate effects [28]. Melatonin has beneficial free radical scavenging actions beyond its stimulatory effects on the cytosolic antioxidant enzyme systems [14,29]. Many studies have been reported that melatonin strongly stimulates cells of the immune system [3,19]. In a previous study we demonstrated the functional activity of human blood phagocytes against bacteria and fungi are modulated by hormones. The melatonin hormone has increased microbicidal activity of mononuclear phagocytes as well as rat macrophages [4,10,15]. This study reported that melatonin presented imunostimulatory effects that may be potentiated by the herbal mixture.

Many medicinal plants have shown their ability to stimulate different immunomodulatory pathways [30,31].

This study confirmed an additive effect between endogenous peptides and plants with medicinal activity. The combination of melatonin and the herbal mixture increased the functional activity of phagocytes. In literature, several studies attempted to elucidate the antimicrobial action of phagocytes. Regarding this aspect it was assigned an important role for oxygen-derived radicals and their possible modulators [1-3,32]. Our previous study confirmed that the herbal mixture can modulate oxygen metabolism pathways [10].

Macrophages play an important role on the mechanisms of the body's defense against infections. The functional activity of macrophages in various biological systems has been associated with immunomodulatory activity [33]. The generation of free radicals has been reported as an important mechanism for protecting the body during the infectious processes [2,32].

During the oxidative stress the cellular mitochondrial and peroxisomal metabolism generates large amounts of the superoxide anion [10,15]. The free radical releasing has been reported as an important body's defense mechanism especially in gut infections [2,32].

The action of reactive oxygen metabolites has been considered an important mechanism of bacterial killing since an ineffective phagocytosis is associated with formation of granulomas [21]. The superoxide anion is the first molecule from the oxygen free radical cascade of oxygen metabolism.

Moderate oxidative stress is often accompanied by an increase in enzymatic antioxidant defenses that can neutralize deleterious effects of free radicals [34]. Among these enzymes superoxide dismutase (SOD) act by catalyzing the dismutation of superoxide into H_2O_2 and O_2 resulting as an important antioxidant role [34].

In this study, we evaluated if the functional activity of phagocytes modulated by the association of melatonin and an herbal mixture is oxidative-dependent we used exogenous SOD. Phagocytes treatment by SOD reduced the microbicidal activity of cells stimulated by both the melatonin and the herbal mixture. This result suggests that both melatonin and the herbal mixture are able to activate the cellular oxidative metabolism and exert important immunomodulatory effects on phagocytosis and microbicidal activity.

SOD comprises a very important mechanism on superoxide anion reduction [35] so the results of this study suggested that the microbicidal activity of human blood phagocytes is dependent on superoxide anion release constituting an important defense mechanism against bacterial infections. It is important to note that both the herbal mixture as the melatonin hormone, in the used dosages, did not exert cytotoxic effects in these cells.

Conclusion

These data suggest that the herbal mixture is a potent immunostimulatory agent, and that the

interaction between plant and hormone may represent an alternative mechanism of defense against infection.

List of Abbreviations

DNA- deoxyribonucleic acid MN phagocytes – mononuclear phagocytes TSB - Trypic Soy Broth TSA - Trypic Soy Agar EPEC – Enteropathogenic *Escherichia coli* SOD – Superoxide dismutase MLT – melatonin hormone HM – Herbal mixture H₂O₂ - hydrogen peroxide

Acknowledgments

We are very grateful to the "Naturoterapia Sinhô Mariano" laboratory. This research received grants from Fundação de Amparo à Pesquisa de Mato Grosso (FAPEMAT N° 738264/2008).

References

- 1. Asad NR, Assad LMBO, Almeida CEB, Leitão, A.C. Lethal interaction between hydrogenperoxide and 0-phenonthroline in *Escherichia coli*. Br J Med Biol Res 1994; 27: 2551-2555.
- 2. Honorio-França AC, Carvalho MPSM, Isaac L, Trabulsi LR, Carneiro-Sampaio MMS. Colostral mononuclear phagocytes are able to kill Enteropathogenic *Escherichia coli* opsonized with colostral IgA. Scand J Immunol 1997;46: 59-66.
- 3. Honorio-França AC, Launay P, Carneiro-Sampaio MMS, Monteiro RC. Colostral neutrophils express IgA Fc receptors (CD89) lacking y chain association that mediate noninflammatory properties of secretory IgA. J Leuk Biol 2001; 69: 280-288.
- França EL, Pereira Jr A, Oliveira SL, Honorio-França AC. Chronoimmunomodulation of melatonin on bactericidal activity of human blood phagocytes. Intern J Microbiol 2009; 6:1-15.

- Ames BN, Shinnenaga MK, Hagen TM. Oxidants, antioxidantes, and the degenerative diseases of agent. Proc Nat Acad Sci 1993; 90: 7915-7922.
- Tiwari U, Rastogi B, Singh P, Saraf DK, Vyas SP. Immunomodulatory effects of aqueous extract of Tridax procumbens in experimental animals. J Ethnopharmacol 2004; 92: 113-119.
- Atal CK, Sharma ML, Kaul A, Khajuria A. Immunomodulating agents of plant origin. I: Preliminary screening. Journal Ethnopharmacol. 1986; 18:133-134.
- 8. Viegas Jr C, Bolzani VS, Barreiro EJ. The natural products and the modern medicinal chemistry. Quím Nova 2006; 29:326-337.
- Corrêa VSC, Maynié JC, França EL, Honorio-França AC. Activity of phagocytes in the Presence of the "Mais Vida" (more life) Herbal Remedy. Braz J Med Plants 2006; 8: 26-32.
- França EL, Feliciano ND, Silva KA, Ferrari CKB, Honorio-França AC. Modulatory Role of Melatonin on Superoxide Release by Spleen Macrophages Isolated from Alloxan-Induced Diabetic Rats.. Bratisl Med J 2009;7:163-173.
- 11. Dardenne M, Savino W. Control of thymus physiology by peptidic hormones and neuropeptides. Immunol Today 1994; 15: 519-523.
- Cassone VM. The pineal gland influences rat circadian activity rhythms in constant light. J. Biol. Rhythms 1992; 7: 27-40.
- 13. Claustrat B, Brun J, Chazot G. The basic physiology and pathophysiology of melatonin. Sleep Med Rev 2005; 9:11–24.
- 14. Pandi-Perumal Sr, Trakht I, Srinivasan V, Spence DW, Maestroni GJM, Zisapel N. Physiological effects of melatonin: Role of melatonin receptors and signal transduction pathways. Prog Neurobiol 2008; 85: 335–353.

- 15. Honorio-França AC, Silva KA, Feliciano ND, Calderon IMP, Rudge MVC, França EL. Melatonin effects on macrophage in diabetic rats and the maternal hyperglycemic implications for newborn rats. Int J Diabet Metabol 2009; 17:87-92.
- Ianãs O, Olinescu R, Bãdescu I. Melatonin involvement in oxidative processes. Rom J Endocrinol 1991; 29:147-153.
- Finocchiaro LME, Nahmod VE, Launay JM. Melatonin biosíntesis and metabolism in peripheral blood mononuclear leucocytes. Biochem J 1991; 280:727-731.
- Harbone, J.B. 1984. Phytochemical Methods. A guide to Modern Techniques of Plant Analysis, Second ed. Chapman and Hall, London, pp. 84-274.
- 19. Pawlak J, Singh J, Lea RW, Skwarlo-Sonta K. Effect of melatonin on phagocytic activity and intracellular free calcium concentration in testicular macrophages from normal and streptozotocin-induced diabetic rats. Mol Cell Biochem 2005; 275: 207-213.
- 20. Bellinati-Pires R, Salgado MM, Hypolito IP, Grumach AS, Carneiro-Sampaio, MMS. Aplication of a fluorochrome-lysostaphin assay to the detection of phagocytic and bactericidal disturbances in human neutrophils and monocytes. J Investig Allergol Clin Immunol. 1995; 5:337-342.
- Babu U, Failla ML. Copper status and fuction of neutrophlis are reversibly depressed in marginally and severely copper-deficient rats. J Nutr 1990; 120:1700-1709.
- Rêgo, TJA. Fitogeografia das plantas medicinais no Maranhão. 2nd ed. EDUFMA, MA, 108-109, 1995.
- 23. Calixto, J.B. Efficacy, safety, quality marketing and regulatory guidelines for herbal medicines (phytotherapeutic agents). Braz J Med Biol Res 2000; 32: 179-189.
- 24. Hasani-Ranjbar S, Larijani B, Abdollah MA. Systematic review of Iranian medicinal plants

useful in diabetes mellitus. Arch Med Sci 2008;.4: 285-292.

- 25. Bussmann RW, Meyer AGK, Kuhlman A, Townesmith A. Herbal mixtures in traditional medicine in Northern Peru. J Ethnobiol Ethnomedicine 2010; 6:2-11.
- 26. Honorio-França AC; Marins C. Boldrini F, França EL. Evaluation of hypoglicemic activity and healing of extract from amongst bark of "Quina do Cerrado" (*Strychnos pseudoquina* ST. HILL). Acta Cir Braz 2008; 23: 504-510.
- 27. Naser B, Lund B, Henneicke-Von Zepellin HH, Köhler G, Lehmacher W, Scaglione F. A randomized, double-blind, placebo-controlled, clinical dose-response trial of an extract of *Baptisia*, *Echinacea* and *Thuja* for the treatment of patients with common cold. Phytomedicine. 2005;12:715-722.
- 28. Besedovsky HO, Del Rey A. Immune-neuroendocrine interactions: Facts and hypotheses. Endocr Rev 1996; 17: 64–102.
- 29. Klepac N, Rudes Z,Klepac R. Effects of melatonin on plasma oxidative stress in rats with streptozotocin induced diabetes. Biomed Pharmacother 2005; 60: 32-35.
- Sudnikovich EJ, Maksimchik YZ, Zabrodskaya SV, Kubyshin VL, Lapshina EA, Bryszewska M. Melatonin attenuates metabolic disorders due to streptoxotocininduced diabetes rats. Eur J Pharmacol 2007; 569: 180-187.
- Manosroi A, Saraphanchotiwitthaya A, Manosroi J. Immunomodulatory activities of Clausena excavata Burm. f. wood extracts. J Ethnopharmacol 2003; 89: 155-160.
- 32. Gokhale AB, Damre AS, Saraf MN. Investigations into the immunomodulatory activity of Argyreia speciosa. J Ethnopharmacol 2003; 84: 109-114.
- 33. França-Botelho AC, Honorio-França AC, França EL, Gomes MA, Costa-Cruz JM. Phagocytosis of *Giardia lamblia* trophozoites

by human colostral leucocytes. Acta Paediat 2006; 95: 438-443.

- 34. Kang NS, Park SY, Lee SM, Lee BG, Shin DH, Pyo S. 2002. Modulation of macrophage function activity by ethanolic extract of larvae of *Holotrichia diomphalia*. J Ethnopharmacol 2002; 79: 89-94.
- 35. Traber MG. Cellular and molecular mechanisms of oxidants and antioxidants. Min Electrol Metabol 1997; 23: 135-139.
- 36. Gort AS, Imlay JA. Balance between endogenous superoxide stress and antioxidant defense. J Bacteriol 1998; 180:1402-1410.