

A study on the antibacterial effect of selected medicinal plants of Western Ghats against dental caries bacteria

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Abstract

This study was conducted to reveal the antibacterial effect of selected medicinal plants from Western Ghats against dental caries bacteria and aware the populace about the importance of using phytomedicines. In this study, the antibacterial effects of nine well known medicinal plants were checked against the dental caries bacteria using well diffusion method. The highest zonation was reported in acetone and methanol extracts. The Chloroform and ethanol extracts revealed sensible activity and water extracts were produced least inhibitory activity. Acetone extract of *Eucalyptus* gained high inhibitory activity against *Bacillus megaterium*. While *Pseudomonas aeruginosa* showed resistance to all the extracts of *Syzygium aromaticum*.

Keywords: Dental caries, Western Ghats, Medicinal plants, Antibacterial activity, Phytochemical investigation

Introduction

Dental decay is the most prevalent disease affecting humanity. Teeth get decayed due to a combination of causes that include bad oral hygiene, stagnation of food on or around the teeth, presence of plaque on the tooth structure and the presence of caries causing microorganisms [1]. The presence of a certain types of microorganism was discovered during the last decade in dental plaques. The nucleating role of the microorganisms in the formation of dental calculus shows similarities to that of nanobacteria in calcification [2]. Periodontal disease has long been recognized as a chronic disease, but literature describes it as a disease derived entirely from the effects of a microbial colonization of the gingival crevice. If this were

so, it would mean that periodontal disease is unique among chronic diseases, all of which represent the long-term cumulative effects of interaction between a host biologic system and the surrounding environment [3]. Antibiotic resistance is the ability of a microorganism to withstand the effects of an antibiotic. It is a specific type of drug resistance. Antibiotic resistance evolves naturally via natural selection through random mutation, but it could also be engineered by applying an evolutionary stress on a population. Once such a gene is generated, bacteria can then transfer the genetic information in a horizontal fashion (between individuals) by plasmid exchange. The patterns of antibiotic usage greatly affect the number of resistant organisms which develop. Overuse of broad-spectrum antibiotics, such as second- and third-generation greatly hastens the development of

resistance. Other factors contributing towards resistance include incorrect diagnosis, unnecessary prescriptions, improper use of antibiotics by patients [4]. Antibiotic resistance in microorganisms recovered from the acute dental abscess has been reported to be increasing (with the exception of Metronidazole) in some populations studied over the last few decades. The resistance problem demands that a renewed effort be made to seek antibacterial agents effective against pathogenic bacteria resistant to current antibiotics. One of the possible strategies towards this objective is the rational localization of bioactive phytochemicals. Plants have a limitless ability to synthesize aromatic substances, most of which are phenols or their oxygen substituted derivatives such as tannins. Many of the herbs and spices used by humans to season food yield have useful medicinal compounds including those having antibacterial activity [5]. Plant derived drugs remain an important resource especially in developing countries to combat serious diseases.

Materials and Methods

Selection of Bacterial Strains

Bacterial strains of six different species (*Pseudomonas aeruginosa*, *Streptococcus salivarius*, *Streptococcus viridans*, *Streptococcus mutans*, *Bacillus megaterium*, and *Neisseria catarrhalis*) were selected from Microbial Technology Laboratory, Malankara Catholic College, Mariagiri, Kaliakkivilai, Tamil Nadu.

Collection of Medicinal Plants

The medicinal plant samples were collected from the Maruthuvarmalai region of Western Ghats of Kanyakumari district. The different parts such as root, stem, leaves and inflorescence of nine well-known plants were selected for testing its antibacterial studies and characterization of secondary metabolites of effective ones.

Preparation of Plant Extracts

Plant samples were shade dried and ground well. 10 gram of powdered sample was filled in screw cap bottles with 10 ml of different solvent systems (acetone, ethanol, chloroform, methanol and water). It was kept at 22⁰ c for fifteen days.

Antibacterial Effect Checking of Medicinal Plant Extracts

Antibacterial effect of medicinal plant extracts were checked by Well- diffusion method.

Well Diffusion Method

The bacterial isolates were effectively swabbed on the Mueller-Hinton agar plates. After allowing the inoculums to dry at room temperature and six millimeter wells were bored on it. The extract was introduced (50 μ l of a 100mg/ml concentration) into three duplicate wells. The plates were allowed to stand at room temperature for one hour for the extract to diffuse into the agar and then they were incubated at 37⁰ c for 18 hours. After incubation the plates were observed for the results.

Results

Antimicrobial Effect of Medicinal Plant Extracts

The antimicrobial activity of nine selected medicinal plant's (*Syzygium aromaticum*, *Piper betle*, *Areca catechu*, *Camellia sinensis*, *Eucalyptus globules*, *Zingiber officinale*, *Gymnema sylvestre*, *Azadirachta indica*, and *Chrysopogon zizanioides*) ethanol, acetone, chloroform, methanol and water extracts against bacterial isolates (*Pseudomonas aeruginosa*, *Streptococcus salivarius*, *Streptococcus viridans*, *Streptococcus mutans*, *Bacillus megaterium*, *Neisseria catarrhalis*) were tabulated.

The acetone extract of *Eucalyptus globules* showed high inhibitory activity against *Streptococcus mutans*, *Streptococcus viridans* and *Bacillus megaterium* with 30mm, 31mm and 36mm respectively. Whereas the ethanol extract revealed activity against *Bacillus megaterium* and *Streptococcus salivarius*.. It was observed that

Table.1. Zone of Inhibition of Different Extracts of *Eucalyptus globulus* against Bacteria

Bacteria	Acetone (mm)	Chloroform (mm)	Ethanol (mm)	Methanol (mm)	Water (mm)
<i>Neisseria catarrhalis</i>	18	10	25	17	13
<i>Streptococcus mutans</i>	30	12	23	17	–
<i>Streptococcus salivarius</i>	18	10	25	17	–
<i>Streptococcus viridans</i>	31	10	–	22	27
<i>Bacillus megaterium</i>	36	13	26	20	14
<i>Pseudomonas aeruginosa</i>	11	9	13	–	–

Table.2. Zone of Inhibition of Different Extracts of *Zingiber officinale* against Bacteria

Bacteria	Acetone (mm)	Chloroform (mm)	Ethanol (mm)	Methanol (mm)	Water (mm)
<i>Neisseria catarrhalis</i>	25	16	23	17	–
<i>Streptococcus mutans</i>	21	15	20	17	–
<i>Streptococcus salivarius</i>	12	15	15	16	–
<i>Streptococcus viridans</i>	16	14	20	14	–
<i>Bacillus megaterium</i>	12	16	16	11	–
<i>Pseudomonas aeruginosa</i>	–	13	11	–	–

Table.3. Zone of Inhibition of Different Extracts of *Syzygium aromaticum* against Bacteria

Bacteria	Acetone (mm)	Chloroform (mm)	Ethanol (mm)	Methanol (mm)	Water (mm)
<i>Neisseria catarrhalis</i>	30	18	23	25	–
<i>Streptococcus mutans</i>	23	20	24	24	24
<i>Streptococcus salivarius</i>	22	15	21	20	11
<i>Streptococcus viridans</i>	21	18	22	18	11
<i>Bacillus megaterium</i>	22	17	18	20	12
<i>Pseudomonas aeruginosa</i>	–	–	–	–	–

Table.4. Zone of Inhibition of Different Extracts of *Camellia sinensis* against Bacteria

Bacteria	Acetone (mm)	Chloroform (mm)	Ethanol (mm)	Methanol (mm)	Water (mm)
<i>Neisseria catarrhalis</i>	25	–	15	18	–
<i>Streptococcus mutans</i>	20	–	–	–	–
<i>Streptococcus salivarius</i>	20	–	18	15	–
<i>Streptococcus viridans</i>	22	–	30	24	–
<i>Bacillus megaterium</i>	18	–	25	23	–
<i>Pseudomonas aeruginosa</i>	15	8	18	9	–

Table.5. Zone of Inhibition of Different extracts of *Piper betle* against Bacteria

Bacteria	Acetone (mm)	Chloroform (mm)	Ethanol (mm)	Methanol (mm)	Water (mm)
<i>Neisseria catarrhalis</i>	10	11	17	11	–
<i>Streptococcus mutans</i>	12	21	13	12	–
<i>Streptococcus salivarius</i>	11	9	18	9	7
<i>Streptococcus viridans</i>	10	23	15	20	–
<i>Bacillus megaterium</i>	22	15	23	20	–
<i>Pseudomonas aeruginosa</i>	18	20	14	18	–

Table.6. Zone of Inhibition of Different extracts of *Gymnema sylvestre* against Bacteria

Bacteria	Acetone (mm)	Chloroform (mm)	Ethanol (mm)	Methanol (mm)	Water (mm)
<i>Neisseria catarrhalis</i>	10	8	13	–	–
<i>Streptococcus mutans</i>	13	–	13	9	–
<i>Streptococcus salivarius</i>	10	10	13	18	–
<i>Streptococcus viridans</i>	13	–	15	11	13
<i>Bacillus megaterium</i>	20	14	14	20	–
<i>Pseudomonas aeruginosa</i>	16	–	10	–	–

Table.7. Zone of Inhibition of Different extracts of *Azadirachta indica* against Bacteria

Bacteria	Acetone (mm)	Chloroform (mm)	Ethanol (mm)	Methanol (mm)	Water (mm)
<i>Neisseria catarrhalis</i>	17	9	13	20	–
<i>Streptococcus mutans</i>	7	12	10	9	–
<i>Streptococcus salivarius</i>	15	14	11	11	–
<i>Streptococcus viridans</i>	12	9	15	10	–
<i>Bacillus megaterium</i>	15	9	8	–	7
<i>Pseudomonas aeruginosa</i>	–	6	6	–	–

Table.8. Zone of Inhibition of Different extracts of *Chrysopogon zizanioides* against Bacteria

Bacteria	Acetone (mm)	Chloroform (mm)	Ethanol (mm)	Methanol (mm)	Water (mm)
<i>Neisseria catarrhalis</i>	12	8	18	12	11
<i>Streptococcus mutans</i>	18	–	13	10	–
<i>Streptococcus salivarius</i>	15	–	–	11	–
<i>Streptococcus viridans</i>	32	–	18	18	–
<i>Bacillus megaterium</i>	25	10	25	25	7
<i>Pseudomonas aeruginosa</i>	12	8	13	9	–

Table.9. Diameter of Zone of Inhibition of Different extracts of *Areca catechu* against Bacteria

Bacteria	Acetone (mm)	Chloroform (mm)	Ethanol (mm)	Methanol (mm)	Water (mm)
<i>Neisseria catarrhalis</i>	20	11	11	18	–
<i>Streptococcus mutans</i>	18	9	12	13	–
<i>Streptococcus salivarius</i>	17	9	12	12	–
<i>Streptococcus viridans</i>	20	–	12	12	–
<i>Bacillus megaterium</i>	18	9	16	19	–
<i>Pseudomonas aeruginosa</i>	12	19	11	–	–

water extract inhibited the growth of *Streptococcus viridans* with a zone of 27mm.

The acetone extract of *Zingiber officinale* inhibited the growth of *Neisseria catarrhalis* and produced zone of 25mm. it was observed that the ethanol extract has the ability to inhibit the growth of *Neisseria catarrhalis* and *Streptococcus mutans*.

The chloroform extract of *Syzygium aromaticum* showed activity against both *Neisseria catarrhalis* and *Streptococcus viridans* with a clear zone of 18mm each. Whereas its acetone extraction revealed high activity against *Neisseria catarrhalis* with a zone of 30 mm. The methanol extract produced high activity against *Streptococcus mutans* with a zone of 24mm. It was observed that the water extract inhibited the growth of *Streptococcus mutans* with a clear zone of 24mm.

Neisseria catarrhalis was sensitive to acetone extract of *Camellia sinesis* with a growth inhibitory zone of 25mm. whereas the chloroform extract showed activity only against *Pseudomonas aeruginosa* with a zone of 8mm. Water extract did not showed activity on any organism.

The crude acetone extract of *Piper betle* showed high inhibitory activity against *Bacillus megaterium* and *Pseudomonas aeruginosa* with a zone of 22mm, 18mm respectively. Chloroform extract revealed activity against *Streptococcus viridans* with a clear zone of 23mm. Whereas ethanol extract showed inhibitory activity against *Bacillus megaterium* with zone of 23mm. it was observed that water extract has ability to inhibit *Streptococcus salivarius* with a zone of 7mm.

The acetone extract of *Gymnema sylvestre* showed highest activity against *Bacillus megaterium* and *Pseudomonas aeruginosa* with 20mm, 16mm respectively. Whereas the water extract showed the maximum activity only against *Streptococcus viridans* with a clear zone of 13mm.

Acetone extract of *Azadirachta indica* showed maximum inhibitory activity against *Neisseria catarrhalis* and *Streptococcus salivarius* with 17mm, 15mm respectively. Whereas the methanol extract showed high activity of 20mm zone against *Neisseria catarrhalis* and 11mm on *Streptococcus salivarius*. It was observed that water extract has activity against *Bacillus megaterium* alone.

Acetone extract of *Chrysopogon zizanioides* showed high activity against *Streptococcus viridans* (32mm) and ethanol extract expressed high activity against *Bacillus megaterium* (25mm). Whereas the methanol extract showed high inhibitory activity against *Bacillus megaterium* and *Streptococcus viridans* with 25mm, 18mm respectively.

The crude acetone extract of *Areca catechu* showed maximum inhibitory activity against *Neisseria catarrhalis* (20mm) and chloroform extract was highly inhibited *Pseudomonas aeruginosa* with a zone of 19mm. The methanol extract expressed high activity against *Bacillus megaterium* and *Neisseria catarrhalis* with 19mm, 18mm respectively.

Discussion

Nowadays the degree of dental caries and related problems are increasing with severe effects. As part of search for effective phytoderivatives against dental pathogens, nine medicinal plants (*Syzygium aromaticum*, *Piper betle*, *Areca catechu*, *Camellia sinensis*, *Eucalyptus globules*, *Zingiber officinale*, *Gymnema sylvestre*, *Azadirachta indica*, *Chrysopogon zizanioides*) were collected from the Maruthuvar Malai region of Western Ghats in Kanyakumari district and its solvent extracts were applied against bacterial isolate. The result evidenced that all the selected plants are with acceptable levels of bactericidal activity. The highest zonation was reported in acetone and methanol extracts. The Chloroform and ethanol extracts showed moderate activity and water extracts were showed relatively least inhibitory activity. Perhaps the variations may be due to the polarity of solvents which determines the type of reaction and solubility of compounds. The acetone and methanol have better extracting capacity which may be attributed to the ability to extract the natural antimicrobial compounds such as alkaloids, flavanoids, terpinoids and phenolic compounds from the plant. Acetone extract of *Eucalyptus* showed high inhibitory activity against *Bacillus megaterium*. Whereas *Pseudomonas aeruginosa* was resistant to all the extracts of *Syzygium aromaticum*. These results strengthen the possibility of application of phytoderivatives against human oral flora.

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References

- [1]. J. Kirkham, R. C. Shore, S. J. Brookes and C. 2002. Robinson. *FEMS Microbiology Ecology*, **39**, Issue 3, 239-244
- [2]. Turgut Demir. 1990. *Archives of Oral Biology*, **35**, Supplement 1, S177-S180

- [3]. V R Dowell, Jr, S Offenbacher, W Snyder, and T Hersh .2006. *Archives of Oral Biology*, **47**, Issue 6, 491-498
- [4]. Parekh J, Karathia N, Chanda S. 2006. Screening of some traditionally used medicinal plants for potential antibacterial activity. *Indian J Pharm Sci*; **68**:832-4
- [5]. Bhavnani, S.M. and Ballow, C.H. 2000. *Curr. Opin. Microbiol.*, **3**: 528