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Review Article

A Brief Review on Phytoconstituents and Ethnopharmacology of *Scoparia Dulcis* Linn. (Scrophulariaceae)

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ABSTRACT

Scoparia dulcis Linn.(S. dulcis)or sweet broom weed commonly known as Mithipatti and Bana Dhania in Western Orissa, it is also known as 'GhodaTulsi'in Hindi. The present review attempts to narrate the chemical constituents of S.dulcis and their uses. S.dulcis is rich in flavones, terpenes and steroids. Main chemical constituents such as scoparic acid A-C, scopadulcic acid A and B, scopadulciol, scopadulin and ammelin have been shown to contribute to the observed medicinal effect of the plant. In this review we have composed the structure and functions of those active ingredients with their melting point and other physical properties individually. Some aspects of the several speculated pharmacological properties of S. dulcis have been validated by scientific research, which includes the presence of hypoglycaemic and antitumour promoting compound. It also has antimicrobial and antifungal effects as well as antihyperlipidemic action.

Keywords: Scoparia dulcis, scoparic acid, ammelin, medicinal effect.

Introduction

Scoparia dulcis Linn.is an erect annual herb with serrated leaves, producing white flowers and measuring up to a half meter in height when fully grown, it is an herb widely distributed in tropical and subtropical regions. Its ethno-medicinal uses amongst various indigenous tribes in the rain-forest zone are well-documented [1]. In fresh or dried form S. dulcis plants have been traditionally used as remedies for Diabetes mellitus in India and hypertension in Taiwan [3]. It is used in curing ailments such as fever, diarrhoea, ulcer, cancer, wounds, skin rash, cough and tuberculosis. The fresh or dried plant has been used for treating stomach aches, inflammation, bronchitis, hemorrhoids and hepatosis. In the western part of Orissa its root is traditionally is used as an effective remedy for Jaundice and diarrhoea. It is also used as an analgesic and antipyretic, in stomach troubles,[2] bronchitis, as well as inhibition of herpes simplex virus

replication, gastric H+,K+-ATPase activation and antitumor activity. It is deemed to be a panacea for all ills. In Gambia, a lotion prepared from the plant is used in curing fever. A hot water infusion or decoction of the leaves or whole plant is used medicinally by indigenous tribes of Nicaragua to treat malaria, stomach disorders, menstrual disorders, insect bites, fevers, heart problems, liver disorders and venereal diseases. It has been used for blood cleansing, in childbirth and as a general tonic. [3] Phytochemical screening has revealed that the plant contains diterpenoids, flavonoids, tannins, alkaloids, triterpenes, hexacosonol. -sitosterol. ketone-dulcitone and ammelin, an antidiabetic compound [2-4]. diterpenoid, scoparic acid A, isolated from the plant has been reported to be a potent -glucuronidase [5]. The constituents. scopadulciol. scopadulcic acid-B and diacetylscopadiol, have been shown to be responsible for the inhibitory activity of the plant on gastric H+-K+ ATPase enzyme [6]. The

diterpenoid, scopadulcic acid-B and flavone, hemenoxin, have been shown to exhibit cytotoxic and antitumor activity [7].

Objective for studying medicinal plants is the discovery of new bioactive components, in the search for promising drugs. This review emphasizes the traditional uses and clinical potential of S.dulcis. Through this review, authors hope to attract the attention of natural productresearchers throughout the world to focus on the unexplored potential of weed like S.dulcis (mithipatti).

The available information on S.dulcis has been divided into four sections, i.e., Plant profile ethnopharmacology, phytoconstituents, pharmacological reports. The reports in which S.dulcis species have been used as a domestic remedy by common men without any prescription for the treatment of various ailments have been discussed under ethnopharmacology.

Plant ProfileofScopariadulcis Linn.-



Fig.1: Plant S.dulcis.

Fig.2: Plant S.dulcis Herbarium.

Vernacular Name

Sanskrit: Asmaghni

Hindi: Mithi Patti, GhodaTulsi, Ban Dhania

English: Sweet broom, Broom weed, Vassourinha

Taxonomy:

Kingdom: Plante

Subkingdom: Trachcobionta Division: Magnoliophyta Class: Magnoliopsida Subclass: Asteridae Family: Scrophulariaceae

Genus: Scoparia **Species**: dulcis

Botanical name: Scoparia dulcis Linn.

Morphology

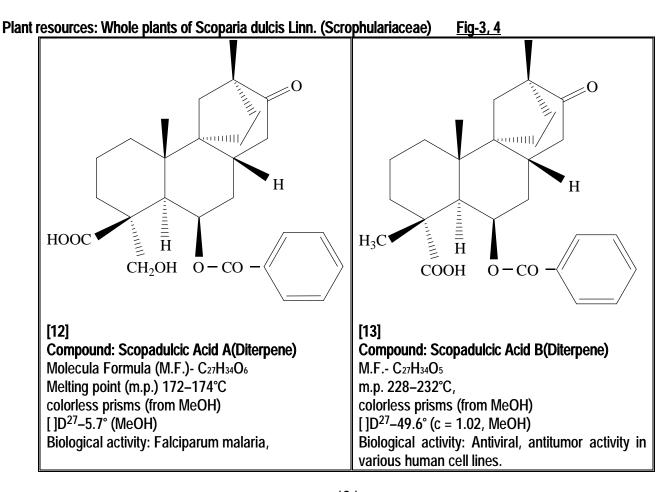
It is a small, much branched, glabrous, leafy annual herb or under shrub with erect or ascending branches; Leaves opposite and 3-notely whorled, rhomboid, elliptic or elliptic lanceolate, obtuse at apex, base tapering, margins serrate; Flowers many, in terminal panicles, pedicelate, pedicels slender, rigid, Calyx lobes 4, oblong, Corolla white, tube very short, Capsule globose; seeds minute, many. [8-9]

Traditional Uses of Scoparia dulcis Linn. [8-11]

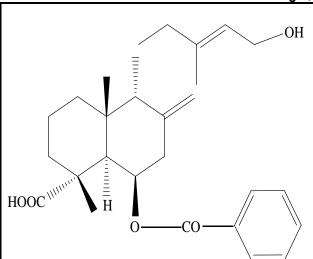
Plant Part				
U S E S	Aerial Part	Leaf	Root	Whole Plant
	Coughs,	Diabetes, diarrhoea, eye problems,	Bronchitis,	Anemia, bronchitis, burns, coughs,
	diarrhoea,	fever, headaches, hemorrhoids,	diarrhoea, fever,	diabetes, diarrhoea, dysentery,
	expectoran	infections, insect bites, intestinal worms,	jaundice, liver	expectorant, fever, gastric disorders,
	t, fever and	kidney disease, liver disorders, malaria,	disorders, malaria,	headache, hemorrhoids, hepatitis,
	stomach	menstrual disorders, migraines, snake	menstrual	hypertension, infections, insect bites,
	pains	bites, stomach disorders, tonic, ulcers,	disorders, skin	intestinal worms, jaundice, liver disease,
		urinary tract disorders, vomiting,	infections, stomach	malaria, menstrual disorders, pain, rash,
		wounds, anemia, burns, and cough	pains	snake bites, swelling and toothache

Phytoconstituents

The available literature on phytochemical reports of the S.dulcis reveals that it comprises mainly terpenes and flavones. Fig. 3 to 38 summarizes phytoconstituents reported from various plant parts of S. dulcis.







[14,15]

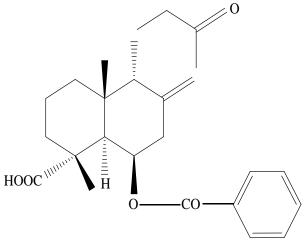
Compound: Scoparic Acid A (Diterpene)

M.F.- C₂₇H₃₆O₅

m.p. colorless amorphous powder

 $[]D^{26}-38.3^{\circ} (c = 1.00, CHCI_3)$

Biological activity: -glucuronidase inhibition



[15]

Compound: Scoparic Acid B (Diterpene)

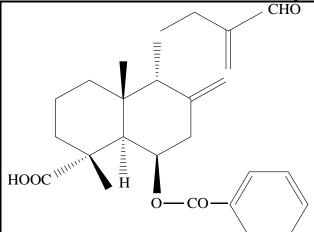
M.F.- C₂₅H₃₂O₅

m.p. colorless amorphous powder

 $[]D^{23}-9.8^{\circ} (c = 0.63, CHCl_3)$

Biological activity: Antiviral

Fig - 7, 8



[15]

Compound: Scoparic Acid C (Diterpene)

M.F.- C₂₆H₃₂O₅

m.p. colorless amorphous powder

 $[]\dot{D}^{22}-13.9^{\circ} (c = 0.69, CHCl_3)$

Biological activity: -glucuronidase inhibition

ОН

[16]

Compound : Apigenin (Flavone)

M.F-C₁₅H₁₀O₅

m.p.315°C

yellow crystalline powder

Biological activity: Antioxidant, radical scavenger, anti-inflammatory, carbohydrate metabolism promoter, immunity system modulater.

Fig-9, 10

[18-20]

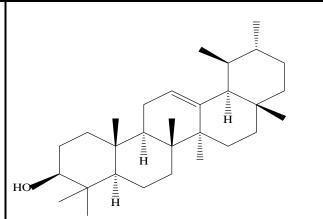
Compound: Acacetin (Flavone)

M.F.-C₁₆H₁₂ O₅

m.p. 268-272°CPale-yellow needles

 $[]D22-13.9^{\circ} (c = 0.69, CHCl3)$

Biological activity: **Inhibits** Human Atrial Repolarization Potassium Currents, Antioxidant, radical scavenger, anti-inflammatory, carbohydrate metabolism promoter, immunomodulater.



[17]

Compound: Amyrin, alpha (Triterpine)

M.F.-C₃₀H₅₀O

m.p- 188°C

White crystalline powder

Biological activity: Anti-elastase activity, modulates the membrane fluidity PGE2 release inhibition, strong anti-inflammatory activity, PKA inhibitor as well as a selective protease inhibitor.

Fig-11, 12

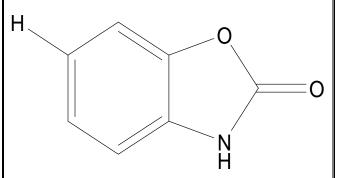
[16, 18]

Compound: Benzoxazin-3-one, 1-4: 2(h): 2-hydroxy (Nitrogen heterocy)

M.F.-C₈H₇NO₂

m.p.- 172-176 °C

Biological activity: Antimicrobial, anticancer and antiinflammatory.



[16, 18]

Compound: Benzoxazolinone (Nitrogen heterocy)

M.F.-C₇H₅NO₂

m.p.- 82-86°C

Light brown-pink Crystalline powder

Biological activity: Adrenergic and antihypertensive

properties.

Fig-13, 14

[21-23] Compound: Betulinic Acid (Triterpene)

 $M.F.-C_{30}H_{48}O_3$

m.p.- 295 - 298 °C (decomposes)

White crystalline powder

Optical Rotation: $+7^{\circ} - +9^{\circ}$ (c=0.9 in pyridine)

Biological activity: Potential anti-melanoma agent, Effective as a non-steroidal anti-inflammatory agent, anti-malarial, anti-HIV, prostaglandin antagonist.

H_3C

[21-22] Compound:Benzoxazolin-2-one, 6-methoxy (Nitrogen heterocy):

M.F.-C8H7NO3

m.p.-151-156 °C(lit.)

light tancolour

Biological activity: Antimicrobial and anti-inflammatory.

Fig-15, 16

[26-28]

Compound: Cirsimarin (Flavone)

M.F.-<u>C₂₃H₂₄O₁₁</u> m.p. - 244-246 °C

Biological activity: Stimulate lipolysis, antiinflammatory, anti-diabetic and hypotensive.

[25]

Compound: Benzoxazolone, 2(3H) 6-methoxy (Nitrogen heterocy):

 $M.F.-\frac{C_8H_7NO_3}{}$

m.p.- 152-156 °C

Biological activity: Antimicrobial, analgesicand antiinflammatory. Fig-17, 18

OH OH OH OH

[28]

Compound: Cirsitakaoside (Flavone)

 $M.F.-C_{23}H_{24}O_{11}$ m.p. - 246-247°C

Biological activity: Respiratory disease, gastric, hepaticdisturbances, anti-inflammatory, anti-diabetic and hypotension.

[29, 30]

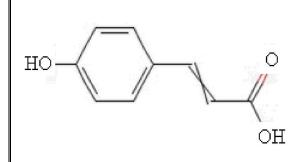
Compound: Cynaroside Flavone)

 $\begin{array}{l} \text{M.F.-C}_{21} \text{H}_{20} \text{O}_{11} \\ \text{m.p. - 266-268 °C} \end{array}$

Yellow amorphous powder

Biological activity: Antioxidant, anti-diabetic.

Fig-19, 20



OH OH

HO

OH

OH

OH

OH

OH

OH

OH

[31]

Compound: Coumaric Acid, para (Phenylpropanoid)

M.F.- $\frac{C_9H_8O_3}{210}$

m.p.-210-213 °C

Biological activity: Inhibits the development of stomach cancer.

[32]

Compound: Dulcitol (Diterpene)

MF:C₆H₁₄O₆ m.p.-188-189 °C

Biological activity: Antiviral and cytotoxic activity.

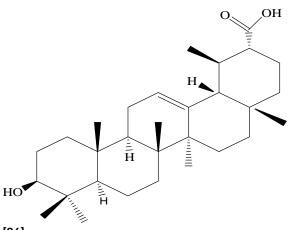
Fig-21, 22

[33]

Compound: Daucosterol(Steroid)

MF: C₃₅H₆₀O₆ m.p.- 295 °C

Biological activity: Immunomodulator



[26]

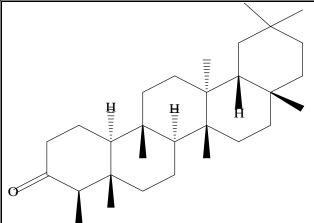
Compound: Dulcioic Acid (Triterpene)

MF: C30H48O3 m.p.-300 °C

Biological activity: Significant inhibitory effect on

cytokine production, antispasmodic.

Fig-23, 24



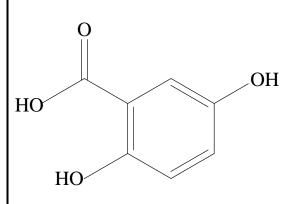
[35]

Compound: Friedelin (Triterpene)

M.F.-C30H50O m.p.-262-265 °C

Biological activity: Estrogenic, Anti-inflammatory,

analgesic and antipyretic.



[34]

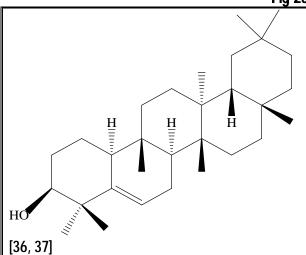
Compound: Gentisic acid (Benzenoid)

M.F.-C7H6O4 m.p.-200 - 205 °C

white to yellow powder

Biological activity: Antispasmodic, local anesthetic, antioxidant and anticonvulsant.

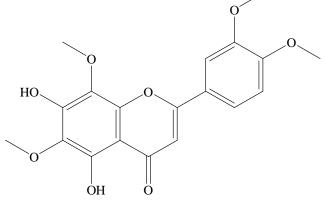




Compound: Glutinol (Triterpene)

M.F.-C30H50O m.p.- 206-208°C

Biological activity: Anti-inflammatory, analgesic.



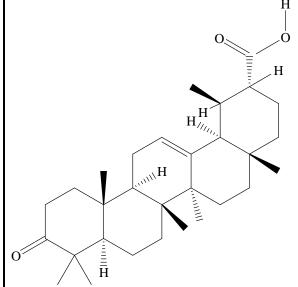
]38, 39]

Compound: Hymenoxin (Flavone)

M.F.-C19H18O8 m.p.-215-. 216°C

Biological activity: Estrogenic, antispasmodic.

Fig-27, 28



[40-41]

Compound: Ifflaionic Acid (Triterpene)

M.F.-C30H46O3 m.p.-303°C

Biological activity: Hypotensive

HO HOMINING OH HOM

[42]

Compound: Linarin (Flavone)

M.F.-C28H32O14 m.p.- 258-260°C

Biological activity: Sedative and sleep-enhancing properties.

Fig-29, 30

[43-46]

Compound: Luteolin (Flavone)

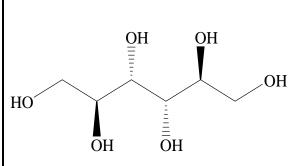
 $M.F.-C_{15}H_{10}O_6$ m.p.->330 °C

yellow crystalline compound

Biological activity:

Anti-oxidant, anti-cancer,

immunomodulator, anti-inflammatory.



[47, 48]

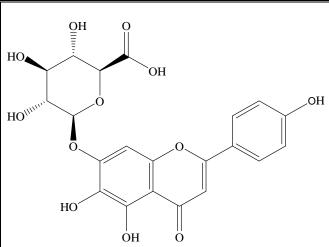
Compound: Mannitol, d (Carbohydrate)

M.F.- C₆H₁₄O₆ m.p.- 164 - 169 °C white, crystalline[

Biological activity: Diuretic, of Alzheimer's disease,

chemotherapy for brain tumors.

Fig-31, 32



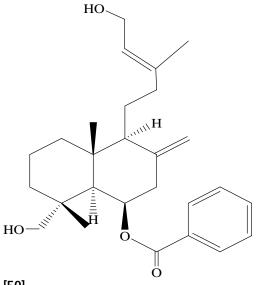
[49, 50]

Compound: Scutellarein (Flavone)

 $M.F.-C_{21}H_{18}O_{12}$ m.p.-218-220" C

Biological activity: Induce apoptosis of ovarian and

breast tumor cells in vitro.



Compound: Scoparinol (Diterpene)

M.F.- C₂₇H₃₈O₄

m.p.-

Biological activity: Anti-inflammatory, analgesic

Fig-33, 34

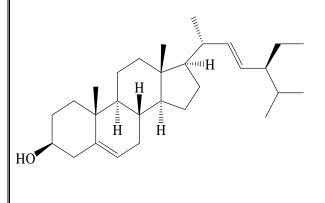
[51]

Compound: Sitosterol, beta (Steroid)

M.F.-C₂₉H₅O m.p.- 136-140 °C

Biological activity: Antioxidant, anti-cancer, anti-

tumor, reduce blood cholesterol levels.



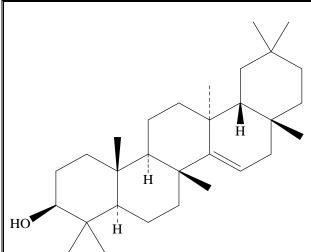
[52, 53]

Compound: Stigmasterol (Steroid)

M.F.- C₂₉H₄₈O m.p.- 161-170 °C

Biological activity: Anti-cancer, lower serum cholesterol, antioxidant, hypoglycemic.

Fig-35, 36

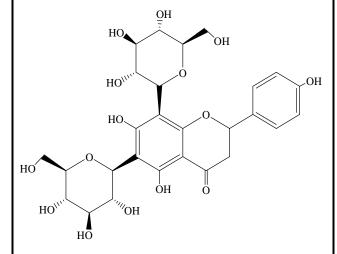


[54]

Compound: Taraxerol (Steroid)

M.F.-C30H50O m.p.- 282-285° C

Biological activity: Anti-cancer, anti-tumor

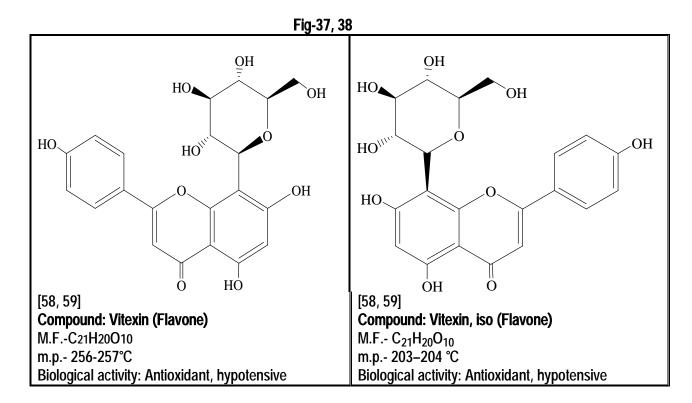


[55- 57]

Compound: Vicenin 2 (Flavone)

M.F.- C₂₇H₃₀O₁₅ m.p.-271-272

Biological activity: Anti-cancer, anti-inflammatory



Pharmacological Activity

The use of whole herb of S. dulcis in painful conditions acting both centrally and peripherally is well documented. It was found that the observed analgesia in S. dulcis was demonstrated by the active constituents, Glutionl, a triterpene [60-61] and Scoparinol, a diterpene [62] isolated from the plant extract through a peripherally acting mechanism similar to thenon-steroidal anti-inflammatory agents, such as indomethacin and diclofenac sodium.

The possible antioxidant property of aqueous extract of S. dulcis was tested in rats exposed to cadmium. Different group of animals were treated with CdCl2 alone or in combination with graded levels of S. dulcis (i.e. 250, 500 and 1000 mg/kg body wt, respectively). The results show that relative to controls, cadmium significantly reduced superoxide dismutase activity while significantly increasing catalase activity and malondialdehyde levels in the liver and kidney.

Another study summarizes the effect of S. dulcis on the population of immune cells during a 28 day experimental T. brucei infection in rabbits. The result

obtained showed that infection resulted in an initial rise in both total white blood cells (WBC) and the absolute number of circulating lymphocytes followed by a progressive decrease in total WBC and all WBC subtypes namely; lymphocytes, monocytes and granulocytes, although the % lymphocytes (lymphocytes expressed as % of total WBC) remained consistently higher than normal throughout the study period. Treatment with S. dulcis at a daily oral dose of 25 mg/Kg body weight significantly reduced the severity of the observed lesions (p < 0.05) when compared with untreated infected animals. Thus the herb demonstrates significant potency in protecting against the parasite induced decrease in the population of immunologically active cells.

The antioxidant efficacy of S. dulcis in STZ diabetic rats was compared with Glibenclamide. A significant increase in the activities of plasma insulin, superoxide dismutase, catalase, glutathione peroxidase, glutathione-S-transferase and reduced glutathione was observed in brain on treatment with 200 mg/kg body weight of S. dulcis plant aqueous extract and glibenclamide for 6 weeks. Both the treated groups

showed significant decrease in thiobarbituric acid reactive substances (TBARS) and hydroperoxides formation in brain, suggesting its role in protection against lipid peroxidation induced membrane damage [62]. It may be concluded that in diabetes, brain tissue was more vulnerable to oxidative stress and showed increased lipid peroxidation. The above observation shows that the aqueous extract of S. dulcis plant possesses antioxidant activity, which could exert a beneficial action against pathological alterations caused by the presence of free radicals in STZ diabetes.

Scoparia dulcis was investigated for anti-HSV-2 activity by plaque reduction assay. It was found that water extract of S. dulcis was active against HSV-2 with 50% effective dose of 1,190.4 μ g/ml and ED50 of ethanol extract of S. dulcis was 13.8 μ g/ml. Ethanol extract of S. dulcis showed highest Therapeutic Index (TI) (2.9) against HSV-2G.

The cytochrome P450 protective activity of the aqueous extract of S. dulcis was evaluated against CCl4 induced prolongation of pentobarbitone sleep time in Sprague-Dawley rats. The results indicate that, the aqueous extract of S. dulcis, at an oral dose of 0.5 g/kg, p.o., shows a significant protective effect against CCl4 induced cytochrome P450 damage and also show a significant intrinsic cytochrome P450 inhibition activity.

Hyperlipidemic effect of oral administration of the herb, S. dulcis, on T. brucei induced changes in plasma lipid profile in rabbits over a period of twenty eight days. Results obtained show that infection with T. brucei resulted in significant increases in plasma total cholesterol, trriacylglycerol, and low density lipoprotein (LDL)-cholesterol, while the level of high density lipoprotein (HDL)-cholesterol was also significantly reduced. Further comparative analysis of data revealed that these lesions were significantly less severe (p<0.05), in the infected and treated group relative to their untreated counterparts. The ability of S. dulcis to mitigate against these plasma lipid anomalies is underscored in the present study The level of total cholesterol, LDL cholesterol and triacylglycerol in

treated animals were significantly lower (p<0.05) relative to the infected but untreated group. Furthermore, the parasite induced decrease in HDL cholesterol was also significantly resisted in the treated group, thus enhancing the HDL: total cholesterol and the HDL: LDL ratios. This phenomenon no doubt favours a reduction in cardiovascular risk.

A group of experiments were performed on normal and experimental male Wistar rats treated with S. dulcis plant extract. The effect of extract was tested on streptozotocin (STZ) treated Rat insulinoma cell lines (RINm5Fcells) and isolated islets in vitro.

The extract markedly reduced the STZ-induced lipid peroxidation in RINm5F cells. Further, extract protected STZ-mediated cytotoxicity and nitric oxide (NO) production in RINm5F cells. Treatment of RINm5F cells with 5mMSTZ and 10g of extract completely abrogated apoptosis induced by STZ, suggesting the involvement of oxidative stress. Flow cytometric assessment on the level of intracellular peroxides using fluorescent probe 2'7'dichlorofluorescein diacetate (DCF-DA) confirmed that STZ (46%) induced an intracellular oxidative stress in RINm5F cells, which was suppressed by extract (21%). In addition, extract also reduced (33%) the STZ-induced apoptosis (72%) in RINm5F cells indicating the mode of protection of extract on RIN islets, and pan-creatic cell mass m5Fcells, (histopathological observations). Present study thus confirms antihyperglycemic effect of extract and also demonstrated the consistently strong antioxidant properties of S. dulcis used in the traditional medicine [63-65]

Much of the recent research on S. dulcis has centered around one powerful phytochemical called scopadulcic acid B (SDB). In a 1993 clinical study, SDB inhibited the growth of tumors in a test tube and in mice. The potency of SDB proved to be stronger than that of other natural antitumor-promoting terpenoids, such as glycyrrhetinic acid. [66].One of the chemical constituent is an aphidicolin-like tetracyclic diterpene named scopadulciol (SDC), which was isolated from S.

dulcis. SDC showed stimulatory effect on antiviral potency of acyclovir (ACV) or ganciclovir (GCV).

The effect of S. dulcis on T. brucei induced anaemia was investigated on rabbits. Changes in Packed cell volume (PCV), Haemoglobin (Hb) concentration, Red blood cell count (RBC), Mean cell haemoglobin (MCH), Mean cell haemoglobin concentration, (MCHC) and Mean cell volume (MCV) were monitored. The results obtained indicate that infection with T. brucei results in a significant decrease in PCV, Hb concentration and RBC. No significant changes were observed in MCH, MCHC and MCV. However the severity of observed anaemia was significantly less pronounced in the infected rabbits that were treated with S. dulcis when compared with their infected but untreated counterparts. It was concluded that S. dulcis therapy may prove useful in the management of T. brucei anaemia, and possibly other forms of anaemia. The herb may possess a measure of trypanocidal activity or immuno-stimulating properties that help to put the parasite in check and thus also control the deleterious effect of uncontrolled parasite proliferation. The plant has also been used in the management of sickle cell anaemia from decades (Hilda Ogbe, personal communication).

Fruit Juice, Seed Extract and leaf extract of S. dulcis was used for the mineralization of calcium oxalate, calcium carbonate and calcium phosphate. Four experimental models namely 'simultaneous flow static model' (S.S.M.), 'simultaneous flow dynamic model' (S.D.M.), 'reservoir static model' (R.S.M.) and 'reservoir dynamic model' (R.D.M.) were used for the study. The study suggests that the increased intake of fruits juice and seed extract of Scoparia dulcis would be helpful in urinary stone prophylaxis.

Conclusion

From this review we can conclude that studies with new active principles obtained from the whole plant of Scopariadulcis can results in novel and effective pattern of treatment. Chemical substances derived from this plant have been used to treat human diseases since the dawn of medicine. This plant may provide leads to find therapeutically useful compounds. Thus more efforts should be made towards isolation and characterization of the active principles and their structure activity relationship. The combination of traditional and modern knowledge can produce better drugs for the treatment of various ailments with fewer side effects.

References

- Branch LC, IMF da Silva. Folk medicine of Alter does chao, para. Branza Acta Amazonica. 1983;(13):737-767.
- 2. Satyanarayana K. Chemical examination of Scoparia dulcis (Linn.) Part-I.J. Indian Chem. Soc. 1969;(46):765-766.
- 3. Edeoga HO, Okwu DE and Mbaebie BO. Phytochemical constituents of some Nigerian medicinal Plants. African Journal of Biotechnology July 2005;Vol. 4(7):pp. 685-688.
- Chow SY, S.M.Chen, C.M. Yang. H.Hsu. Pharmacological studies on Chinese herbs. 1. Hypotensive effect of 30 Chinese herbs. Taiwan Yi Xue Za Zhi. 1974;(73):729-739.
- Hayashi K, Toshimitsu H, Naokata M. Cytotoxic and antitumor activity of scopadulcic acid from Scoparia dulcis L. Phytother Res 1992;6:6-9.
- Asano S, Mizutani M, Hayashi T, Morita N, Takeguchi N. Reversible inhibitions ofgastric H+ K+ ATPase by scopadulcic acid B and diacetyl scopadol: Newbiochemical tools of H+ K+ ATPase. J Biol Chem 1990;265:22167-22173.
- Hayashi K, Jung BL, Yoshie M, Naoki T, Hideo N, Toshimitsu Hayashi. The role ofa HSV thymidine kinase stmulating substance scopadulciol in improving the efficacyof cancer gene therapy. J Gene Med 2006;8:1056-1062.
- The Wealth of India. A Dictionary of Indian Medicinal and Industrial Product. First supp. Series (Raw materials) 5V, National Institute of science communication and information resources, CSIR, New Delhi. 2004; p. 87.
- Ayurvedic Pharmacopoeia of India, 2001. Indian system of Medicine & Homeopathy, Govt. of India Ministry of Health and Family Welfare, The

- Controller of Publication Civil Lines, Delhi, I ed. 2001;V –III: p.234.
- Branch LC, daSilva IMF. Folk medicine of Alter do chao, Para, Brazil, Acta Amazonica, 1983;13(5/6):737-797.
- 11. Denis P. Herbal Medicine among the Miskito of Eastern Nicaragua. Econ.Bot. 1988;42(1):16-28.
- Hayashi T., Kishi M., Kawasaki M., Arisawa M., Shimizu M., Suzuki S., Yoshizaki M., Morita N, Tezuka Y, Kikuchi T, Berganza LH, Ferro E, Basualdo I. Scopadulcic acid-A and -B, new diterpenoids with a novel skeleton, from a Paraguayan crude drug "typychá kurat " (Scoparia dulcis L.). Tetrahedron Lett. 1987;(28):3693–3696.
- Hayashi T, Okamura K, Kakemi M, Asano S, Mizutani M, Takeguchi N, Kawasaki M, Tezuka Y, Kikuchi T, Morita N, Chem. Pharm. Bull. 1990;38(10):2740–2745.
- 14. Kawasaki M, Hayashi T, Arisawa M, Shimizu M, Horie S, Ueno H, Syogawa H, Suzuki S, Yoshizaki M, Morita N, Tezuka Y, Kikuchi T, Berganza LH, Ferro E, Basualdo I, Structure of scoparic acid A, a new labdane-type diterpenoid from a paraguayan crude drug "typycha kurata" (Scoparia dulcis I.). Chem. Pharm. Bull. 1987;(35)3963–3966.
- Hayashi T, Kawasaki M, Okamura K, Tamada Y, Morita N, Tezuka Y, Kikuchi T, Miwa Y, Taga T, Scoparic acid A, a beta-glucuronidase inhibitor from Scoparia dulcis. J. Nat. Prod., 1992;(55)1748–1755.
- Hee-Kap Kang, Diane Ecklund, Michael Liu,Syamal K Datta http://arthritisresearch.com/content/11/2/R59. 09/02/12
- 17. **naturactiva.net**/articles/benefits_shea_butter.doc . 09/02/12
- 18. http://www.alibaba.com/showroom/acacetin.ht ml. 09/02/12
- 19. Hashimoto N, Yamashita T, Tsuruzoe N. Tertiapin, a selective IKACh blocker, terminates atrial fibrillation with selective atrial effective refractory period prolongation. Pharmacol Res. 2006;(54):136 –141.
- 20. http://circ.ahajournals.org/cgi/content/full/CIRC ULATIONAHA.108.769554/DC1

- Faujan NH, Alitheen NB, Yeap SK, Ali AM, Muhajir AH, Ahmad FBH. Cytotoxic effect of betulinic acid and betulinic acid acetate isolated from Melaleuca cajuput on human myeloid leukemia (HL-60) cell line. African Journal of Biotechnology. 2010;9(38):6387-6396.
- 22. Perumal Yogeeswari, Dharmarajan Sriram. Betulinic Acid and Its Derivatives: A Review on their Biological Properties . Current Medicinal Chemistry. 2005;(12):657-666.
- Simone Fulda. Betulinic Acid for Cancer Treatment and Prevention. .Int. J. Mol. Sci. 2008;(9):1096-1107.
- 24. Zikmundova M., Drandarov K., Bigler L., Hesse M., Werner C. Biotransformation of 2-Benzoxazolinone and 2-Hydroxy-1,4-Benzoxazin-3-one by Endophytic Fungi Isolated from Aphelandra tetragona. Applied And Environmental Microbiology. 2002;68(10):4863–4870.
- 25. http://www.znaturforsch.com/ac/v59c/s59c017 7.pdf. 09/02/12
- Girotti C., Ginet M., Demarne F.C., Lagarde M., Géloën A. Lipolytic activity of cirsimarin extracted from Microtea debilis.Planta Med. 2005;71(12):1170-2.
- 27. www.chemblink.com/products/13020-19-4.htm. 09/02/12
- 28. Saikat Sen, Raja Chakraborty, Biplab De, Joydeep Mazumder. Plants and phytochemicals for peptic ulcer: An overview. Phcog Rev. 2009;3:(6):270-279.
- Park JC, Park JG, Kim HJ, Hur JM, Lee JH, Sung NJ, Chung SK, Choi JW. Effects of extract from Angelica keiskei and its component, cynaroside, on the hepatic bromobenzenemetabolizing enzyme system in rats. Phytother Res. 2002;16 1:S24-7.
- Xiao Sun, Gui-boSun, MinWang,Jing Xiao,Xiao-bo Sun. Protective effects of cynaroside against H2O2-induced apoptosis in H9c2 cardiomyoblasts. Journal of Cellular Biochemistry. 2011;112(8)2019–2029.

- 31. Ferguson LR, Shuo-tun Z, Harris PJ. Antioxidant and antigenotoxic effects of plant cell wall hydroxycinnamic acids in cultured HT-29. Molecular Nutrition & Food Research. 2005;49(6)585–693.
- 32. www.chemblink.com/products/608-66-2.htm. 09/02/12
- Lee JH, Lee JY, Park JH, Jung HS, Kim JS, Kang SS, Kim YS, Han Y. Immunoregulatory activity by daucosterol, a beta-sitosterol glycoside, induces protective Th1 immune response against disseminated Candidiasis in mice. Vaccine, 2007;25(19):3834-40.
- 34. Moore WE, Bope FW, Christensens BV. Some new derivatives of gentisic acid. II. Pharmacological tests upon some new derivatives of gentisic acid. Journal of the American Pharmaceutical Association. 1954:6(43):334–337.
- 35. www.chemblink.com/products/559-74-0.htm_ 09/02/12
- www.molecularnetworks.com/biopath3/biopath/mols/Glutinol. 09/02/12
- Kamaliah Mahmood, Hapipah Mohd Ali, Rohana Yusof, Hamid Hadi A, Mary Pais. Chemical Components from the Light Petroleum Soluble Fraction of Uvaria cordata (Dunal) Alston. Pertanika J. Sci. & Techno. 1995;3(2):197-202.
- 38. Watson WH, Kashyap RP, Gao F, Mabry TJ. Structure of the flavone hymenoxin. ActaCrystallogr C. 1991;15(47)(Pt 2):459-61.
- 39. Atta-ur- Rahman, Studies in Natural Products Chemistry, Bioactive Natural Products, 1st Edn. Elsevier. 2005;Volume 32:pp.1127-1201.
- 40. www.tradingchem.com/lfflaionic_acid/6805-19-2.html. 09/02/12
- 41. Atta-ur- Rahman, Studies in Natural Products Chemistry, Bioactive Natural Products (Part B), Part 2, 1st Edn. Elsevier. 2000;pp.690-693.
- Sebastián Fernández, Cristina Wasowski, Alejandro C Paladini, Mariel Marder. Sedative and sleep-enhancing properties of linarin, a flavonoid-

- isolated from Valeriana officinalis. Pharmacology Biochemistry and Behavior. 2004;77(2):399-404.
- Chowdhury AR, Sharma S, Mandal S, Goswami A, Mukhopadhyay S, Majumder HK. Luteolin, an emerging anti-cancer flavonoid, poisons eukaryotic DNA topoisomerase Biochem. J. 2002;366:653-661.
- 44. en.wikipedia.org/wiki/Luteolin. 09/02/12
- 45. Kelley KW, Johnson RW. Luteolin reduces IL-6 production in microglia by inhibiting JNK phosphorylation and activation of AP-1. Proc. Natl. Acad. Sci. U.S.A. 2008;105(21):7534–9.
- 46. Mann John., Secondary Metabolism. 2nd Edn, Oxford, UK: Oxford University Press. 1992;pp.279–280.
- 47. Ikeda M, Bhattacharjee AK, Kondoh T, Nagashima T, Tamaki N. Synergistic Effect of Cold Mannitol and Na⁺/Ca²⁺ Exchange Blocker on Blood–Brain Barrier Opening Biochem. Biophys. Res. Commun. 2002;291:669-674.
- 48. http://en.wikipedia.org/wiki/Mannitol. 09/02/12
- http://www.chemicalbook.com/Search_EN.asp x?keyword=Scutellarein. 09/02/12
- 50. www.chemfaces.com/manual/Scoparinol-CFN99397.pdf. 09/02/12
- 51. Bouic PJ. The Role of Phytosterols and Phytosterolins in Immune Modulation: A Review of the Past 10 Years. CurrOpinClinNutrMetabCare. 2001; 4:471-475.
- 52. www.selleckchem.com/products/Stigmasterol(Stigmasterin).html. 09/02/12
- 53. Panda S, Jafri M, Kar A and Meheta BK. Thyroid inhibitory, antiperoxidative and hypoglycemic effects of stigmasterol isolated from Butea monosperma. Fitoterapia. 2009;80(2):123-126.
- 54. http://www.drugfuture.com/chemdata/luteolin.h tml. 09/02/12
- 55. Nagaprashantha LD, Vatsyayan R, Singhal J, Fast S, Roby R, Awasthi S, Singhal SS. Anti-cancer effects of novel flavonoid vicenin-2 as a single agent and in synergistic combination with docetaxel in prostate cancer. BiochemPharmacol, 2011;82(9):1100-9.

- 56. http://pubchem.ncbi.nlm.nih.gov/summary/summary.cgi?sid=12381.09/02/12
- 57. http://www.jstage.jst.go.jp/article/cpb/51/10/51 __1204/_article/-char/en. 09/02/12.
- 58. Kim JH, Lee BC, Kim JH, Sim GS, Lee DH, Lee KE, Yun YP, Pyo HB. The isolation and antioxidative effects of vitexin from Acer palmatum. Arch Pharm Res. 2005;28(2):195-202.
- 59. en.wikipedia.org/wiki/Vitexin. 09/02/12
- Freire SMF, Torres LMB, Roque NF, Souccar C. Analgesic activity of a triterpene isolated from ScopariadulcisL. (Vassourinha). Memórias do InstitutoOswaldo Cruz. 1991;(86):149-151.
- FariasFreie SM, Silva Emin JA, Lapa AJ, Souccar C &Brandao Torres LM. Analgesic and antiinflammatory properties of Scoparia dulcis L. extract and glutinol in rodents. Phytotherapy Research. 1993;(7):408-414.
- 62. Ahmed M, Shikha HA, Sadhu SK, Rahman MT, Datta BK. Analgesic, diuretic, and anti-

- inflammatory principle from Scopariadulcis. Pharmazie, 2001;56(8):657-660
- Latha M, Pari L, Sitasawad S, Bhonde R. Insulin secretagogue activity and cytoprotective role of the traditional antidiabetic plant Scoparia dulcis ,Life Sci. 2004;75(16):2003-2014.
- 64. Pari L, Latha M. Antihypoglycaemic activity of Scopariadulcis: effect on key metabolic enzymes of carbohydrate metabolism in streptozotocininduced diabetes. Pharm. Biol. 2004;42(8):570-576.
- Pari L, Venkateswaran. Hypoglycaemic activity of Scopariadulcis L. extract in alloxan-induced hyperglycaemic rats, PPhytotherapy Research. 2002;16(7):662-664.
- Nishino H Anti-tumor promoting activity of Scopadulcic Acid B, Isolated from the medicinal plant Scoparia dulcis. L. Oncology. 1993;50(2):100-103.